Production and Characterisation of Cellulase by Streptobacillus species- APS-8

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(Received: 12 September 2007; accepted: 17 October 2007)

A cellulolytic bacterial species APS-8 was isolated from the soils of Visakhapatnam region. This strain is characterized as Gram positive *Streptobacillus* species. The strain grew well in medium containing Carboxymethylcellulose (CMC), producing maximum enzyme at 37°C and pH 8.0. The activity of extra cellular enzyme was found to be high at temperature 50°C and at pH 5.0. Metal ions like MgSO₄, ZnSO₄ and FeCl₃ enhanced the enzyme activity whereas MnSO₄ inhibited the enzyme activity under same assay conditions.

Keywords: Cellulase, Congo red, Carboxymethylcellulose (CMC), Carboxymethylcellulase (CMCase) assay, APS-8.

Life on Earth depends on photosynthesis, which results in production of plant biomass having cellulose as the major component¹. Cellulose is found in nature almost exclusively in plant cell walls, although it is produced by some animals (e.g., tunicates) and a few bacteria¹⁻². Like starch, cellulose is composed of glucose units; whereas all the glucose molecules in a starch chain share the same orientation ("alpha linkages"), adjacent constituents of a cellulose chain are flipped 180 degrees ("beta linkages"). Due to this molecular arrangement cellulose is tough and fibrous and difficult to break down3. Cellulase was identified as one of the key enzyme degrading cellulose⁴⁻⁵ Cellulase refers to a family of enzymes, which act in concert to hydrolyze cellulose⁶. It is produced chiefly by fungi like Aspergillus,

*Trichoderma, Cladosporiu*⁷ and bacteria like *Clostridium, Ruminococcus, Cellulomonas*¹. Enzymes have been used since the dawn of mankind in various industrial processes. In this regard cellulases have profound applications. In textile industries it is used for keeping colour brightness of the fabrics for longer time and in stone washing of jeans to make them appear faded in very less time⁸⁻¹⁰. Cellulases are used in the Processing of animal feed, coffee beans and in clarification of fruit juice⁸. They are also used to bleach the wood pulp and to recycle the paper¹¹⁻¹².

One of the most recent developments for the use of cellulases has been the production of ethanol for fuel under the trade name Eoethanol¹³. Screening for cellulolytic micro organisms has been restricted for a long time mainly for molds. In this investigation, a new different cellulolytic bacterium that might be suited for any of such a process was isolated and the characterization of enzyme was done using different parameters.

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MATERIAL AND METHODS

All chemicals used were of analytical grade and obtained from Merck, Glaxo, Qualigens (Mumbai, India). Culture media was obtained from Hi Media (Mumbai, India).

Composition of Congo red agar medium (gm/ 100ml)

The Congo red agar medium contains Gelatin-200mg, Carboxymethyl Cellulose-188 mg, K_2 HPO₄- 50mg, MgSO₄- 25mg, Congo red -20 mg, Agar -2g, the pH was adjusted to 7.0.

Isolation of Cellulolytic Bacterial Species

Soil samples were collected from a depth of 5cm from the moist part underneath a stack of decaying hay into a sterile Ziploc bag and serial dilution was carried out. Each dilution (100μ I) was plated separately onto Congo red agar medium and incubated at 37° C for 3 to 7 days¹⁴⁻¹⁹. After 3-7 days of incubation the cellulolytic organism was identified by a yellow halo against a red background (Fig. 1). The colonies with zone of clearance were selected and plated separately on to a fresh medium to obtain a pure culture²⁰. A bacterial strain with large zone of hydrolysis was selected and designated as APS-8.

Cellulase Assay

The enzyme assay was performed according to the Method²¹. An aliquot of 0.5 ml of supernatant was taken in a test tube and mixed with 0.5 ml of 1% CMC in 0.1 M phosphate buffer (pH 6.7) and incubated at 50°C for 30 min and the liberated reducing sugar was determined by DNS method ²² and the Cellulase activity is expressed as micro moles of glucose liberated per minute per ml of the culture filtrate²³.

Enzyme production

Production of cellulose is carried out by inoculating $(1X10^8 \text{ Cells/mL})$ overnight grown APS-8 culture into a 500 ml flask containing 100ml of production media and incubated at 37° C at 150 rpm in orbital shaker (Remi-Ris-24, Mumbai, India) for 7 days. The production medium contained Gelatin-200mg, CMC- 188 mg, K₂HPO₄ -50mg, MgSO₄ -25mg, in 100ml of distilled water, the pH was adjusted to 7.0 with 1.0N NaOH. The growth and the cellulase production were determined for every 24hrs. Enzyme production was tested by CMCase method and the cell density by measuring the optical density at 660nm respectively. At the end of each fermentation period, the whole fermentation broth was centrifuged at 10,000g for 15 min (Plastocraft Super Spin- RV/FM High speed, Mumbai, India) and the clear supernatant was used as crude enzyme.

Effect of different parameters such as temperature ($20^{\circ}C-90^{\circ}C$), pH (2.0-9.0) and varying salt concentrations (1.0mM-5.0mM) on the production of cellulase by APS-8 were also studied.

Characterization of enzyme Effect of pH on enzyme activity

The activity of cellulase was tested over a pH range from 2.0-12.0 under standard assay conditions using appropriate buffers (0.1M Acetate pH 2.0, Citrate pH 3.0-5.0), Phosphate pH 6.0-7.0, Tris-HCl pH8.0-9.0, Glycine-NaOH pH 10.0 and Carbonate- Bicarbonate pH11.0-12.0).

Effect of temperature on enzyme activity

The effect of temperature on enzyme activity was studied by performing the standard CMCase assay procedure over a temperature range of 20°C-80°C.

Effect of metal ions on enzyme activity

The effect of different metal ions on cellulase activity was studied by using 0.2 M concentration of metals such as Na⁺, Fe⁺³, Mg⁺², Mn^{+2} Co⁺ and Zn⁺².

Table 1. Morphological characterization of the APS-8

Colony Morphology	
Configuration	Round
Margin	Entire
Surface	Smooth and shiny
Elevation	Raised
Opacity	Opaque
TEST	APS-8
Gram's Reaction	+ve
Cell Shape	Rods
Size(µm)	2.3-3.5µ in length
Arrangements	In chains
Spores	Absent
Motility	Non motile







Fig. 4. Effect of varying salt concentration on cellulase production



Fig. 5. Effect of pH on cellulase production

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Fig. 8. Effect on metal ions on cellulase activity

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enzyme activity significantly. Wher as, $ZnSO_4$ and $FeCl_3$ found to enhance the enzyme activity moderately. However, $MnSO_4$ inhibited the activity by 50%.

ACKNOWLEDGEMENTS

The authors thank Management of the GITAM University for providing necessary facilities in the Department of Biotechnology for carrying out this work.

REFERENCES

- Lee R. Lynd, Paul J. Weimer, Willem H. van Zyl, and Isak S. Pretorius, Microbial cellulose utilization: Fundamentals and Biotechnology. *Microbiology and Molecular Biology Reviews*: 2002; 506–577.
- Lynd, L. R., Wyman C. E., and Gerngross T. U., Biocommodity engineering. *Biotechnol. Prog.* 1999; 15:777–793.
- 3. Environmental and Energy Study Institute (EESI), January 8, 2006, www.eesi.org
- Kotchoni O.S., Shonukan O.O., and Gachomo W.E., *Bacillus pumilus* BPCRI 6, a promising candidate for cellulase production under conditions catabolite repression. *Afr. J. Biotechnol.* 2003; 2(6): 140-146.
- Muthuvelayudham, R., and Viruthagiri, T. Fermentative production and kinetics of cellulose protein on *Trichoderma reesei* using sugarcane bagasse and rice straw. *African Journal of Biotechnology*. 2006; 5(20): 1873-1881.
- 6. Deiveegan, S., Muthuvelayudham, R., and Viruthagiri, T. Triggering cellulase protein production using cellulose with lactose by *Trichoderma reesei. Asian Jr. of Microbiol. Biotech.Env.Sc.* 2006; **8**(2) :249-251.
- Carlile M. J., and Watkinson S. C., The fungi. Academic Press, New York, N.Y. 1997; 269–275.
- Bhat M.K., Cellulases and related enzymes in biotechnology. *Biotechnology Advances*. 2000; 18: 355-383
- Haki, G.D.; Rakshit, S.K., Developments in industrially important thermostable enzymes: a review. *Bioresource Review*. 2000; 89: 17-34.
- Csiszar E., Losonczi A, Szakacs G, Rusznak I, Bezur L., Reicher J, Enzymes and chelating agent in cotton pretreatment. *Journal of Biotechnology*. 2001; 89: 271-279.

- Pelach, M.A., Pastor, F.J., Puig, J., Vilaseca, F., and Mutje, P. Enzymic drinking of old newspapers with cellulase. *Process Biochemistry*. 2003; 38: 1063-1067.
- Dienes, D., Egyhazi, A., and Reczey, K. Treatment of recycled fiber with *Trichoderma* cellulases. *Industrial Crops and Products* 2004; (article in press).
- Iogen doubles EcoEthanol Capacity. April 28, 2003. www.iogen.ca/news/28_03_2003.html (accessed May 17,2003).
- Gitaitis, R. D., Chang, C. J., Sijak, K., and Dowler, C. C. A differential medium for semiselective isolation of *Xanthomonas campestris* pv. *Vesticatoria* and other cellulytic xanthomonads from natural sources. *Plant Dis.* 1991; **75**: 1274–1278.
- Kluepfel, D. Screening of prokaryotes for cellulose and hemicellulose degrading enzymes. *Methods Enzymol.* 1988; 160:180–18.
- Mateos, P. F., Jimenez-Zurdo, J. I., Chen, J., Squartini, A. S., Haack, S. K., Martinez-Molina, E., Hubbell, D. H., and Dazzo, F. B. Cell-associated pectinolytic and cellulolytic enzymes in *Rhizobium leguminosarum* biovar trifolii. *Appl. Environ. Microbiol.* 1992; 58:1816–1822.
- Charles W. Hendricks, Jack D. Doyle and Bonnie Hugley., A New Solid Medium for Enumerating Cellulose Utilizing Bacteria in Soil. *Applied and environmental microbiology*. 2005; 2016–2019
- Wood, P. J., Erfle, J. D., and Teather, R. M. Use of complex formation between Congo red and polysaccharides in detection and assay of polysaccharide hydrolases. *Methods Enzymol.* 1988; 160: 59–74.
- Teather, R. M., and P. J. Wood. Use of Congo red-polysaccharide interactions in enumeration and characterization of cellulolytic bacteria from the bovine rumen. *Appl. Environ. Microbiol.* 1982; 43:777–780.
- Aslim B., Yuksekdag Z.N., and Beytli Y, Determination of PHB growth quantities of Certain Bacillus Species Isolated from Soil. Turkish Electronic. J. Biotechnol. Special issue. 2002; 24-32
- 21. Mandels, M and Sternberg, D. J. Ferment. Technol. 1976; 54:267
- 22. Miller, G.L. Use of Dinitrosalicylic Acid Reagent for Determination of Reducing Sugar. *Anal.chem.* 1959; **31**:426.
- Debabrat Baishya and Manab Deka. Asian Jr. of Microbiol. Biotech. Env.Sc. 2006; 8(2): 345-348.

J. Pure & Appl. Micro., 1(2), Oct. 2007