

Functional Characterization of the Tmg-4 Magnesium Transporter with the GQN Motif from *Neurospora crassa*

Sireesha Radarapu¹, Neelima Gorityala² , Uday Kumar Banala³ ,
Someswar Rao Sagurthi²  and Premeagar Korripally^{1*} 

¹Department of Biotechnology, UCS, Mahatma Gandhi University, Nalgonda, Telangana, India.

²Department of Genetics and Biotechnology, Osmania University, Hyderabad, Telangana, India.

³Radiological and Environmental Safety Division, Indira Gandhi Centre for Atomic Research, Kalpakkam, Tamil Nadu, India.

Abstract

To unveil the physiological relevance of magnesium and its transport pathways in *Neurospora crassa*, the vegetative, asexual, and sexual phases of development were investigated. Notably, a regular rate of hyphal growth extension was observed in media without magnesium supplementation. Further, conidia and perithecia formation was completely abolished under the same conditions. By estimating the levels of mycelial cations, magnesium was identified as the 3rd most abundant ion and its transport was found to be mediated by four putative CorA magnesium transporters: Tmg-1, Tmg-2, Tmg-3, and Tmg-4. Among these, the Tmg-4 transporter encoded by the NCU07816.5 (*tmg-4*) gene possesses a GQN motif instead of the universally conserved GMN motif of CorA magnesium transporters. Phenotypic analysis of the knockout mutant strain, $\Delta tmg-4$, revealed stunted vegetative growth, acquired partial cobalt resistance, and reduced levels of mycelial magnesium compared to that of the wild type strain. Further, *tmg-4* gene expression remained unchanged during vegetative development but was upregulated by three-fold in the sexual cycle. Collectively, these results validate *tmg-4* and its encoded protein as functional novel variant in the CorA superfamily magnesium transporters of fungi.

Keywords: Tmg-4, GQN, STMg, GMN, CorA Superfamily, *Neurospora crassa*, Sexual Cycle

*Correspondence: korripally@gmail.com

Citation: Radarapu S, Gorityala N, Banala UK, Sagurthi SR, Korripally P. Functional Characterization of the Tmg-4 Magnesium Transporter with the GQN Motif from *Neurospora crassa*. J Pure Appl Microbiol. 2023;17(4):2609-2619. doi: 10.22207/JPAM.17.4.60

© The Author(s) 2023. **Open Access.** This article is distributed under the terms of the [Creative Commons Attribution 4.0 International License](https://creativecommons.org/licenses/by/4.0/) which permits unrestricted use, sharing, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

INTRODUCTION

The mineral, magnesium, is vital to the structural integrity of cell organelles, macromolecules, and hundreds of enzymes involved in diverse biochemical reactions.¹⁻⁴ The levels of magnesium must be finely regulated in all cellular forms, and magnesium transporters play an important role in this process. An array of Mg²⁺ transporters has been discovered and characterized in both prokaryotes (CorA, MgtE, and MgtA/B) and eukaryotes (Mrs2, ALR1 and 2, MNR2, LPE10, SLC41, and the MGT family).⁵ The crystal structure of TmCorA from *Thermotoga maritima* was previously determined. TmCorA has a pentameric cone with two transmembrane helices. The GMN motif of TmCorA is universally conserved and present in the first transmembrane domain.^{6,7} To maintain pentameric integrity and magnesium transport, every amino acid in the GMN motif is extremely important.^{8,9} Studies using baker's yeast revealed the presence of a CorA superfamily magnesium transporter with a GMN motif. These transporters are subgrouped as ALR, MNR, and MRS2, which are localized to the plasma membrane, vacuoles, and mitochondrial membrane, respectively.¹⁰⁻¹³ These dissimilar transporter proteins of the CorA super family mediate the assimilation of magnesium from the environment to the cytoplasm and its distribution to the vacuoles and mitochondria, respectively. Besides magnesium transport in eukaryotes, CorA transporters are necessary for ascospore development, pathogenicity, regulation of virulence factor expression, and melanin synthesis in fungi.¹⁴⁻¹⁶ Root endophytic fungus *P. indica* Mg transporter (PiMgT1) is involved in the improvement of magnesium nutrition for host plants.¹⁷ This family of magnesium transporters is also necessary in pollen development, and salt and other abiotic stresses in plants.¹⁸⁻²⁰

Our group recently discovered the CorA magnesium transporter protein, Tmg-4 (NCU07816.5), which contains a GQN motif, from *Neurospora crassa*. The Tmg-4 homolog proteins with the GQN motif and CorA domain are mainly present in the Sordariomycetes class of fungi and are named STMg.²¹ The present study focused on magnesium and the role of the Tmg-4 transporter protein in various phases of *N. crassa*. Our findings

confirm that Tmg-4 bearing the GQN motif is a novel variant of the CorA superfamily magnesium transporter involved in intracellular magnesium accumulation. Overall, this study suggests that Tmg-4 with the GQN motif is necessary for the sexual phase in the heterothallic fungi, *N. crassa*.

MATERIALS AND METHODS

Strains, media, and culture conditions

The *N. crassa* strains used in this study are listed in Table 1. The strains were cultured in basal medium (BM) for vegetative growth and synthetic crossing medium (SCM) for crossing.^{22,23} The conidiospore suspension of the strains (100 µl of 1.0 O.D. at 470 nm) was used to inoculate all cultures; the strains were grown in liquid or solid medium (supplemented with 1.5% agar).

Phenotypic characterization of wild type and the knockout mutant

The gene loci, NCU07816.5, named *tmg-4* and the corresponding gene knockout strain of *N. crassa* (Δ *tmg-4*) were obtained from the FGSC (Fungal Genetic Stock Centre) for phenotypic characterization. The conidiospore suspensions of the wild type and Δ *tmg-4*,²⁴ were inoculated into 10 ml basal medium and incubated at 30°C for three days under stationary conditions for vegetative growth. To estimate mycelial growth, the wild type and Δ *tmg-4* were inoculated in liquid basal media containing magnesium concentrations ranging from 0.1 to 2.0 mM. To determine the sensitivity of the wild type and knockout toward CoCl₂ and NiCl₂, conidiospores of the wild type and Δ *tmg-4* were inoculated in liquid basal media supplemented with 0.2 mM MgSO₄ and the above cations in individual experiments. The mycelia from liquid cultures were harvested using strainers and washed thrice with deionized water. Filter papers were used to remove excess moisture from mycelia and their dry weights were recorded after drying for 3 h at 80°C or overnight at 60°C. The *N. crassa* strains were grown on solid basal media with or without MgSO₄ in race tubes to measure the rate of hyphal growth extension.²⁵

Conidia and perithecia formation

To evaluate the requirement of magnesium for the proper performance of the

asexual and sexual cycles, the *N. crassa* strains were analyzed by growing on solid basal media (2% Sorbose) or synthetic crossing media with (2.0 mM) or without MgSO₄, and with hexamine cobalt (III) chloride (HCC) (150 μM), a competitive inhibitor of CorA magnesium transport.²⁶ For the asexual cycle, wild type *N. crassa* conidia were inoculated and cultured for 7 days at 30°C. The petri plates were photographed at different time intervals to record growth and development. For the sexual cycle, opposite mating types of wild type *N. crassa* (mat A, mat a) conidia were implanted on synthetic crossing media on opposite edges of the petri plates. These cultures were maintained under light at 30°C for 9 days and the plates were subsequently photographed.

Element analysis

The *N. crassa* strains, wild type and knockout *Δtmg-4*, were grown in the basal media supplemented with various concentrations of MgSO₄ for 3 days at 30°C. The mycelia were then harvested, washed, and dried at 60°C overnight. The respective weights of the mycelia were then recorded. The dried mycelia were subjected to acid digestion in a closed microwave apparatus, and element analysis was performed using Agilent 7700x ICP-MS (Inductively coupled plasma-Mass spectrometry).

RNA isolation

For RNA isolation, the conidia of *N. crassa* were inoculated, as described previously.^{27,28} For vegetative growth, conidia (~1 × 10⁷ cells/ml) were inoculated into a 50-ml conical flask containing 10 ml of basal media and incubated at 30°C in stationary mode for 3 days. The effect of magnesium deficiency on *tmg-4* gene expression was assessed by harvesting pre-grown mycelia and transferring these mycelia to fresh basal media containing EDTA or HCC. The culture was incubated for 3 h at 30°C under stationary conditions. The wild type *N. crassa* Mat "A" (#2489) was grown on a petri dish containing synthetic crossing media covered with sterile cellophane for five days, fertilized with heavy conidia of mat "a" (#4200), and further incubated for three days. The mycelial mats (30 to 50 mg) of the vegetative cultures, unfertilized and fertilized mycelia, and perithecia were collected via scraping from crossing cultures and snap frozen with liquid nitrogen. To isolate total RNA, frozen mycelia were grounded into powder using a chilled mortar and pestle with 0.5 mm acid-washed glass beads. The remaining procedure was performed according to the manufacturer's instructions (Sigma Plant/Fungi RNA Purification Kit). The integrity of RNA was verified on a 2% agarose gel, and quantification was performed spectrophotometrically.

Table 1. Strains and Oligonucleotides used

| <i>Neurospora crassa</i> Strains | Genotype/Comment | Source / cloning site |
|----------------------------------|-----------------------------|-----------------------|
| Wild-type | 74-OR23-1a (#4200)mat a | FGSC |
| Wild-type | 74-OR23-1A(# 2489) mat A | FGSC |
| <i>Δtmg-4</i> | (#11968) mat a | FGSC |
| <i>Δtmg-4</i> | (#11969) mat A | FGSC |
| Oligonucleotide | | |
| Name | Sequence | |
| Rtmg4-F | 5'CACGATTGCCTTGCGACATA3' | |
| Rtmg4-R | 5'AGACCATTGACGCTGATCCA3' | |
| Rt gpd1-F | 5'CATCGTCGAGGGTCTCATGA3' | |
| Rt gpd1-R | 5'GTGCTGCTGGGAATGATGTT3' | |
| tmg4-F | 5'GAATTCATGGGATCCAGGTCGCC3' | EcoR1 |
| tmg4-R | 5'AAGCTTCTACACCTGCTTCTTCC3' | HindIII |

cDNA synthesis and quantitative real-time PCR

Reverse transcription of target mRNA in the total RNA was performed using oligo(dT)18 primers and Thermo Scientific Maxima First Strand cDNA Synthesis Kit, according to the manufacturer's instructions. Primer 3 software was used to design the primers used in this study.²⁹ Real-time PCR was performed on an ABI 7500 FAST (Applied biosystems) master cycler using Maxima SYBR Green/ROX qPCR Master Mix (2X). The 2- $\Delta\Delta$ Ct method was used for analysis.³⁰ *gpd-1* and beta-tubulin were used as internal controls. All qRT-PCR results were expressed relative to that of *gpd-1*. The oligonucleotides used in this study are summarized in Table 1.

RESULTS

Magnesium is required for the asexual and sexual phases of *N. crassa*

The *N. crassa* wild type strain abundantly formed asexual conidia and perithecia in media containing normal levels of magnesium (2 mM) (Figure 1). However, under magnesium depleted

conditions, asexual conidia were not produced, and approximately 50-fold less perithecia were formed in the sexual cycle, highlighting the importance of magnesium in the life cycle of *N. crassa*. Upon media supplementation with HCC (CorA specific inhibitor), the number of perithecia was reduced to a single digit in the petri dish, suggesting that magnesium uptake through the CorA transporter aids in the maintenance of intracellular Mg²⁺ in *N. crassa*. Thus, a minimum concentration of magnesium transported through the CorA transporter is critical for the development phases of *N. crassa*.

Magnesium is the 3rd most abundant cation in the mycelia of *N. crassa*

Magnesium availability plays a significant role in the asexual and sexual phases. Therefore, we estimated the levels of Mg²⁺, other cations, macro elements, and trace elements in the mycelia using ICP-MS to determine their cellular levels. Based on the results, magnesium and calcium compete for 3rd position among the tested metals, including K, Na, Mg, Ca, Fe, Zn, Mn, Cu, Ni, and

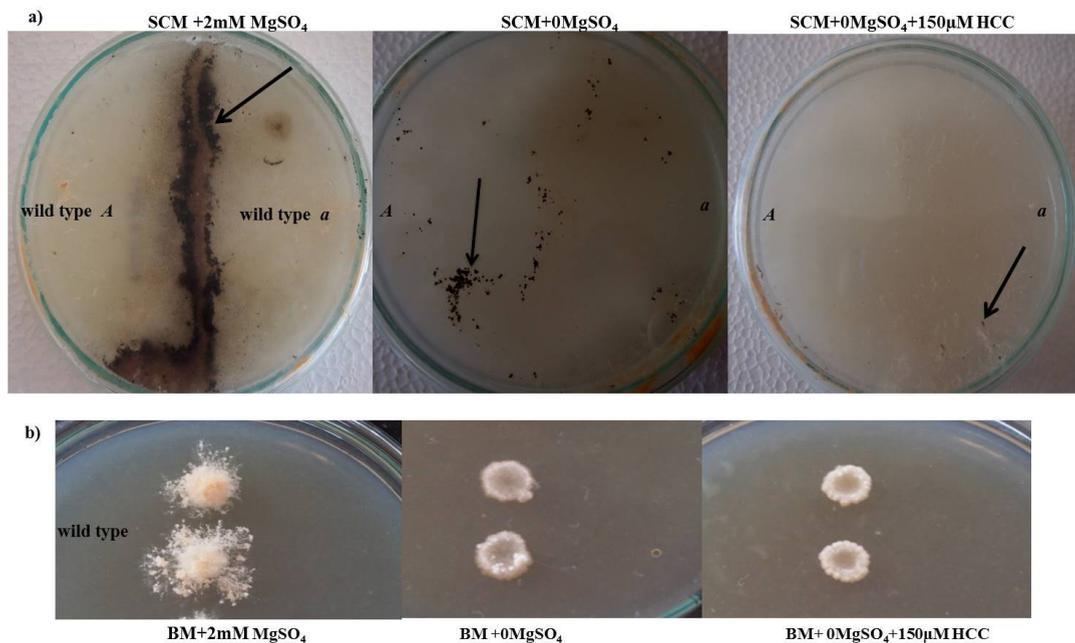


Figure 1. Perithecia and conidia formation. a) For perithecia formation, *N. crassa* opposite mating types ("A" and "a") were placed on SCM supplemented with and without MgSO₄, HCC and plates were photographed after 9 days incubation at 30°C (Arrows pointed towards perithecia) b) For conidia formation wild type strain (#4200) was grown on Solid BM with and without MgSO₄, HCC and photographed after 7 days incubation at 30°C

Co (Table 2). The sexual and asexual phases of *N. crassa* could be dependent on reported levels of magnesium in the mycelium (Figure 1).

Phenotypic characterization of the $\Delta tmg-4$ knockout mutant and wild type strains

During vegetative growth on solid media for 7 days, no significant difference was found between $\Delta tmg-4$ and wild type *N. crassa* (Figure 2a). The difference in hyphal growth extension between $\Delta tmg-4$ and the wild type was marginal when grown on solid basal media with and without $MgSO_4$ supplementation (Figure 2b). Further, to determine the growth difference between wild type and $\Delta tmg-4$, the strains were grown in liquid basal media supplemented with 2.0, 0.4, 0.2, and 0.1 mM $MgSO_4$. When magnesium concentration was decreased to 5-fold, $\Delta tmg-4$ exhibited a 20% reduction in growth compared to the wild type. Further, a 20-fold decrease in the magnesium

concentration affected the growth of $\Delta tmg-4$ by approximately 68% and wild type strain by 43% (Figure 2c). The cellular levels of magnesium in the wild type and $\Delta tmg-4$ strains of *N. crassa* were estimated under various concentrations of magnesium in the media. Under 0.4 mM to 0.1 mM $MgSO_4$, $\Delta tmg-4$ exhibited reduced levels of magnesium compared to the wild type strain (Figure 3). However, cellular magnesium levels significantly decreased in $\Delta tmg-4$ compared to that of the wild type strain under 0.4 mM $MgSO_4$. The $\Delta tmg-4$ knockout strain showed less hyphal growth extension, reduced dry weights and cellular magnesium levels, suggesting that *tmg-4* is involved in magnesium transport.

$\Delta tmg-4$ strain acquires resistance to cobalt

When the cobalt concentration increased from 50 μM to 100 μM in the media, the growth of $\Delta tmg-4$ was not affected whereas the wild type

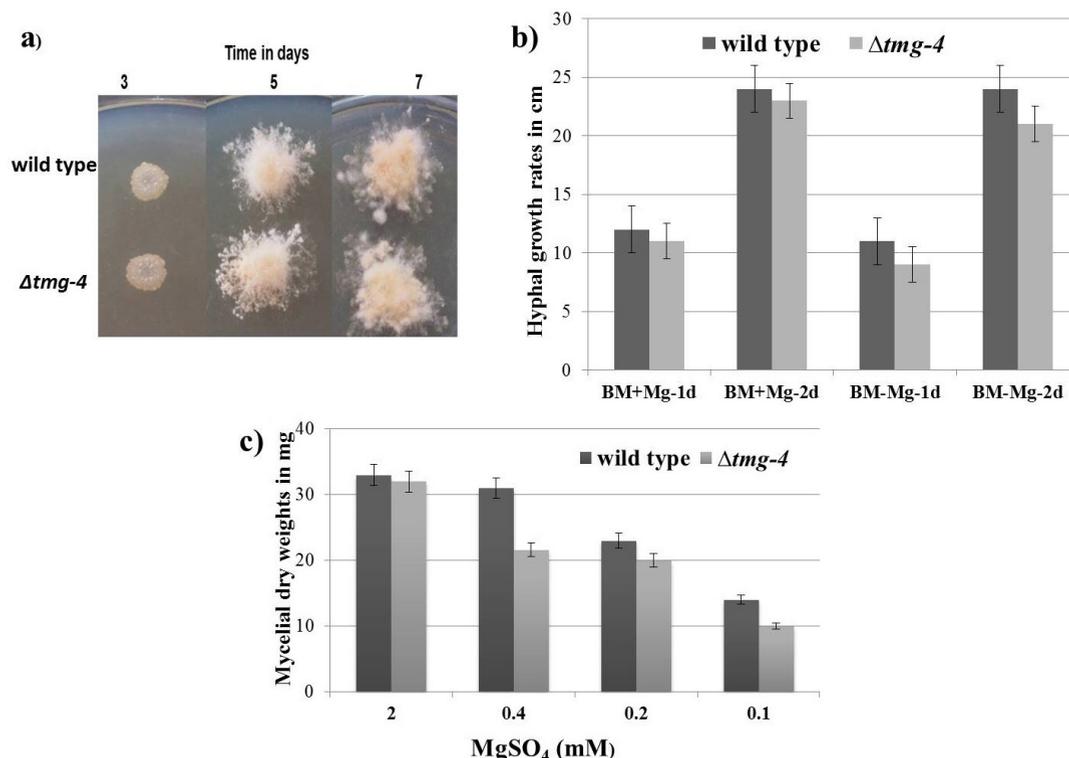


Figure 2. Phenotypic characterization of *N. crassa* strains a) 5ul of 1 O.D conidiospores suspension of wild type and $\Delta tmg-4$ were placed on basal media and incubated at 30°C. b) *N. crassa* strains were inoculated on solid BM prepared (wit and without $MgSO_4$) and in race tubes and hyphal growth extension rates were measured at 1 day and 2 days. c) *N. crassa* strains grown in liquid BM containing various concentration of $MgSO_4$ for 3 days and mycelial mats were harvested, dried and weights were recorded

strain suffered growth loss of approximately 15% (Figure 4a). Thus, the acquired resistance of $\Delta tmg-4$ towards cobalt suggests that the Tmg-4 protein transports cobalt when the media concentration of magnesium is 10-fold less. When the toxic effects of nickel were examined, this metal did not induce a growth difference between the wild type and $\Delta tmg-4$ (Figure 4b), indicating sensitivity to nickel by both strains, and highlighting that Tmg-4 may not aid in nickel transport.

Table 2. Metal ion composition. *N.crassa* wild type strain grown in Basal media with 2mM of $MgSO_4$ and subjected to acid digestion for major metal ion analysis by ICP-MS

| Metal Name | Metal content in ppb/mg mycelia |
|------------|---------------------------------|
| K | 12409.13 ±1005 |
| Na | 4092.35 ±100 |
| Mg | 1007.35 ±55 |
| Ca | 1001.00 ±60 |
| Fe | 482.42±20 |
| Zn | 93.29±10 |
| Mn | 25.99±6 |
| Cu | 16.43±2 |
| Ni | 5.02±1 |
| Co | 0.33±0.1 |

tmg-4 gene expression is marginally upregulated in the sexual cycle

The gene expression of *tmg-4* was studied in wild type grown in the presence of 100 μM , 200 μM , and 400 μM $MgSO_4$ for 3 days. The transcript

Table 3. Gene expression analysis by qRT-PCR: The *tmg-4* gene expression analyzed by qRT-PCR in various conditions. In 1) and 2) mycelia grown in Basal media containing 2mM $MgSO_4$ was taken as control to analyze the *tmg-4* expression in indicated Conditions. In 3) unfertile mycelia, before crossing was used as control. The *gpd-1* transcript was used for normalization

| 1) Condition: Vegetative growth (3 days) | Fold change |
|---|-------------|
| Basal Media(BM) with 400 μM $MgSO_4$ | 1.201±0.15 |
| BM 200 μM $MgSO_4$ | 1.143±0.10 |
| BM 100 μM $MgSO_4$ | 1.162±0.13 |
| 2) Condition: Vegetative growth (3hours) | |
| BM without $MgSO_4$ | 1.301±0.11 |
| BM with 100 μM EDTA | 1.173±0.14 |
| BM with 300 μM HCC | 1.152±0.12 |
| 3) Condition: Sexual cycle | |
| After crossing (3 days) | 3.203±0.16 |
| Perithecia (8 days) | 1.546±0.13 |

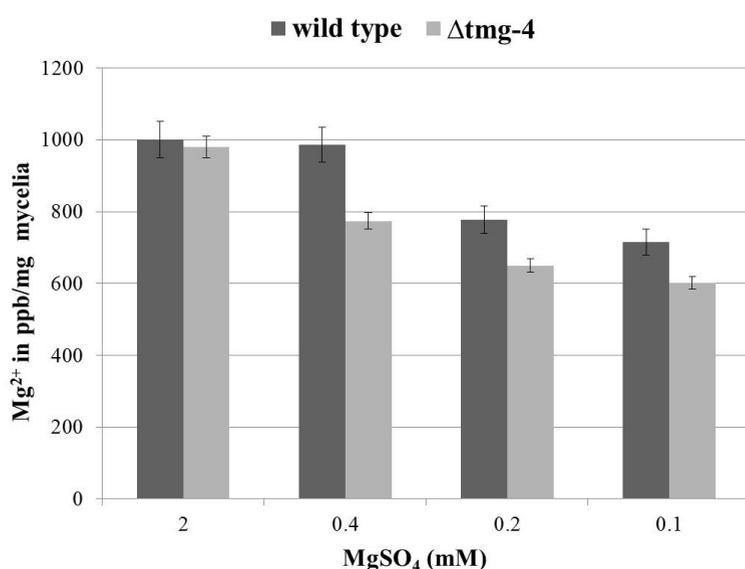


Figure 3. Graphical representation of mycelial magnesium levels in wild type and $\Delta tmg-4$ strains. *N. crassa* strains grown in BM containing various concentration of $MgSO_4$ at 30°C for 3 days. The dried mycelia were subjected to the acid digestion and elemental analysis was carried out by ICP-MS

levels of the *tmg-4* gene remained the same, irrespective of the concentration of magnesium in the media, indicating no influence of Mg^{2+} on the gene expression (Table 3). The gene expression of *tmg-4* in basal media (BM) without magnesium, BM with 100 μ M EDTA chelator of the divalent ion, and 300 μ M HCC for 3 h was also checked. In all cases, the transcript levels of *tmg-4* were similar to those in media supplemented with 2

mM $MgSO_4$, confirming that the external supply of divalent metal chelator or competitive inhibitor of magnesium could not induce a change in gene expression (Table 3). Interestingly, *tmg-4* gene expression was upregulated by 3-fold in the culture undergoing sexual development compared with the culture before crossing. In contrast, *tmg-4* expression in perithecia exhibited the same trend as that observed under vegetative conditions

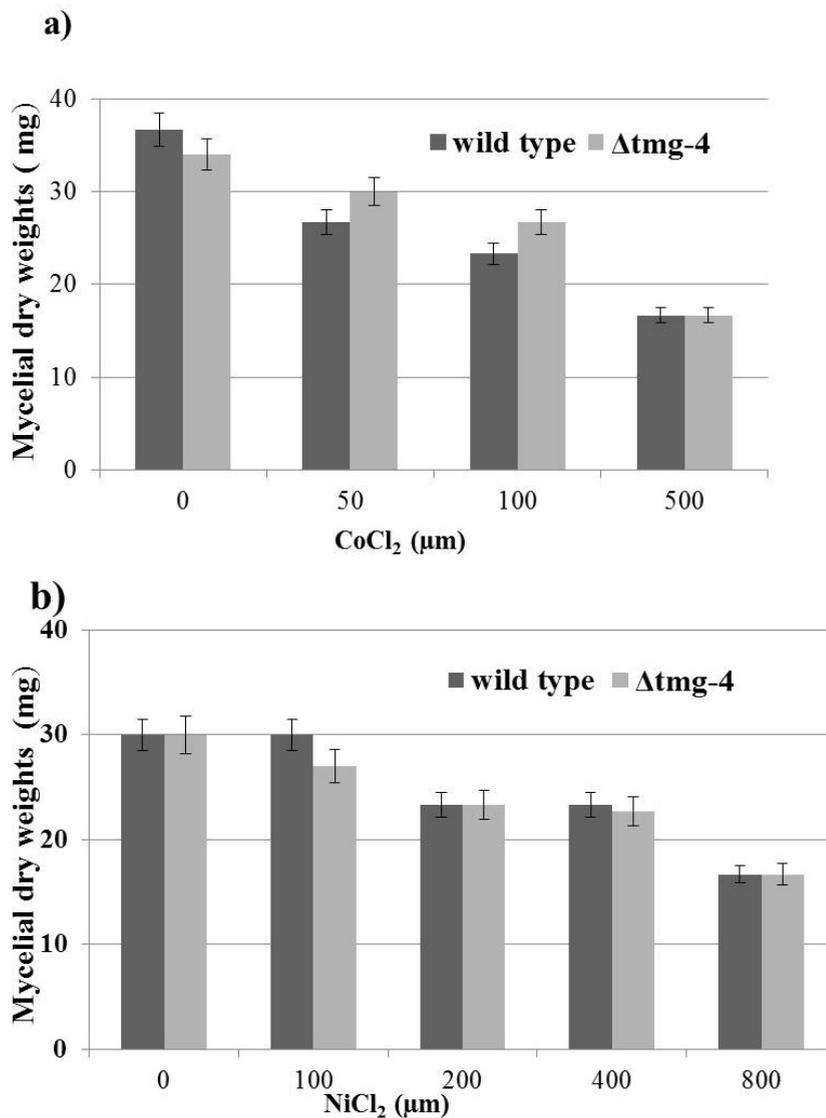


Figure 4. Mycelial growth in the presence of Cobalt and Nickel. Conidiospores of wildtype, Δ tmg-4, strains were inoculated into liquid BM containing various levels of $CoCl_2$ and $NiCl_2$. After 3 days of stationary growth mycelia were harvested and dried at 60°C for overnight and weights were recorded

(Table 3). Thus, the *tmg-4* gene may be required for the sexual development of *N. crassa*.

DISCUSSION

Importance of magnesium and its transporters in *N. crassa*

To understand the importance of magnesium in *N. crassa*, the asexual and sexual developmental phases were investigated. Herein, the asexual and sexual phases of *N. crassa* were successful, with the production of abundant conidia and perithecia in the presence of magnesium (2 mM). The lack of magnesium supplementation and chemical inhibition of Mg^{2+} transport by the CorA inhibitor, HCC, completely abolished the formation of conidia and perithecia. This remarkable effect suggests that CorA-mediated magnesium transport is the only means of transporting magnesium from media to *N. crassa*. We proceeded to examine whether a decrease in Mg^{2+} concentration affects the metal ion composition. Element analysis revealed that magnesium is the 3rd most abundant metal after sodium and potassium. In addition, the level of cellular magnesium reduced to 30% with a 10-fold decrease in magnesium concentration in basal media. Therefore, the percent of magnesium in the media might play a significant role in the physiology and development of the model fungal organism, *N. crassa*.

Tmg-4 is a CorA magnesium transporter with a GQN motif and a novel variant from Sordariomycetes

Owing to the importance of magnesium in the lifecycle of *N. crassa*, we opted to analyze the genome to determine the genes responsible for magnesium uptake from milieu to macromolecules. Preliminary studies revealed that the four gene loci belonging to the CorA superfamily are responsible for magnesium uptake.⁵ Three of the four (*tmg-1*, *tmg-2*, and *tmg-3*) genes encoded proteins comprising 2TMS and the GMN motif, which are universally conserved and hallmarks of the CorA superfamily.^{8,9} In silico analysis of four clades and structural superposition of the crystal structure of CorA (GMN motif) with a homology model of Tmg-4 (GQN motif) confirmed GQN as a novel motif and suggests its potential candidate for magnesium

transport in *N. crassa*.²¹ Similar to the Tmg-4 with GQN observation in *N. crassa*, the glycine residue on the GMN motif of CorA transporters was substituted in few plant species, which exhibited functional activities. For example, Arabidopsis AtMRS2-11 encodes an Mg^{2+} channel that mediates fast Mg^{2+} transport. The glycine residue in the GMN motif of AtMRS2-11 is essential for the transport of magnesium at low physiological Mg^{2+} concentrations. However, replacing the glycine of GMN with alanine, serine, valine, or tryptophan revealed that mutants can transport the magnesium at higher concentrations.³¹ A natural replacement for glycine was found in a CorA/MRS2-like protein of *Brachypodium distachyon* (BdMGT5-1), which forms a WMN motif.³² Wild-type maize ZmMGT6 containing the AMN motif but not the GMN motif retains its transport activity and complements yeast Mg^{2+} -deficient strains.³³ A total of 24 MGT genes were identified in wheat, and only TaMGT1A, TaMGT1B, and TaMGT1D had GMN mutations to AMN.²⁰

To verify the in silico results that the *tmg-4* encoded protein is a magnesium transporter or any other divalent transporter and validate the real nature of the gene, the *tmg-4* knockout strain was used for functional analysis. In this study, a growth defect was observed in $\Delta tmg-4$ relative to the wild type under Mg^{2+} limiting conditions (0.4 mM $MgSO_4$), whereas marginal differences in hyphal growth rates were observed between the wild type and $\Delta tmg-4$ strains. In addition, a decrease in the level of magnesium in media reveals that both strains suffered growth loss. The severity of the growth defect was further intensified by double knockout ($\Delta tmg-1$ and $\Delta tmg-4$), resulting in complete loss at 16 μM $MgSO_4$ relative to that of the wild type strain, which retained 30% growth (data not shown). This result indicates that the loss of one gene may affect the growth of the knockout strain up to 20 to 30% under magnesium-depleted conditions. Thus, our results supporting the phenotypic defects suggest that these four genes might play a significant role at different levels of magnesium transport, similar to other fungal systems. Consequently, mycelial growth in the presence of cobalt and nickel revealed that $\Delta tmg-4$ is partially resistant to cobalt compared with the wild type strain; however, nickel did

not lead to differences. We assume that cobalt toxicity in the wild type is due to the presence of four transporters, which may accumulate more cobalt than the $\Delta tmg-4$ with three transporters. Elemental analysis also revealed that $\Delta tmg-4$ accumulated 19% less cellular magnesium than the wild type. These results suggest that lack of the *tmg-4* gene in *N. crassa* affected its growth, reduced magnesium accumulation, and led to its acquisition of partial resistance to cobalt chloride. This result highlights the *tmg-4* gene encoded protein as a magnesium transporter that may be decorated in the plasma membrane of the cell. Although pBLAST analysis of the Tmg-4 revealed the homologs to the CorA and ZntB³⁴ (efflux or influx of Zinc and Cadmium) family, our analysis proves that Tmg-4 of the STMg group is a magnesium transporter that co-transporters cobalt, and is not a zinc transporter.

Functional role of Tmg -4 with the GQN motif in *N. crassa*

To gain insights into the functional role of Tmg-4, we evaluated the expression of the *tmg-4* gene in the wild type. The expression of the *tmg-4* gene was unaltered in the wild type vegetative culture in response to various concentrations of magnesium, EDTA, or HCC. In contrast, a 3-fold upregulation of gene expression was observed in the sexual cycle in fertilized mycelia relative to the unfertilized or before crossing. The *tmg-4* expression under vegetative conditions was found to be similar to that of its prokaryotic homolog, CorA of *Salmonella typhimurium*, whose expression did not markedly change under various conditions.⁵ Nevertheless, morphogenesis may be influenced by *tmg-4* expression; however, the degree of expression change is marginal. Transcriptional profiling of the wild type vegetative cultures grown on three major monosaccharides, namely D-glucose, D-xylose, and L-arabinose, revealed that NCU03312.5, NCU09091.5, and NCU11312.5 (*tmg-1*, *tmg-2*, and *tmg-3*) have two to three-fold more reads/RPKM values than *tmg-4*.³⁵ On the other hand, transcriptional profiling of the whole genome of *N. crassa* before and after crossing was investigated, which revealed that NCU07816.5 had the highest expression among

magnesium transporters.³⁶ In our study, this locus was identified as *tmg-4*, and the expression results imply that this gene may be required for the proper development of the sexual cycle in *N. crassa*.

CONCLUSION

In summary, we conclude that the G-M-N motif has a high degree of conservation in CorA magnesium transporters in prokaryotes and eukaryotes. A second conserved residue of GMN, methionine, can be functionally replaced by glutamine to form a GQN motif in the Tmg-4 transporter protein of *tmg-4* gene. Based on the experimental approaches, Tmg-4 with a novel motif (GQN) may serve as a magnesium transporter and may play a role in sexual reproduction in *N. crassa*.

ACKNOWLEDGMENTS

The authors would like to thank the Department of Biotechnology for providing the necessary facilities to carry out the work.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

AUTHORS' CONTRIBUTION

PK conceptualized the study, acquired funding, performed supervision and project administration. SRS collected resources. SR and PK designed the experiments. SR applied methodology, performed investigation and validation. NG performed formal analysis and data curation. UKB performed visualization. NG, UKB, PK and SRS wrote the manuscript. NG, UKB and SRS reviewed and edited the manuscript. All authors read and approved the final manuscript for publication.

FUNDING

This work was supported by the Science and Engineering Research Board, Government of India (grant SB/YS/LS-84/2013).

DATA AVAILABILITY

All datasets generated or analyzed during this study are included in the manuscript.

ETHICS STATEMENT

Not applicable.

REFERENCES

1. Kretsinger, RH. Magnesium in Biological Systems. In Kretsinger, RH, Uversky VN, Permyakov, EA (eds.), *Encyclopedia of Metalloproteins*, Springer, New York, N.Y. 2013:1250-1255. doi: 10.1007/978-1-4614-1533-6_258
2. Anastassopoulou J, Theophanides T. Magnesium-DNA interactions and the possible relation of magnesium to carcinogenesis. Irradiation and free radicals. *Crit Rev Oncol Hematol*. 2002;42(1):79-91. doi: 10.1016/S1040-8428(02)00006-9
3. Cowan JA. Metal activation of enzymes in nucleic acid biochemistry. *Chem Rev*. 1998;98(3):1067-1088. doi: 10.1021/cr960436q
4. Elin RJ. Magnesium: The fifth but forgotten electrolyte. *Am J Clin Pathol*. 1994;102(5):616-622. doi: 10.1093/ajcp/102.5.616
5. Maguire ME. Magnesium transporters: Properties, regulation and structure. *Front Biosci Land*. 2006;11(3):3149-3163. doi: 10.2741/2039
6. Eshaghi S, Niegowski D, Kohl A, Molina DM, Lesley SA, Nordlund P. Crystal structure of a divalent metal ion transporter CorA at 2.9 angstrom resolution. *Science*. 2006;313(5785):354-357. doi: 10.1126/science.1127121
7. Lunin VV, Dobrovetsky E, Khutoreskaya G, et al. Crystal structure of the CorA Mg²⁺ transporter. *Nature*. 2006;440(7085):833-837. doi: 10.1038/nature04642
8. Szegedy MA, Maguire ME. The CorA Mg²⁺ transport protein of *Salmonella typhimurium*. Mutagenesis of conserved residues in the second membrane domain. *J Biol Chem*. 1999;274(52):36973-36979. doi: 10.1074/jbc.274.52.36973
9. Palombo I, Daley DO, Rapp M. Why is the GMN motif conserved in the CorA/Mrs2/Alr1 superfamily of magnesium transport proteins? *Biochemistry*. 2013;52(28):4842-4847. doi: 10.1021/bi4007397
10. MacDiarmid CW, Gardner RC. Overexpression of the *Saccharomyces cerevisiae* magnesium transport system confers resistance to aluminum ion. *J Biol Chem*. 1998;273(3):1727-1732. doi: 10.1074/jbc.273.3.1727
11. Gregan J, Bui DM, Pillich R, Fink M, Zsurka G, Schweyen RJ. The mitochondrial inner membrane protein Lpe10p, a homologue of Mrs2p, is essential for magnesium homeostasis and group II intron splicing in yeast. *Mol Gen Genet*. 2001;264(6):773-781. doi: 10.1007/s004380000366
12. Weghuber J, Dieterich F, Froschauer EM, Svidová S, Schweyen RJ. Mutational analysis of functional domains in Mrs2p, the mitochondrial Mg²⁺ channel protein of *Saccharomyces cerevisiae*. *FEBS J*. 2006;273(6):1198-1209. doi: 10.1111/j.1742-4658.2006.05157.x
13. Pisat NP, Pandey A, MacDiarmid CW. MNR2 regulates intracellular magnesium storage in *Saccharomyces cerevisiae*. *Genetics*. 2009;182(1):874-884. doi: 10.1534/genetics.109.106419
14. Grognet P, Lalucque H, Silar P. The PaAlr1 magnesium transporter is required for ascospore development in *Podospora anserina*. *Fungal Biol*. 2012;116(10):1111-1118. doi: 10.1016/j.funbio.2012.08.004
15. Reza MH, Shah H, Manjrekar J, Chattoo BB. Magnesium uptake by cora transporters is essential for growth, development and infection in the rice blast fungus *magnaporthe oryzae*. *PLoS One*. 2016;11(7):e159244. doi: 10.1371/journal.pone.0159244
16. Suo CH, Ma LJ, Li HL, et al. Investigation of *Cryptococcus neoformans* magnesium transporters reveals important role of vacuolar magnesium transporter in regulating fungal virulence factors. *Microbiologyopen*. 2018;7(3):e00564. doi: 10.1002/mbo3.564
17. Prasad D, Verma N, Bakshi M. et al. Functional characterization of a magnesium transporter of root endophytic fungus *Piriformospora indica*. *Front Microbiol*. 2019;9:3231. doi: 10.3389/fmicb.2018.03231
18. Mohamadi SF, Babaeian Jelodar N, Bagheri N, Nematzadeh G, Hashemipetroudi SH. New insights into comprehensive analysis of magnesium transporter(MGT) gene family in rice(*Oryza sativa* L.). *3 Biotech*. 2023;13(10):322. doi: 10.1007/s13205-023-03735-4
19. Chen J, Li LG, Liu, ZH, et al. Magnesium transporter AtMGT9 is essential for pollen development in Arabidopsis. *Cell Res*. 2009;19(7):887-898. doi: 10.1038/cr.2009.58
20. Tang Y, Yang X, Li H, et al. Uncovering the role of wheat magnesium transporter family genes in abiotic responses. *Front Plant Sci*. 2023;14:1078299. doi: 10.3389/fpls.2023.1078299
21. Sireesha R, Neelima G, Udaykumar B, Someswar RS, Premsagar K. *In silico* analysis of CorA superfamily magnesium transporters of fungi and identification of magnesium transporter STMg with GQN motif from *Neurospora crassa*. *Global Journal For Research Analysis*. 2023;12(9). doi: 10.36106/gjra
22. Mohan PM, Sastry KS. Interrelationships in trace-element metabolism in metal toxicities in nickel-resistant strains of *Neurospora crassa*. *Biochem J*. 1983;212(1):205-210. doi: 10.1042/bj2120205
23. Westergaard M, Mitchell HK. *Neurospora* V. A Synthetic Medium Favoring Sexual Reproduction. *Am J Bot*. 1947;34(10):573-577. doi: 10.2307/2437339
24. Colot HV, Park G, Turner GE, et al. A high-throughput gene knockout procedure for *Neurospora* reveals functions for multiple transcription factors. *Proc Natl Acad Sci*. 2006;103(27):10352-10357. doi: 10.1073/pnas.0601456103
25. Ryan FJ, Beadle GW, Tatum EL. The tube method of measuring the growth rate of *Neurospora*. *Am J Bot*. 1943;30(10):784-799. doi: 10.2307/2437554
26. Kucharski LM, Lubbe WJ, Maguire ME. Cation hexaamines are selective and potent inhibitors of the CorA magnesium transport system. *J Biol Chem*. 2000;275(22):16767-16773. doi: 10.1074/jbc.M001507200
27. Nelson MA, Kang S, Braun EL, et al. Expressed sequences from conidial, mycelial, and sexual stages of *Neurospora crassa*. *Fungal Genet Biol*. 1997;21(3):348-

363. doi: 10.1006/fgbi.1997.0986
28. Nelson MA, Metzenberg RL. Sexual development genes of *Neurospora crassa*. *Genetics*. 1992;132(1):149-162. doi: 10.1093/genetics/132.1.149
29. Rozen S, Skaletsky H. Primer3 on the WWW for general users and for biologist programmers. *Methods Mol Biol.* 2000;132:365-386. doi: 10.1385/1-59259-192-2:365
30. Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2- $\Delta\Delta$ CT method. *Methods*. 2001;25(4):402-408. doi: 10.1006/meth.2001.1262
31. Ishijima S, Shiomi R, Sagami I. Functional analysis of whether the glycine residue of the GMN motif of the Arabidopsis MRS2/MGT/CorA-type Mg²⁺ channel protein AtMRS2-11 is critical for Mg²⁺ transport activity. *Arch Biochem Biophys*. 2021;697:108673. doi: 10.1016/j.abb.2020.108673
32. Regon P, Chowra U, Awasthi, JP, Borgohain P, Panda SK. Genome-wide analysis of magnesium transporter genes in *Solanum lycopersicum*. *Comput Biol Chem.* 2019;80:498-511. doi: 10.1016/j.compbiolchem.2019.05.014
33. Huang K, Chen X, Liu T, et al. Identification, and functional and expression analyses of the CorA/MRS2/MGT-type magnesium transporter family in maize. *Plant and Cell Physiol.* 2016;57(6):1153-1168. doi: 10.1093/pcp/pcw064
34. Worlock AJ, Smith RL. ZntB is a novel Zn²⁺ transporter in *Salmonella enterica* serovar Typhimurium. *J Bacteriol.* 2002;184(16):4369-4373. doi: 10.1128/JB.184.16.4369-4373.2002
35. Li J, Lin L, Li H, Tian C, Ma Y. Transcriptional comparison of the filamentous fungus *Neurospora crassa* growing on three major monosaccharides D-glucose, D-xylose and L-arabinose. *Biotechnol Biofuels*. 2014;7(31):6. doi: 10.1186/1754-6834-7-31
36. Lehr NA, Wang Z, Li N, Hewitt DA, Lopez-Giraldez F, Trail F, Townsend JP. Gene expression differences among three *Neurospora* species reveal genes required for sexual reproduction in *Neurospora crassa*. *PLoS One*. 2014;9(10):e110398. doi: 10.1371/journal.pone.0110398