

## Effect of Thidiazuron with Sucrose Pulsing and Low Temperature Storage with Polyfilm Packaging on Floret Opening and Abscission of Tuberose Cut Spikes

Homraj Sahare<sup>1</sup>, Alka Singh<sup>2</sup>, Gurpreet Singh<sup>1\*</sup>, R. Sharma<sup>1</sup> and P. Kumari<sup>1</sup>

<sup>1</sup>School of Agriculture, Lovely Professional University Punjab India.

<sup>2</sup>Department of Floriculture and Landscape Architecture, ASPEE College of Horticulture and Forestry, NAU, Navsari - 396 450, Gujarat India.

<http://dx.doi.org/10.22207/JPAM.12.3.41>

(Received: 10 June 2018; accepted: 18 July 2018)

**Tuberose is a perennial plant and mostly use as cut flower as well as loose flower. Low temperature storage of tuberose flower spike decreases or inhibits floret opening and triggers floret abscission adversely affecting the flower quality. The investigations were conducted to study the influence of pre-storage pulsing with TDZ with sucrose and low temperature storage with poly film packaging on post storage quality of tuberose cut spikes. Pre-storage pulsing, poly film packaging and low temperature storage at 2 °C significantly influenced post-harvest floret opening tuberose flower spikes. Pre-storage pulsing of tuberose spikes with solution containing 0.5 mM TDZ +15% sucrose or for six hours, followed by 2 °C low temperature storage under seal packaging with HDPE 40 μ for fifteen days, significantly improved physiological parameters like water uptake (ml), total soluble sugars and electrolyte leakage (%) in florets tissue. This further lead to improved per cent floret opening and decreased abscission during post storage life.**

**Keywords:** Thiadiazuron, Sucrose, Electrolyte leakage, water uptake, Fresh weight, abscission, tuberose, floret opening.

The sturdy fragrant flower spikes of tuberose (*Polianthes tuberosa* L.) with serially arranged florets are very popular but are perishable in nature with vase life of 6-8 days. Opened fresh florets determine its postharvest quality. Low temperature storage adversely affects its flower quality with reduced floret or inhibited floret opening that further deteriorates with increase in storage duration<sup>37</sup>. Further, low temperature storage has also been known to promote ethylene production that further triggers floral abscission and early senescence in tuberose. Storage temperature and duration also influence metabolic activities of cut

flower. Various enzymatic activities like protease, lipoxygenase, *etc* are known to increase during senescence process<sup>37</sup>. Improvement in postharvest quality of fresh tuberose spikes by pulsing with Thidiazuron has been earlier observed<sup>15</sup>.

Wet storage ensures flower quality but only for short duration as long duration storage results into increase in the stage of bud opening which again declines its market value. Benefits of passive modified atmosphere storage system along with pre or post-storage treatments have been earlier observed gladiolus spikes<sup>31</sup>. Passive modified atmosphere using films like Polypropylene (PP), High Density Polyethylene (HDPE), Low Density Polyethylene (LDPE) *etc*. Having differential air permeability property can be employed for long term storage of flowers. The key to successful modified storage of fresh flowers

\* To whom all correspondence should be addressed.  
E-mail: gurpreetraje@rediffmail.com  
Homraj.sahare@rediffmail.com.

is to use packaging films of suitable permeability so as to ensure and establish the optimal Equilibrium Modified Atmosphere (EMA) at low temperature<sup>4</sup>. Thus, there is need to cold store tuberose cut spike when there is market glut causing market fluctuations. However, vase life and floret opening of cut tuberose spikes are known to decrease after cold storage<sup>37</sup>. According to prior research, tuberose cut spikes can be stored only for 3 day at 2°C and Packaging has been known to play an important role in flower quality, appearance, reduces water loss and opening ability in stored cut flowers<sup>37, 36</sup>.

Pulsing treatments constituting of germicides and sugar are used to improve flower opening, flower size, shape, colour and longevity of cut flowers like tuberose, gladiolus and rose<sup>16</sup>. Sucrose replaces the depleted endogenous carbohydrates utilized during postharvest life of flowers<sup>24</sup>. From the market point of view, the most important components of quality are the shape, size, surface, and freshness of the flower. The marketability of flowers depends on maintenance of these components and freshness at optimum. During post-harvest life physiological activities like respiration and transpiration causes continuous depletion of CHOs and water in cut flowers<sup>10, 13</sup>.

A lot of research on postharvest aspects is being conducted on tuberose; however, systematic research on storage of cut tuberose spikes is meager. Thus there is need to evaluate optimum pulse, storage technology for tuberose flowers using chemicals and packaging films. Hence, the experiment was planned to standardize packaging film for dry storage aiming for prolonging storage for cut tuberose under cold storage condition at 2°C.

## MATERIALS AND METHODS

Fresh spikes of tuberose, var. 'Prajwal' were obtained from Floriculture farm, ASPEE College of Horticulture and Forestry, Navsari Agricultural University, Navsari (Gujarat) and brought to the Floricultural laboratory at ambient temperature (20-25°C). The experiment was laid out in Completely Randomized Design with Factorial concept (FCRD). There were 15 treatment combinations and each treatment was replicated three times. Tuberose spikes at two basal florets

opened stage were sorted and selected for uniform sizes (80± 5) cm and fresh weight (90± 5g). They were randomly assigned to treatments and re-cut to 70 cm. Polyfilms used for seal packaging of tuberose spikes were polypropylene (PP of 40 and 30 μ), High Density Polythene (HDPE of 40 μ) and Low Density Polythene (LDPE of 40 μ). Fifteen treatment combinations of pre-storage treatments (0.5 mM TDZ along with 15% sucrose) and storage methods comprising of seal packaging with polyfilms (HDPE, PP and LDPE), wet storage (water) as controls were prepared. Spikes were divided into three groups *viz.*, A, B and C, given pre-storage treatment of distilled water, 15% sucrose and 0.5 mM TDZ with 15 % sucrose respectively. Pre-storage treatment was given at ambient temperature 20-25°C, 60±5% RH for a period of 6 h. Thereafter, spikes from each group were divided into bundles each having 10 spikes. From each group, equal numbers of bundles were seal packed with PP, HDPE and LDPE poly films in size 75 cm x 15 cm. Further, same numbers of tuberose spikes from each group were held in water and same numbers of tuberose spikes were also kept as such without any packaging as controls. All the treated tuberose spikes (packed and those held in water) along with controls were then immediately placed at 2°C low temperature storage for a period of 15 days. 15 days later, spikes were taken out from low temperature storage, removed from packaging and held in distilled water for recording observations in laboratory at ambient temperature (20-25°C), 60±5% relative humidity and average radiation around 18 μmolm<sup>-2</sup>s<sup>-1</sup> for a period of 8+2h day<sup>-1</sup>. Total soluble petal sugars, and electrolyte leakage of cell membrane in the petal tissue were estimated (in triplicate) from spikes held in distilled water in vase after 15 days of storage using third, fourth and fifth floret from the basal end on day 3 and day 5. Per cent relative fresh weight of tuberose spikes was also estimated on day 3 and day 5.

### Post storage vase life and total water uptake measured at alternate day (ml) during the vase life period

Post storage vase life was measured in days at the time of holding the tuberose spikes in vase after LT storage until the first sign of wilting or necrotic drying to the death of the 50 % florets spike during the vase life. The water uptake by

**Table 1.** Effect of pre-storage pulsing, packaging and storage at 2°C on total water uptake (ml), Total soluble sugar (%) by tuberose cut spikes cv. Prajwal

Treatments	Total water uptake (ml)			Total Soluble Sugar (µg/ml) on day 2			Total Soluble Sugar (µg/ml) on day 4			Total Soluble Sugar (µg/ml) on day 6		
	(T <sub>0</sub> ) No pulsing	(T <sub>1</sub> ) Sucrose 15 %	(T <sub>2</sub> ) TDZ 0.5 mM +15 % Sucrose	(T <sub>0</sub> ) No pulsing	(T <sub>1</sub> ) Sugar 15 %	(T <sub>2</sub> ) TDZ 0.5 mM +15 % Sucrose	(T <sub>0</sub> ) No pulsing	(T <sub>1</sub> ) Sugar 15 %	(T <sub>2</sub> ) TDZ 0.5 mM +15 % Sucrose	(T <sub>0</sub> ) No pulsing	(T <sub>1</sub> ) Sugar 15 %	(T <sub>2</sub> ) TDZ 0.5 mM +15 % Sucrose
P <sub>1</sub> 40 µ LDPE	115.62	118.54	122.21	10.53	11.20	12.85	9.03	9.36	11.42	7.28	8.15	9.37
P <sub>2</sub> 40 µ HDPE	-µ	160.82	166.54	175.39	13.44	14.78	16.65	11.27	12.73	15.01	9.01	9.35
P <sub>3</sub> 20 µ PP	117.71	119.71	139.45	11.23	12.23	13.37	8.06	8.76	12.08	6.46	7.46	9.26
P <sub>4</sub> 40 µ PP	131.45	133.86	147.32	10.98	11.31	14.05	9.85	10.02	12.56	8.05	8.32	10.43
P <sub>5</sub> control (Holding in water)	47.59	51.69	49.29	9.05	9.85	10.15	*	*	*	*	*	*
	T	P	TXP	T	P	TXP	T	P	TXP	T	P	TXP
LSD	1.19	1.68	2.91	0.28	0.40	0.69	0.23	0.33	0.56	0.11	0.16	0.28

\* (-) No observation was recorded as vase life was over due to chilling injury.

the cut spike was recorded at regular intervals (volume ml) and total water uptake was determined by summation at the end of vase life evaluation.

#### Total soluble sugar (TSS)

Floret petals (100 mg) were extracted with 5 ml of 80% ethanol and centrifuged at 3000 rpm for 10 minutes. Extraction was repeated 4 times with 80% ethanol and supernatants were collected into 25 ml volumetric flasks. Final volume of the extract was made to 25 ml with 80% ethanol. The

extract (0.3 ml) was taken from treatments into separate test tubes and the tubes were placed in a boiling water bath for 3 minutes to evaporate the ethanol. One ml of Millipore water and 4 ml of 0.2% anthrone reagent (200 mg in 100 ml H<sub>2</sub>SO<sub>4</sub>) was added in each test tube and placed in ice cold water. Reagent blank was prepared by adding 1 ml of distilled water and 4 ml of anthrone reagent. The intensity of colour was read at 600 nm on spectrophotometer. A standard curve was prepared

**Table 2.** Effect of pre-storage pulsing and packaging films on per cent floret opening in tuberose cut spikes cv. Prajwal

Treatments	% Florets opening on day 2			% Florets opening on day 4			% Florets opening on day 6		
	(T <sub>0</sub> ) No pulsing	(T <sub>1</sub> ) Sucrose 15 %	(T <sub>2</sub> ) TDZ 0.5 mM +15 % Sucrose	(T <sub>0</sub> ) No pulsing	(T <sub>1</sub> ) Sucrose 15 %	(T <sub>2</sub> ) TDZ 0.5 mM +15 % Sucrose	(T <sub>0</sub> ) No pulsing	(T <sub>1</sub> ) Sugar 15 %	(T <sub>2</sub> ) TDZ 0.5 mM +15 % Sucrose
P <sub>1</sub> 40 µ LDPE	20.33	22.00	24.33	58.30	60.63	76.39	21.69	24.35	30.63
P <sub>2</sub> 40 µ HDPE	32.66	33.33	36.00	63.44	70.10	80.60	26.50	30.50	35.43
P <sub>3</sub> 20 µ PP	25.33	26.66	31.00	58.42	61.09	75.59	21.27	25.27	31.37
P <sub>4</sub> 40 µ PP	23.66	27.33	30.33	61.56	64.89	76.51	23.90	26.24	33.52
P <sub>5</sub> control (Holding in water)	20.00	21.66	23.33	-	-	-	-	-	-
	T	P	TXP	T	P	TXP	T	P	TXP
LSD	0.52	0.74	1.27	0.91	1.29	2.23	0.29	0.41	0.71

\* (-) Control spikes failed to show any floret opening due to chilling injury.

**Table 3.** Effect of pre-storage pulsing and packaging films on maximum numbers of floret open per spike at one time and total number floret abscission per spike in tuberose cut spikes cv. Prajwal

Treatments	Maximum numbers of open florets / spike at one time (Number)			Total number of floret abscission per spike (Number)		
	(T <sub>0</sub> ) No pulsing	(T <sub>1</sub> ) Sugar 15 %	(T <sub>2</sub> ) TDZ 0.5 mM +15 % Sucrose	(T <sub>0</sub> ) No pulsing	(T <sub>1</sub> ) Sugar 15 %	(T <sub>2</sub> ) TDZ 0.5 mM +15 % Sucrose
P <sub>1</sub> 40 µ LDPE	2.83	3.00	4.00	4.00	3.00	3.00
P <sub>2</sub> 40 µ HDPE	4.48	5.15	6.00	3.00	2.86	2.00
P <sub>3</sub> 20 µ PP	3.00	3.33	3.33	4.00	3.51	3.25
P <sub>4</sub> 40 µ PP	4.00	4.33	5.00	3.15	3.00	3.00
P <sub>5</sub> control (Holding in water)	2.03	3.00	3.00	5.00	4.00	3.75
	T	P	TXP	T	P	TXP
LSD	0.09	0.13	0.23	0.11	0.14	0.24

using 100 mg glucose per 100 ml distilled water<sup>7</sup>. The T.S.S. in the petal tissue were calculated with the formula given below.

#### Formula,

Total soluble sugar ( $\mu\text{g}/\text{mg}$ ) = sample O.D. x Standard O.D. x dilution factor

#### Per cent floret opening

Per cent floret opening in spike was recorded on the basis of number of florets observed per spike in per cent form at alternate days during vase life.

$$\text{Floret opening} = \frac{\text{Number of floret opens on particular day}}{\text{Total florets open}} \times 100$$

#### Maximum number of florets opened/ spike at one time measurement

Maximum no. of florets opened/ spike at one time was recorded on the basis of number, maximum number of florets visible per spike at one time during vase life.

#### Total number of florets abscission/spike measurement

The total number of florets abscised/spike was measured in number from the time of keeping the spike in vase until the first sign of wilting or necrotic drying to the death of the 50 % florets of during the vase life.

#### Electrolyte leakage

The electrolyte leakage was measured by taking five petal discs ( $1\text{cm}^2$ ) of flower on days 2, 4 and 6. The petal discs were rinsed well in deionized water prior to incubation in 5 ml of deionized water for 3 h. at room temperature. After incubation, conductivity of the bathing solution was measured with the conductivity meter (value A). The petal discs were boiled with bathing solution for 10 minutes to kill tissue. After cooling to room temperature, the conductivity of bathing solution was again measured (value B).

Formula, Electrolytic leakage (%) = (value A/ value B) X 100

#### Statistical Analysis

The data recorded during the course of investigation were subjected to the statistical analysis. The design of analysis used was Completely Randomized Design with Factorial concept as described by<sup>23</sup>. The significance of treatment differences was tested by 'F' test on the basis of null hypothesis. Mean comparisons were performed using LSD test to determine whether the difference between the variables were significant

at  $P \leq 0.05$ .

## RESULTS AND DISCUSSION

Significantly maximum total water uptake (175.39 ml) was recorded in treatment combination comprising of pre-storage treatment of 0.5 mM TDZ + 15% sucrose and in 40  $\mu$  HDPE packaging. Minimum water uptake (47.59 ml) was recorded in low temperature wet stored flower spikes (control). Tuberose spikes given pre-storage treatment of 0.5 mM TDZ with 15% sucrose and stored in 40  $\mu$  HDPE polyfilm packaging at 2°C low temperature storage showed maximum post storage vase life of 22 days. Tuberose spikes stored by holding in water (control) at low temperature storage displayed minimum vase life, irrespective of the pre storage treatment.

Maximum total soluble sugars (16.65, 15.01 and 11.53  $\mu\text{g}/\text{mg}$  Fw) on day 2, day 4 and day 6 respectively was recorded in petal tissue of low temperature stored tuberose spikes treated with 0.5 mM TDZ + 15% sucrose and stored in 40  $\mu$  HDPE seal packaging. Low temperature wet stored tuberose spikes retained low total soluble sugars in petal tissue on day 2 while observation could not be recorded on day 4 and 6 due to early senescence.

HDPE and PP were more effective in modifying atmospheric conditions with high  $\text{CO}_2$  and lower  $\text{O}_2$  as compare to LDPE within the seal packages as the permeability of these films to  $\text{CO}_2$  and water vapour is lower as compared to LDPE<sup>6</sup>. HDPE and PP packaging thus, contributed to higher water uptake and minimal cell damage during storage and retaining of normal cell condition after storage as indicated by minimal electrolyte leakage in the petal tissue (Table 3) and high TSS in the petal tissue. Further, sucrose being osmoticum<sup>12,2,13</sup> as in pre-storage treatment contributed in influencing high water uptake as well as high TSS in petal tissue of tuberose cut spikes as also observed in gladiolus<sup>31</sup>. Positive influence of sucrose pulsing with modified atmosphere low temperature storage on petal sugar level and water uptake has been earlier indicated in gladiolus<sup>19</sup>. Low water uptake in wet stored (controls) tuberose cut spikes after storage was due to gradual advancement in senescence stage as evident from electrolyte leakage (Table 4).

Restriction in water uptake due to damaged xylem structure on account of desiccation during low temperature storage has been indicated in cut flowers<sup>6</sup>. In addition to this, cytokinins have also been known to enhance enzymatic activities for increasing sucrose availability in the cell<sup>1</sup>, that further facilitate water uptake and fresh weight retention in cut flowers as also observed in gladiolus<sup>30</sup>. Since, thidiazuron (TDZ; N-phenyl-N\_-1,2,3-thiadiazol-5-ylurea) is a non-metabolizable compound with strong cytokinin like activity<sup>20</sup> would have been highly effective.<sup>18</sup> also indicated strong role of TDZ in regulation of flower metabolism.

Further, the retention of high petal sugar level (higher TSS) along with low electrolyte

leakage in the petal tissue (Table 4) and high water uptake by the cut spikes contributed to higher bud opening in prestorage treated and HDPE or PP packaged tuberose cut spikes. Water balance<sup>11</sup> along with intact cell membrane of petal tissue was suggested to improve bud opening in cut flowers as also observed in rose<sup>32</sup>. Further, carbohydrate content in petals or tepals has been directly link to flower opening in tuberose<sup>35</sup>. Modified atmosphere conditions within the package have been known to regulate metabolic activity in flowers and influence flower opening<sup>8</sup>. Improved percent of floret opening per spike with TDZ in along with sucrose was observed earlier also in iris<sup>18</sup>.

0.5 mM TDZ along with 15% sucrose as pre-storage treatments and seal packaging with

**Table 4.** Effect of pre-storage pulsing, packaging and storage at 2° C on Electrolyte leakage 6 DAS and Total vase life by tuberose cut spikes cv. Prajwal

Treatments	Electrolyte leakage on day 2 (%)			Electrolyte leakage on day 4 (%)			Total vase life (Days)		
	(T <sub>0</sub> ) No pulsing	(T <sub>1</sub> ) Sucrose 15 %	(T <sub>2</sub> ) TDZ 0.5 mM +15 % Sucrose	(T <sub>0</sub> ) No pulsing	(T <sub>1</sub> ) Sucrose 15 %	(T <sub>2</sub> ) TDZ 0.5 mM +15 % Sucrose	(T <sub>0</sub> ) No pulsing	(T <sub>1</sub> ) Sugar 15 %	(T <sub>2</sub> ) TDZ 0.5 mM +15 % Sucrose
P <sub>1</sub> 40 μ LDPE	45.52	43.52	39.27	50.52	48.64	45.03	18.30	18.10	20.33
P <sub>2</sub> 40 μ HDPE	38.37	35.37	30.16	39.22	36.78	34.52	19.70	20.00	22.00
P <sub>3</sub> 20 μ PP	45.86	43.53	42.37	53.03	50.32	48.60	17.42	19.03	20.66
P <sub>4</sub> 40 μ PP	47.41	45.41	44.03	55.18	52.95	49.70	19.55	19.67	21.00
P <sub>5</sub> control (Holding in water)	75.22	74.22	70.31	-	-	-	17.23	17.39	18.39
	T	P	TXP	T	P	TXP	T	P	TXP
LSD	0.38	0.54	0.93	0.86	1.21	2.10	0.16	0.20	0.35

\* (-) No observation was recorded as vase life was over due to chilling injury.

polyfilms appreciably influenced florets opening of tuberose flower spikes as recorded after storage on day 2, day 4 and day 6 when spikes were held in vase water after removal from packages. Significantly maximum number of florets opened (36.00, 80.60 and 35.43% respectively) was recorded on day 2, 4 and 6 were recorded in treatment combination comprising of pre-storage treatment of 0.5 mM TDZ + 15% sucrose and in 40 μ HDPE packaging. Minimum per cent florets opening (20%) on day 2 were recorded in low temperature wet stored flower spikes while on day

4 and day 6 observation could not be recorded as the post storage vase life was over.

Tuberose spikes given pre-storage treatment of 0.5 mM TDZ with 15% sucrose and stored in 40 μ HDPE polyfilm packaging at 2°C low temperature storage showed maximum number of florets opened per spikes (6.00). Tuberose spikes stored by holding in water (control) at low temperature storage displayed minimum number of florets opened per spikes (2.83), irrespective of the pre storage treatment. In the parameter of floret abscission, Tuberose spikes given pre-storage

treatment of 0.5 mM TDZ with 15% sucrose and stored in 40 µ HDPE polyfilm packaging at 20°C low temperature storage noted minimum number of florets abscised per spikes (2.00). Tuberose spikes stored by holding in water (control) at low temperature storage recorded maximum total number of abscised florets per spikes (5).

Minimum per cent electrolyte leakage (30.16 and 34.52%) on day 2 and day 4 respectively was recorded in treatment combination comprising of pre-storage treatment with 0.5 mM TDZ+ 15 % sucrose and stored with 40 µ HDPE packaging. Electrolyte leakage was extremely high (75.22 and 55.18% respectively) in the petal tissue of untreated low temperature stored tuberose spikes on day 2 and day 4 when held in water after storage.

Abscission in flowers is triggered by elevated ethylene<sup>2,33</sup> deficient carbohydrates status<sup>8</sup> other hormonal imbalance of auxin and abscisic acid and water and temperature stress<sup>3</sup>. Pre-storage pulsing of cut flowers with sucrose has been reported to maintain the cell membrane integrity and reducing their sensitivity to ethylene during low temperature storage<sup>17</sup>. Further, <sup>37</sup>associated low carbohydrates reserves dry cold storage with ethylene sensitivity and stimulates floret abscission in tuberose. Pre-storage sucrose pulsing also inhibited the premature abscission of gladiolus young petals<sup>37</sup>. Cytokinin help to reduce the flower tissue sensitivity to ethylene and thus increase vase life of tuberose cut spikes. Cytokinines known reduce the ethylene biosynthesis and response<sup>5,17</sup>. Further TDZ is also known to regulate endogenous cytokinin biosynthesis and metabolism<sup>21</sup>. In addition to this TDZ may act by mimicking auxin activity<sup>22</sup> and high level of auxin are directly linked with reduction in abscission<sup>28</sup>. In addition, inclusion of 5–45 µM TDZ in vase water was found to reduce ethylene-mediated flower abscission and senescence on phlox and lupin stems, respectively<sup>25,26</sup>. α-lipoic acid being an anti-oxidant and anti-ageing agent as a free radical scavenger<sup>27</sup> consequently delayed senescence (indicated by enhanced vase life, Table-4) that, further contributed to increase in water uptake and fresh weight retention. These results are in agreement with the work of<sup>29</sup> in gladiolus. The modified atmosphere have been known to minimize the flower abscission of reduce the respiration rate,

alleviations ethylene damage and preventing water loss from the cut stems<sup>8</sup>.

## ACKNOWLEDGMENTS

This research was supported by Navsari Agricultural University, Gujarat. We are thankful to our professors, technical staff and lab technician for greatly support and guide during this research.

## REFERENCES

- Balibrea, M. E.; Gonzalez, M. C.; Fatima, T. Ehness, R.; Lee, T. K.; Proels, R. and Roitsch, T. Extracellular invertase is an essential component of cytokinin mediated delay of senescence. *Pl. Cell.* 2004; **16**: 1276-1287.
- Borochoy, A.; Tirosh, T. and Halevy, A. H. Abscisic acid content of senescing petals of cut rose flowers as affected by sucrose and water stress. *Pl. Physiol.* 1976; **58**: 175-178.
- Cameron, A. C. and Reid, M. S., 1-MCP blocks ethylene –induced petal abscission of pelargonium peltatum but the effect is transient. *Postharvest Biol. Technol.* 2001; **22**: 169-177.
- Day, B. P. F. Modified atmosphere packaging of fresh fruits and vegetables. *Acta Horticulture.* 2001; **553**: 585-590.
- Eisinger, W. Role of cytokinin in carnation flower senescence. *Pl. Physiol.*, 1977; **59**: 707-709.
- Farber, J. N.; Harris, L. J.; Parish, M. E.; Beuchat, L. R.; Suslow, T. V.; Gorney, J. R.; Garrett, E. H. and Busta, F. F. Microbiological safety of controlled and modified atmosphere packaging of fresh and fresh-cut produce. *Comprehensive review in food science and food safety.* 2003; **2**: 142- 158.
- Franciscitt, W., David, F.B. and Robert, M.D. The estimation of the total soluble carbohydrate in cauliflower tissue. *Experiment in plant physiology*, Van, Nostrand. Reinhold Camp., New York, 1971; pp.16.
- Goszczynska, D.M. and Rudnicki, R.M.. Storage of cut flowers. *Hort. Rev.* 1988; **10**: 35-62.
- Grover, J. K.; Singh, K. and Kumar, A. Effect of passive hypobaric pressure on storage of gladiolus spikes. *Indian J. pl. physiol.* 2008; **13**(3): 238-242.
- Halevy, A. H. and Mayak, S. Senescence and postharvest physiology of cut flowers. *Part I. Hort. Rev.* 1979; **1**: 204-236.
- Halevy, A. H. and Mayak, S. Senescence and post-harvest physiology of cut flowers. *Hort. Rev.*, 1981; **3**: 59- 143.

12. Ho, L. C. and Nicholus, R. The role of phloem transport in the translocation of sucrose along the stem of carnation cut flowers. *Ann. Bot., (London)*, 1975; **39**: 439-446.
13. Hutchinson, M. J.; Chebet, D. K. and Emongor V. E. Effect of accel, sucrose and silver thio sulphate on the water relation and postharvest physiology of cut tuberose cut flower. *African crop sci. j.* 2003; **11**(4): 279 -287.
14. Ichimura, K. and Suto, K. Effects of the time of sucrose treatment of vase life, soluble carbohydrate concentrations and ethylene production in cut sweet pea flowers. *Pl. growth regulation*, 1999; **28**: 117-122.
15. Jature, S. D.; Nawaghare, P.D.; Shinde, S. J.; Naik D. M. and Borade, R. S. Effect of chemicals on vase life of cut tuberose (*polianthes tuberosa* L.) cv. Single *The Asian J. of Hort.* 2010; **4** (2) 478-480.
16. Jeenbuntug, J.; Buanong, M. and Kanlayanarat S. Study of sucrose pulsing treatment on physiological changes of tuberose (*Polianthes tuberosa*) after harvest. *Acta. Horticulture.* 2007; **775**: 425-428 .
17. Mayak, S. and Halevy, A. S. The action of kinetin in improving water balance and delaying senescence process of cut rose flowers. *Pl. Physiol.*, 1974; **50**: 341-346.
18. Machnish A. J; Jiang C. Z and Reid M. S. Treatment with thidiazuron improves opening and vase life of iris flowers. *Postharvest boil. technol.* , 2010; **56**: 77-84.
19. Meir, S.; Philosoph, H.D.; Michaeli, R.; Davidson, H.; Fogolman, M.; Fjeld, T. and Stomme, E., Improvement of keeping quality of mini gladiolus spikes during prolong storage by sucrose pulsing and modified atmosphere packaging. *Acta. Horticulture.*, 1995; **405**: 335-342.
20. Mok, M.C.; Mok, D.W.S.; Armstrong, D.J., Shudo, K.; Isogai, Y. and Okamoto, T. Cytokinin activity of N-phenyl-N'-1,2,3- thiazol-5-ylurea (Thaidizuron). *Phyokemistry.*, 1982; **21**: 1509-1511.
21. Mok, M. C.; Martin, R. C. Mok, and D. V. S. Cytokinins: biosynthesis, metabolism and perception. *In vitro cell., Dev. Biol. Pl.*, 2000; **36**: 102-107.
22. Murthy, B. N. S.; Murch, S.J. and Saxena, P. K. Thidiazuron: a potent regulator of in vitro plant marphogenesis. *In vitro. Cell . Dev. Biol. Plant.*, 1998; **34**: 267-275.
23. Panse, V. G. and Sukhatme, P. V. Statistical Methods for Agricultural Workers. ICAR, New Delhi, India 1967.
24. Salunkhe, D. K.; Bhatt, N. R. and Desai, D. B. Post-harvest biotechnology of flowers and ornamentals plants. *Springer verlag, Berlin*1990. pp. 192.
25. Sankhla, N., Mackay, W. A. and Davis, T. D Reduction of flower abscission and leaf senescence in cut phlox inflorescence by thiadizuron. *Acta. Hort.*, 2003; **628**: 837-841.
26. Sankhla, N., Mackay, W. A. and Davis, T. D. Effect of thiadizuron on senescence of flower in cut inflorescence of lupinnus densiflorus Benth. *Acta. Hort.*, 2005; **669**: 239-243.
27. Sen, C. K. and Packer, L. Thiol homeostasis and supplementation in physical exercise. *Amer. J. Chin. Nutri.*, 2000; **72**: 653-658.
28. Sexton, R. The role of ethylene and auxin in abscission. *Acta. Hort.*, 1998; **463**: 55-57.
29. Singh, V. P. and Jegadheesan, A. Effect of á-lipoic acid on senescence in Gladiolus flowers. *Indian J. Pl. Physiol.*, 2003; **516**-521.
30. Singh, A.; Kumar, J. and Kumar, P. Effects of plant growth regulators and sucrose on post-harvest physiology, membrane stability and vase life of cut spikes of gladiolus. *Pl. Growth Regul.*, 2008; **55** : 221-229.
31. Singh, A.; Dhaduk, B. K. and Ahlawat, T.. Storage of jasmine (*Jasminum sumbac*) in passive MAP. *Acta. Horticulture.* 2009; **847**: 321-326.
32. Singh, S.B. and Punitha, P. Entrepreneurship development through Anthurium Flower - A case study of Mizoram, North-East India. *Indian Res. J. Ext. Edu.*, 2012; **12** (3).
33. Spikman, G. The effect of water stress on ethylene production and ethylene sensitivity of fressia inflorescence. *Acta. Horticulture.* 1986; **181**: 135-140.
34. Torre, S.; Borochoy, A. and Halevy, A. H.. Calcium regulation of senescence in rose petals. *Physiologia. Plantarum.*, 1999; **107**: 214-219.
35. Van der Meuler-Muisers, J. J. M.; Van Overen, J. C.; Van der Plas, L. H. W and Van Tuyl, J. M.. Post-harvest flower development in Asiatic hybrid lilies as related to petal carbohydrate status. *Postharvest Biol. Technol.*, 2001; **21**: 201-211.
36. Varu, D. K. and A.V. Barad. Effect of different packing methods on vase life and quality of cut flowers in tuberose (*Polianthes tuberosa* L.) cv. Double. *Asian J. Bio Sci.*, 2008; **3** (1): 159-162.
37. Waithaka, K.; Reid, M. S. and Dodge, L. Cold storage and flower keeping quality of cut tuberose (*Polianthes tuberosa* L.). *J. Hort. Sci. Biotech.*, 2001; **76** (3): 271-275. .