Relationship Between Aflatoxin B1 Exposure and Etiology of Liver Disease in Saudi Arabian Patients

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Exposure to chronic low levels of aflatoxin B1 (AFB1) contamination can lead to immune suppression and nutritional consequences that might greatly contributed in the increase of hepatocellular carcinoma (HCC). The toxicity of AFB1 is greatly vary between different population, affected by age, gender, and environmental factors. Aflatoxin B1 (AFB1) was measured in 50 blood samples collected from non B, C hepatitis viruses and non CMV-Ab liver disease patients from different general hospitals and polyclinic in KSA during period 01-2013 to 06-2014. All Patients demonstrate elevation of ALT and AST with unknown etiology. Serum samples were obtained and kept at "20°C for AFB1detection. Out of the 50 blood samples, 38 demonstrate a detectable serum level of AFB1 while the remaining 12 patients were AFB1 negative. ALT was significantly higher in AFB1 positive samples compared to control AFB1negative. AFB1 was positively correlated with AST and ALT as liver function enzymes and with age as a risk factor of long duration of AFB1 chronic exposure. Multiple linear regression analysis ascertained the association between AFB1 chronic exposure and ALT increase in liver dysfunction Saudi patients. Measurement of elevated ALT as marker of liver injury in AFB1 chronically exposed Saudi patients can help to avoid the future development of HCC. Moreover, early detection of AFB1 exposure, together with early vaccination against HBV and HCV can remove the synergistic effects of these two etiological factors and thus decrease the risk of developing liver cancer.

Keywords: Aflatoxin B1, Transaminases, Hepatocellular carcinoma, Saudi Patients, liver dysfunction.

Aflatoxins is a very potent hepatotoxic and carcinogenic mycotoxin which is frequently reported as a food contaminant. AFB1 are probably the best known toxic metabolites produced by certain fungi usually developed on food due to bad storage conditions. AFB1 chronic exposure demonstrate genotoxic, mutagenic, immunosuppressive, and

remarkable increase of hepatocellular carcinoma (HCC)^{1,2}. Based on the fact that AFB1 recorded the most potent carcinogenic effects in both humans and animals, they have received greater attention compared to other mycotoxins. As it is realized that absolute safety is very difficult to be achieved, many developing countries among which is Saudi Arabia have attempted to decrease the toxicity of aflatoxins by regulations that control exposure that restrict the limits of these toxins in food and animal feed

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As aflatoxins are excreted in cow's milk, the exposure to these toxins has received a great health importance. It is well known that aflatoxin M1, as a metabolite of aflatoxin B1, is found in the milk of dairy cattle that have ingested feed contaminated with the AFB1. Hence, milk and dairy products can be targeted to decrease the exposure to AFB1 toxicity^{3,4}.

Moreover AFB1 contamination demonstrates synergistic interaction with hepatitis B virus infection which usually lead to the development of human hepatocellular carcinoma5. The human co-exposure to both AFB1 and hepatitis B was found to enhance early pre-neoplastic lesions in rat liver⁶. Up to this information, the risk of developing HCC was found to be 30 times higher in hepatitis B and C virus chronic infection^{4,7}. Understanding the mechanism through which HCC is related to AFB1 exposure can help to control the development of liver cancer9. This information initiate our interest to study the relationship between AFB1 chronic exposure, elevation of transaminases as measure of liver injury and age as risk factors to develop HCC in non B, C hepatitis viruses and non CMV-Ab Saudi patients. This might help to decrease the development of HCC in AFB1 -exposed individuals through early vaccination against HBV and HCV as synergistic risk factors in case of AFB1 chronic exposure.

MATERIAL AND METHODS

Samples collection

50 Blood samples were collected from liver disease patients have elevation in ALT and AST with unknown etiology; non B, C hepatitis viruses and non-cytomegalovirus- antibody (CMV-Ab) from different general hospitals and polyclinic in KSA during period 01-2013 to 06-2014. Serum samples were prepared from the blood and kept at "20 °C for aflatoxin detection.

Laboratory investigations

The patients non B, C hepatitis viruses and non-CMV-Ab and have elevation in liver profiles; were investigated with unHBsAg was carried by hemagglutination assay (Murex Diagnostics Limited, Dartford, UK) with radioimmunoassay testing of negative samples (Sorin Biomedica Diagnostics, Vercelli, Italy). anti-HCV and CMV-Ab were determined by third-generation enzyme-

linked immunosorbent assay (ELISA; ORTHO Clinical Diagnostics, Neckargemund, Germany). Liver profiles ALT and AST detected by Kinetic biochemical testing (Promega, USA).

Detection of aflatoxin concentration

Aflatoxins were extracted from serum samples by using the methods of Wray and Hayes 10 . Where identification and quantification aflatoxins B1 by using HPLC (Beckman Model 110A) method of Tchana $\it et~al~(11)$. The HPLC column used was an ODS-Sil-X-10 μm reverse phase column (4 \times 250 mm, Perkin Elmer). Serum samples were injected (25 μL) through a Perisorb RP 18, 10"40 μm pre-column (4.9 \times 50 mm, Waters) at 2,000 psi pressure. The mobile phase used was bi-distilled water-acetonitrile-methanol (16:5:5 v/v/v), at a rate of 1 mL .min $^{-1}$. Aflatoxin was detected using a Beckman fluorescent detector (Type 157) set at excitation wavelength of 365 nm and at emission 430 nm.

Confirmed ELISA assay of aflatoxin B1

The presence of AFB1 in serum samples were confirmed by using an immuno- enzymatic commercial kit based on monoclonal antibodies specific for it (Transia, Lyon France). The extract was evaporated to dryness and taken up in the sample buffer provided in the kit, then 50 µL of the serum sample was pipetted into duplicate wells of an ELISA plate which sensitised with the monoclonal antibody specific for aflatoxin AFB1. After incubation and washing 50 µL of the specific monoclonal antibody conjugated to horseradish peroxidase dissolved in the conjugate buffer provided in the kit was added. Incubation followed by washingwere done before the addition of the substrate. The intensity of the developed color was read using ELISA reader.

Statistical analysis

The data were analyzed using the Statistical Package for the Social Sciences (SPSS, Chicago, IL, USA). The results were expressed as mean± standard deviation of the mean (SD). All statistical comparisons between the 50 blood samples from liver disease individuals (elevation in ALT and AST with unknown etiology, N=12 (negative for AFB1, used as control); non B, C hepatitis viruses and non-cytomegalovirus- antibody, CMV-Abs, and AFB1 positive individuals and used as patients; N=38) were performed using the one-way analysis of variance (ANOVA) Pearson's correlations were

performed between the measured parameters. A p value less than 0.05 was statistically considered significant

RESULTS

Table 1 demonstrates levels of AST, ALT and AFB1 in sera of 50 liver disease patients

with unknown etiology (i.e non B, C hepatitis viruses and non CMV-Ab). Out of the 50 samples, 38/50 had a detectable serum level of AFB1 with concentrations varies from 0.5-1.675µg/L and 12/50 were negative and used as control participants. While AST was non-significantly different in AFB1 exposed patients, ALT was

Table 1. The independent T-Test between Control and Patients groups in all ALT (μ/L) and AST (μ/L)

Parameter	Group	N	Min.	Max.	Mean \pm S.D.	P value
ALT (μ/L)	Control Patients	12 38	56.05 56.45	67.74 115.30	59.65 ± 3.39 68.10 ± 14.69	0.002
AST (μ/L)	Control Patients	12 38	35.23 35.25	62.01 72.32	43.32 ± 7.01 45.65 ± 8.76	0.405

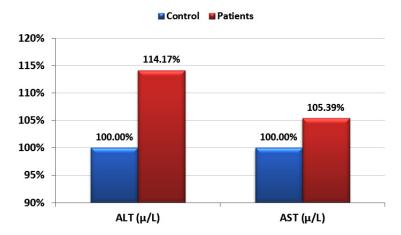


Fig. 1. Percentage change of ALT (μ/L) and AST (μ/L) in patients group compared to control group

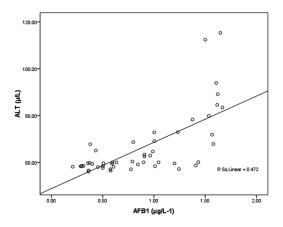


Fig. 2. Correlation between AFB1 (μ g/L-1) and ALT (μ /L) with best fit line curve (positive correlation)

significantly higher in AFB1 positive samples compared to control AFB1-negative.

Figure 1 demonstrates the percentage change in AST and ALT in AFB1 positive compared to control. Tables 2 and 3 demonstrate the correlation between AFB1 exposure and AST, ALT and age. It can be easily noticed that AFB1 was positively correlated with AST and ALT as liver function enzymes and with age as a risk factor of long duration of AFB1 exposure.

Table 4 shows multiple linear regression analysis using ALT as dependent variable and AFB1 and age as independent variables or predictors. R² value of 0.461 shows that 46.1% of

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the change in ALT is associated or related to the elevated AFB1.

DISCUSSION

Aflatoxins is considered the most agent naturally causing hepatocarcinogens; and the impact of aflatoxin on public health is detected in both acute and chronic exposure (10, 27). Acute exposure of aflatoxin (aflatoxicosis) can result fulminant liver failure (11, 28); and chronic exposure of aflatoxin has been linked to development of HCC (12, 27). In this research include difficulties in detection clinical outcomes in some resource-constrained environments

Table 2. Correlations between AFB1, AST, and ALT

Parameters	R (Person Correlation)	Sig.	
$\overline{ AFB1 (\mu g/L-1) \sim ALT (\mu/L) } $ $AFB1 (\mu g/L-1) \sim AST (\mu/L)$		0.001 0.001	Pa Pa

^{**} Correlation is significant at the 0.01 level.

and difficulty in accurately detecting aflatoxin exposure.

Data presented in table 1 demonstrate that AFB1 exposed patients has a significantly higher ALT enzyme compared to control. As ALT is a biomarker of liver dysfunction, this observation can help to suggest that a high level of the hepatotoxic AFB1 was involved in the developed liver injury. This is in good agreement with the previous reports of Barton *et al*¹² which recorded that interperitonial injection of AFB1 to rats demonstrated a dose-dependent hepatotoxicity measured as marked elevations in serum ALT and AST activities concomitant with histologic changes. As ALT exceeds AST in toxic hepatitis,

Table 3. Correlation between AFB1 and age

Parameters	R (Person Correlation)	Sig.	
AFB1 (μ g/L-1) ~ Age	0.619**	0.001	Pa

^{**} Correlation is significant at the 0.01 level.

Table 4. Multiple regression using stepwise method for ALT as a dependent variable

Predictor Variable	Beta	P value	Adjusted R square	Model F value P value		
AFB1	0.024	0.001	0.461	42.879	0.001	_

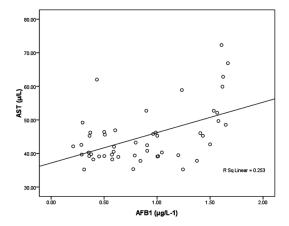


Fig. 3. Correlation between AFB1 (μ g/L-1) and AST (μ /L) with best fit line curve (positive correlation)

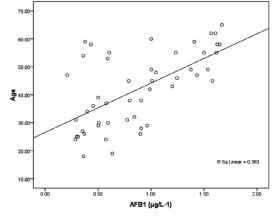


Fig. 4. Correlation between AFB1 (μg/L-1) and Age with best fit line curve (positive correlation)

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a Positive correlation.

^a Positive correlation.

viral hepatitis, and cholestatic hepatitis, the higher concentration of ALT compared to AST presented in Table 1, in AFB1 negative (control) and AFB1 positive (patients) ascertained that all investigated samples demonstrate liver injury¹³. This in turn can confirm the hepato-toxic effect of AFB1 as much higher increase in ALT compared to AFB1 negative participants. Use of ALT as predictive marker of AFB1 liver toxicity can find support in the recent work of Yang et al. (14) in which the elevated level of serum ALT was one of the cascade three steps model of liver injury induced by total saponins of plant origin. On the other hand, the non-significant elevation of AST in response to AFB1 toxicity, reported in the present study, is not in good agreement with the previous study of Allameh et al.15 which showed that serum lactate dehydrogenase (LDH) and AST demonstrate more dependency on the exposure duration and concentration of aflatoxins.

Tables 2 together with figures 2 and 3 demonstrate Pearson's correlations between ALT, AST, age and AFB1 as the four measured variables in this study It can be easily noticed that AFB1 was positively correlated (PÂ0.001) with AST and ALT. This can suggest the importance of both enzymes as markers of AFB1 hepatotoxicity. The positive correlation between AFB1 and age (Table 3 and figure 4) in AFB1 positive samples can prove that detection limit of AFB1 fall within certain limits which need longer period to be reached (i.e older participants). This suggestion can find support in the recent work of Rieswijk et al¹⁶ which proves for the first time that AFB1 induced an epigenetic persistent toxic effect eventually leads to HCC. They were able to identify methyl DNA-mRNAinteractions as marker for a persistent epigenetic effect associated with the early onset of AFB1induced HCC.

The general purpose of multiple regressions is to learn more about the relationship between several independent or predictor variables (AFB1 and age) in the present study, and a dependent or criterion variable, ALT through the recorded R^2 and β coefficient values. In this analysis R^2 describes the proportion or percentage of variance in the dependent variable explained by the variance in the independent variables together which sometimes called the predictor variables.

Table 4 demonstrates the contribution of AFB1 as predictor variable in the elevation of ALT as dependent variable. This suggestion can find support in the previous study of Wild *et al*¹⁷ which proved that increased liver injury may modulate AFB1 metabolism. The relationship of ALT elevation and AFB1 and vice versa can also be supported through considering the early work of Thaxton and Hamilton¹⁸ and Gawai *et al*¹⁹ in which the proved the elevation of ALT, as indication of acute liver injury, in birds that had been given a single dose of aflatoxin compared to control untreated birds.

The reported increase of ALT in AFB1 positive patients can also find support in more recent studies which demonstrate the significant increase of ALT among other biochemical parameters in rats fed AFB120-22. Therefore this study exhibits similarity to the previously research which prove that AFB1-induced liver injury can be attributed to the oxidative stress. Based on this similarity regarding the elevation of ALT in Saudi patients with liver dysfunction, as subject of the present study, curcumin and green tea as two supplements show ameliorative effects of AFB1-induced liver energy can be used to avoid the carcinogenic effect of AFB1 as long as possible in Saudi populations who demonstrate AFB1 positivity together with elevated ALT.

In addition to these recommended supplementations, a reasonable and promising clinical intervention to reduce aflatoxin-related HCC is early vaccination against HBV and HCV. This can be supported through the scientific fact that early vaccination of children against HBV has, over the past 30 years, significantly decreased HBV infection in several regions, including Europe^{23,24}, Taiwan²⁵ and Thailand²⁶. This might help to control the carcinogenic effect of aflatoxin, because removing the synergistic impact between HBV and aflatoxin exposure would remarkably reduce HCC risk^{27, 28, 29}.

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