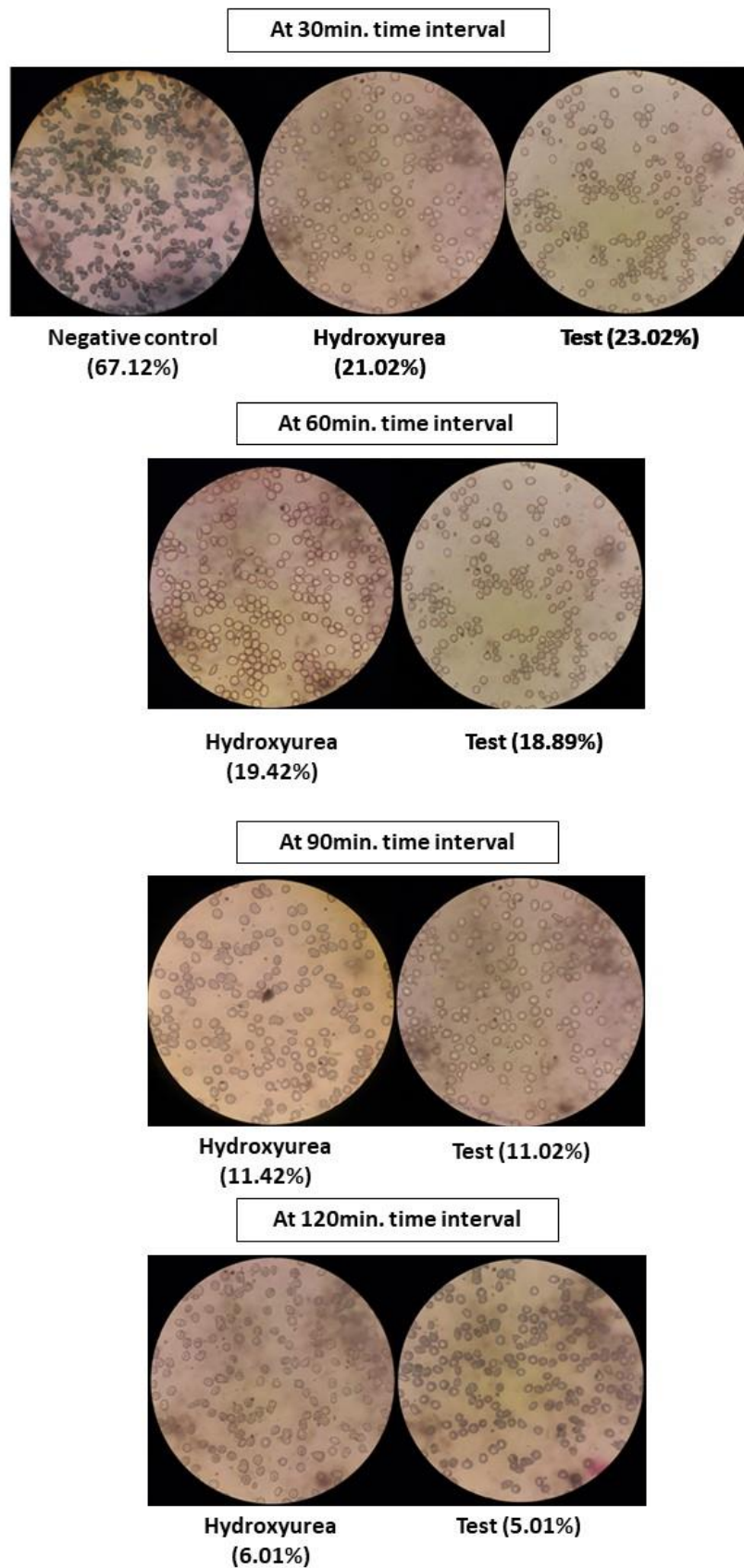


Supplementary Information

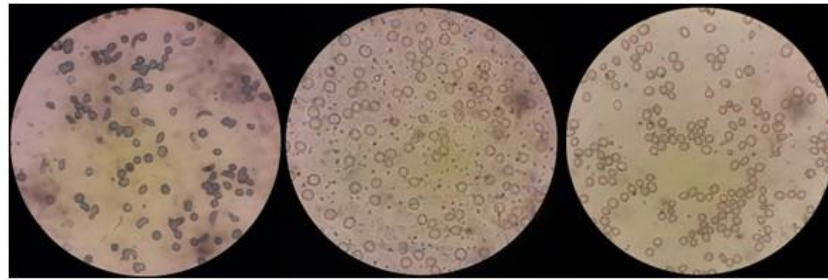
Figure S1.

A) Percentage sickling at 30% (v/v)



B) Percentage sickling at 60% (v/v)

At 30min. time interval

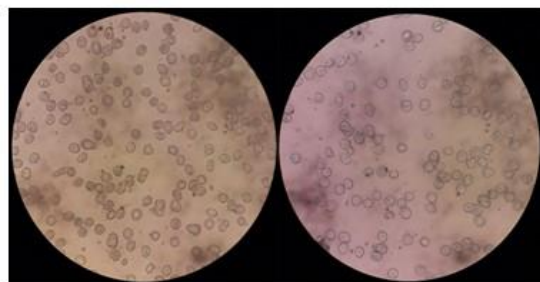


Negative control
(87.2%)

Hydroxyurea
(3.92%)

Test (3.02%)

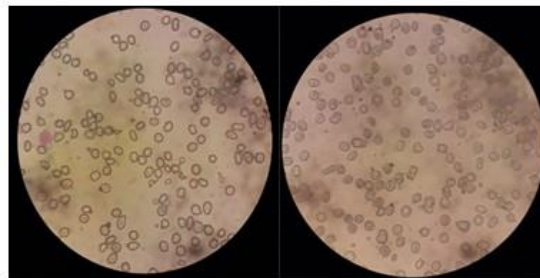
At 60min. time interval



Hydroxyurea
(4.42%)

Test (4.99%)

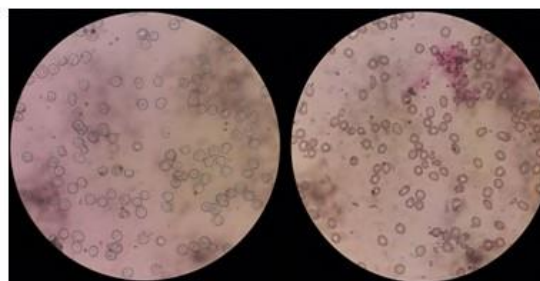
At 90min. time interval



Hydroxyurea
(2.92%)

Test (2.81%)

At 120min. time interval



Hydroxyurea
(0.98%)

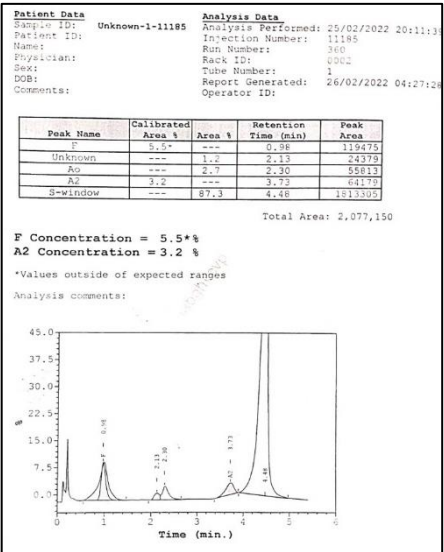
Test (1.91%)

Figure S1. Microscopic Observation (40X) of percentage sickling at different dose and time duration of *L. plantarum* (CS) and hydroxyurea.

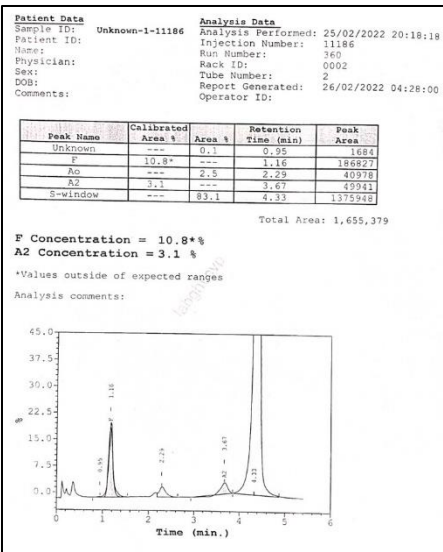
- A) Percentage sickling at 30% (v/v) culture supernatant and hydroxyurea at different time duration:** The percentage of sickling decreased in *L. plantarum* (CS) as well as hydroxyurea of 30%(v/v) at 30 min, 60 min, 90 min and 120 min of time intervals compared to the Negative control. The least percentage of sickling in *L. plantarum* (CS) was observed at 60 min which was comparable to Hydroxyurea (Drug Control).
- B) Percentage sickling at 60% (v/v) culture supernatant and hydroxyurea at different time duration:** The percentage of sickling decreased in *L. plantarum* (CS) as well as hydroxyurea of 60%(v/v) at 30 min, 60 min, 90 min and 120 min of time intervals compared to the Negative control. The least percentage of sickling in *L. plantarum* (CS) was observed at 60 min which was comparable to Hydroxyurea (Drug Control).

Figure S2.

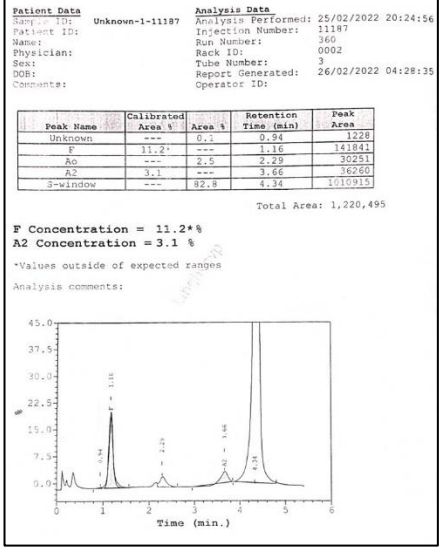
(A) Percentage increase in fetal hemoglobin at 60 min. interval



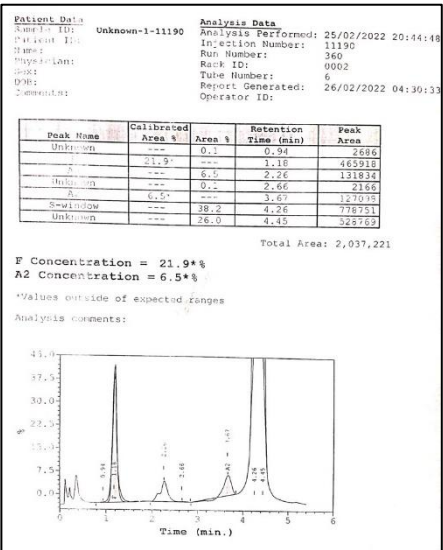
Negative



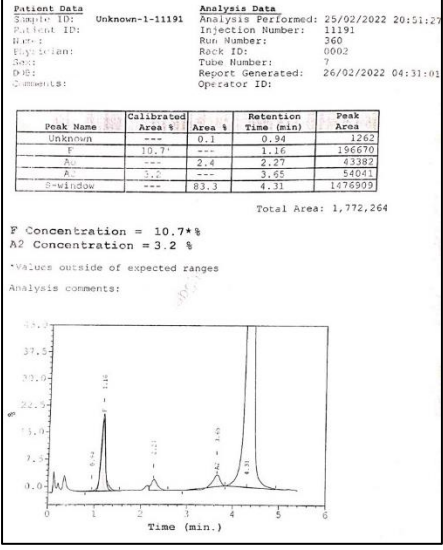
L. plantarum at 30% (v/v)



L. plantarum at 60% (v/v)

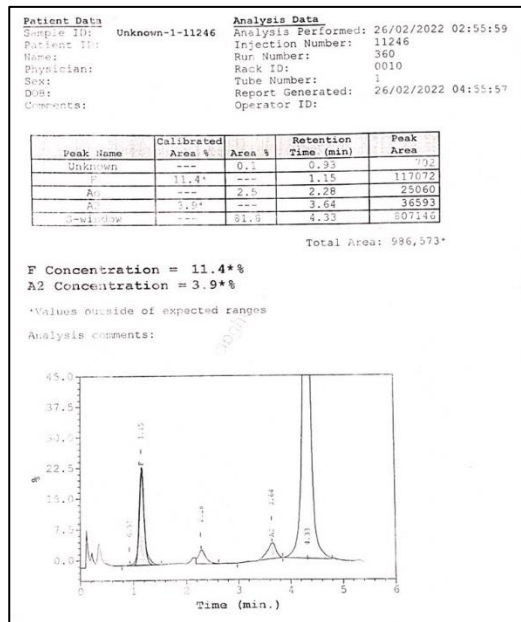


Hydroxyurea at 30% (v/v)

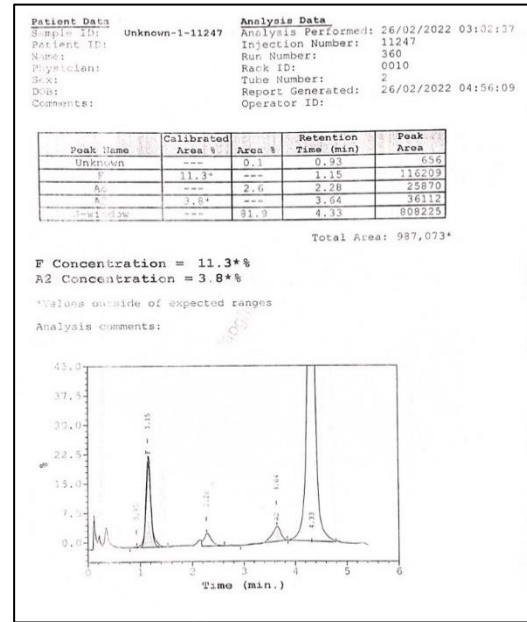


Hydroxyurea at 60% (v/v)

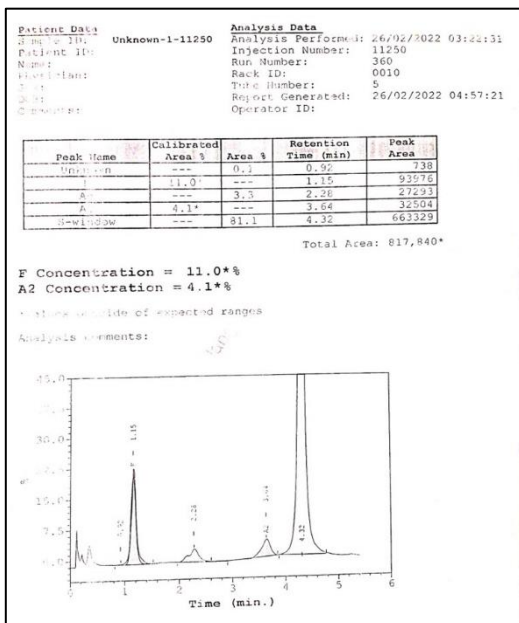
(B) Percentage increase in fetal hemoglobin at 120 min. interval



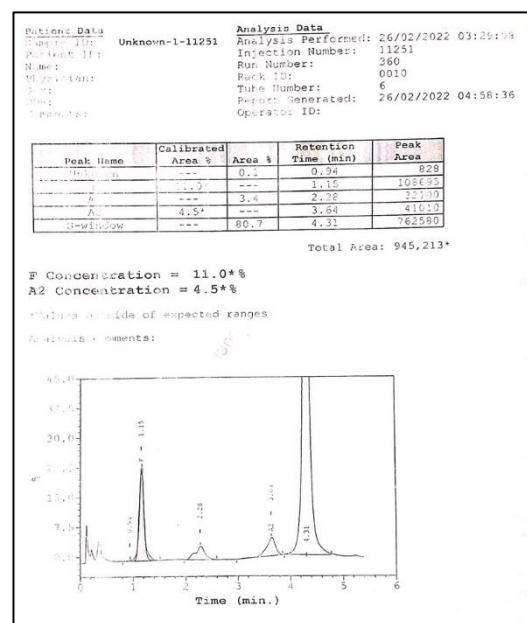
***L. plantarum* at 30% (v/v)**



***L. plantarum* at 60% (v/v)**



Hydroxyurea at 30% (v/v)



Hydroxyurea at 60% (v/v)

Figure S2. The representative HPLC reports for the percentage increase in fetal hemoglobin (HbF%) upon treatment with *L. plantarum* (CS) at different time intervals.

Percentage increase in fetal hemoglobin at 60 min. interval: The percentage of fetal haemoglobin (Fetal Hemoglobin Concentration denoted as F%) increased in *L. plantarum* (CS) as well as

hydroxyurea of 30% (v/v) & 60% (v/v) concentration at 60 min of time intervals compared to the Negative control.

Percentage increase in fetal hemoglobin at 120 min. interval: The percentage of fetal haemoglobin (Fetal Hemoglobin Concentration denoted as F%) increased in *L. plantarum* (CS) as well as hydroxyurea of 30% (v/v) & 60% (v/v) concentration at 120 min of time intervals compared to the Negative control.

Table S1. Percentage of sickling at 30% v/v & 60% v/v of *L. plantarum* (CS) and Hydroxyurea.

Concentration of <i>L. plantarum</i> (CS) and Hydroxyurea	Percentage Sickling at different time intervals (%) (Mean \pm SEM)			
	(N=15)			
30 mg/ml	30 min	60 min	90 min	120 min
Negative Control	56.80 \pm 6.250			
<i>L. plantarum</i> (CS)	9.066 \pm 1.740	7.753 \pm 7.703	7.717 \pm 2.617	7.133 \pm 2.212
Hydroxyurea	9.803 \pm 2.018	7.795 \pm 2.100	8.134 \pm 3.516	5.344 \pm 2.160
p* value	p<0.0001	p<0.0001	p<0.0001	p<0.0001
60 mg/ml				
<i>L. plantarum</i> (CS)	5.892 \pm 1.278	6.846 \pm 2.344	8.053 \pm 2.465	6.828 \pm 2.224
Hydroxyurea	8.910 \pm 2.917	5.819 \pm 1.436	6.4 \pm 2.694	4.741 \pm 1.170
p* value	p<0.0001	p<0.0001	p<0.0001	p<0.0001

*p values refer to comparison made between negative control (normal saline) and *L. plantarum* (CS) at different intervals.

Table S2. Percentage of Fetal Hemoglobin Concentration at 30% v/v & 60% v/v of *L. plantarum* (CS) and Hydroxyurea.

Concentration of <i>L. plantarum</i> (CS) and Hydroxyurea	Percentage of Fetal Hemoglobin Concentration at different time intervals (%)	
	(Mean \pm SEM) (N=10)	
30% (v/v)	60 min	120 min
Negative Control	9.438 \pm 1.368	
<i>L. plantarum</i> (CS)	22.98 \pm 5.301	23.83 \pm 5.771
Hydroxyurea	23.43 \pm 5.916	23.18 \pm 5.591
p* value	p<0.0001	p<0.0001
60% (v/v)		
Test Extract	26.40 \pm 4.150	23.18 \pm 5.591
Hydroxyurea	23.35 \pm 5.827	23.10 \pm 5.474
p* value	p<0.0001	p<0.0001

*p values refer to comparison made between negative control (normal saline) and *L. plantarum* (CS) at different intervals.

Table S3. Comparison of anti-sickling & HbF inducing activity of *L. plantarum* (CS) between erythrocytes of severe and less severe groups of SCD patients.

Treatment (30%v/v for 120 min.)	(A) Percentage Sickling (Mean ± SEM)		p= value	Significant Difference (p<0.05)
	Severe (N=6)	Less Severe (N=9)		
Negative Control (Normal Saline)	(65.32 ± 2.592)	(59.34 ± 1.317)	0.100 ^a	No
<i>L. plantarum</i> (CS)	(17.89± 3.890)	(15.47± 3.174)	0.635 ^b	No
Hydroxyurea	(21.33± 3.751)	(29.67± 3.851)	0.146 ^c	No
	(B) HbF Percentage (HbF%) (Mean ± SEM)			
Negative Control (Normal Saline)	(6.260± 0.647)	(8.838± 0.60)	0.019 ^a	Yes
<i>L. plantarum</i> (CS)	(17.42± 3.713)	(19.93± 4.797)	0.685 ^b	No
Hydroxyurea	(18.14± 3.488)	(21.15± 4.545)	0.608 ^c	No

‘N’ represents number of SCD Patients,

^a Severe patients vs. Less severe patients for Negative control,

^b Severe patients vs. Less severe patients for *L. plantarum* (CS),

^c Severe patients vs. Less severe patients for Hydroxyurea.

Genotyping of *rs334* (A/T; M34058) polymorphism for identification of homozygous Sickle cell disease patients

Genomic DNA was extracted from SCD patients' blood samples to confirm the HbSS genotype. 300µl of venous blood was used to isolate genomic DNA using the QIAamp DNA Blood Kit (Qiagen, Germany). Following extraction, DNA was kept for further examination at 20°C, and 0.8% agarose gel electrophoresis was used to determine its purity. The *rs334* (A/T) polymorphism of the β -globin gene was genotyped using allele-specific-polymerase chain reaction (AS-PCR). (GenBank accession no. M34058), as previously described.²⁵ The following wild-type primer set was utilized for amplification of 517 bp fragment: WT-AS [5'-ATG GTG CAC CTG ACT CCT GA-3'] and WT-CP517 [5'-CCC CTT CCT ATG ACA TGA ACT-3']; and for 267 bp fragment amplification (SCD) following mutant primer set was used: MUT-AS [5'-CAG TAA CGG CAG ACT TCT CCA-3'] and MUT-CP267 [5'-GGG TTT GAA GTC CAA CTC CTA-3']. By using 2.0% agarose gel electrophoresis, the three genotypes of the *rs334* (A/T) polymorphism were determined: AA homozygous (Healthy individual), AT heterozygous (Carrier), and TT homozygous (SCD patient).

The 25 µl of the PCR reaction mixture contained 15 µl nuclease free water, 3 µl (50 ng) genomic DNA, 2.5 µl 2.5 mM dNTPs (Takara, Japan), 0.4 µL (2.5 U/µL) Taq Polymerase (Takara, Japan), 2.5 µl 10X PCR buffer and 1 µl of 10 µM corresponding forward and reverse primers (Eurofins, Luxembourg). The following protocol was used for performing the amplification using the Bio-Rad T100 Thermal Cycler: 95°C for two minutes (initial denaturation), 30 cycles of 95°C for 30 sec. (denaturation), 60°C for 30 sec. (primer annealing), and 72°C for 35 sec. (extension). All amplified products were evaluated by 2.0% agarose gel electrophoresis.