

Pathogenicity of EUS Associated *Aeromonas hydrophila*, Artificially Injected to *Channa striatus* (Bloch)

S.A. Mastan

P.G. Department of Biotechnology, D.N.R College,
PG Courses and Research Centre, Bhimavaram- 534 202, India.

(Received: 14 May 2008; accepted: 21 August 2008)

Experiments have been conducted with *A. hydrophila*, which was isolated from EUS affected *Mystus cavasius* and injected to *Channa striatus* intraperitoneally and observations were made at regular intervals for 96hrs to study the disease progress. Visible clinical and behavioural symptoms were noticed. Moribund and dead fish were autopsied to observe pathological changes in different organs. The L_{p50} value obtained was 2.98×10^3 cells/ml. The statistical analysis such as regression equation, chi-square, $\text{Log}L_{p50} \pm$ standard deviation (SD) and fiducial limits (FL) of the data were recorded as $Y = -3.070 + 2.322x$, 4.027 , 3.4742 ± 0.0651 , 2.22×10^3 and 4.00×10^3 cells/ml, respectively. The chi-square value was found to be significant at $P \leq 0.05$.

Key words : Pathogenicity, EUS *Aeromonas hydrophila*, *Channa striatus*.

Epizootic Ulcerative Syndrome (EUS) is a devastating disease affecting many species of wild and cultured freshwater fishes in Asia (Chinabut, 1998). In India also, the EUS has assumed the shape of epizootic proportion and has taken heavy toll of millions of fry, fingerling and adult fishes. The first outbreak of EUS was reported from India in the month of May, 1988 from Northeastern region. Thereafter, it spread to Northern and Southern states of the country (Das, 1988; Jhingran and Das, 1990; Pal and Pradhan, 1990;

Dastidar and Chakraborty, 1992; Das and Das, 1993; Mohan and Shankar, 1994; Qureshi *et al.*, 1995; Mukherjee, 1996; Lio-po *et al.*, 1996; 1998; Mastan, 1998 and Mastan and Qureshi, 1999; 2001; 2001; 2001; 2003; 2004; 2005). EUS is reported to affect over thirty freshwater species of cultured as well as other wild fish population, although it was most notable in Snakehead and Catfishes (Tonguthai, 1985). Wild fish species have been found to be seriously affected by the outbreaks resulting in severe dermal ulceration and large-scale mortality.

Unfortunately, the problem has become complex due to the fact that many vital clues regarding the causative organisms and other factors responsible for EUS outbreaks are yet to be unveiled and this problem is still evading a plausible solution, despite global efforts (Jhingran and Das, 1990; Das and Das, 1993 and Mastan, 1998). *Aeromonas hydrophila* is consistently associated with EUS affected fish in all the south

* To whom all correspondence should be addressed.

east-Asian countries. Thus, the present study is aimed to test the pathogenicity of *A. hydrophila* against *C. striatus* in relation to EUS.

Materials and methods

For the purpose of present study, healthy and disease free fingerlings of *Channa striatus* averaging 12.4 ± 0.28 gm weight and 10.40 ± 0.12 cm in length were used. They were introduced in aquaria of the size of 90 x 45 x 45 cm washed with $KMnO_4$ filled with 100 litres of freshwater each. In each concentration, twenty fishes were used which were acclimatized in laboratory conditions for one week, prior to the experiment. They were fed with pelleted diet once a day approximately 75% of the water was changed everyday Feeding was stopped 24hrs prior to experiment.

Bacterial suspension was prepared by culturing *A. hydrophila* on TSA plates at 30°C temperature for 24hrs and harvesting them with 500ml of 0.85% physiological saline. Colony forming unit was then determined by plating 10 fold dilution series. Later, it was diluted with distilled water to make a cell number in the powers of 10^3 /ml and used for exposure to fish. The counting of cells were done with the help of haemocytometer. Bacterial strain used in this study was originally isolated from naturally EUS affected fishes. Symptoms and mortality was recorded upto 96hrs after injection. Each experiment was carried in triplicate sets.

RESULTS AND DISCUSSION

Fingerlings of *Channa striatus*, experimentally injected with the challenge organism, *A. hydrophila*, revealed the following behavioural and clinical symptoms. The fishes were inactive and were lying at the bottom showing loss of movement. Sometimes, they swam in circling manner, sometimes upside-down and sometimes in flating position. Some fishes were seen lying down on the bottom of aquaria before death.

Other symptoms shown by them were relative more mucus secretion and the appearance of haemorrhages on the general surface of body. Haemorrhages were seen in all the fishes within 18hrs after injection. Superficial ulcers developed at the point of injection. In some fishes scales got

Table 1. Showing the results of experimental infection produced in *Channa striatus* with *Aeromonas hydrophila*, isolated from EUS affected fishes

S. No.	Conc. in (cells/ml)	Symptoms observed	Mortality	Regression equation (Y=a+bx)	Chi-square ($X^2(n-2)$)	L_{p50}	$\text{Log}L_{p50} \pm SD$	95% FL (cells/ml)
1.	1.40×10^3	-	30	$Y = -3.070 + 2.322x$	4.027*	2.98×10^3	3.4742 ± 0.0651	$LL: 2.22 \times 10^3$ $UL: 4.0 \times 10^3$
2	2.52×10^3	Haemorrhages	45					
3	3.50×10^3	Haemorrhages	65					
4	7.60×10^3	Haemorrhages	80					
5	Control	-	0					

FL = Fiducial limit; LL : Lower limit; UL : Upper limit

*Value is significant P \leq 0.05

raised and became loose while in others they fell down. Severity of symptoms increased with the increase in bacterial concentration.

Table 1 exhibits results of statistical analysis of data for pathogenicity of *A. hydrophila* against fingerlings of *Channa striatus*. When four different concentrations of *a. hydrophila* suspension, ranging from 1.4×10^3 to 7.6×10^3 cells/ml were inoculated to *Channa striatus*, they caused 30 to 80% mortality within 96hrs after injection. The L_{D50} value obtained was 2.98×10^3 cells/ml. the statistical analysis such as regression equation, chi-square, Log L_{D50} Standard deviation (SD) and fiducial limits (FL) of the data were recorded as $Y = -3.070 + 2.322 x$, 4.027, 3.4742 ± 0.0651 , 2.22×10^3 and 4.00×10^3 respectively. The chi-square value was found to be significant at $P \leq 0.05$.

Though, the fish and the pathogens live together in an aquatic environment, the disease manifestation occurs only when the susceptible fish and the virulent pathogen happen to meet under certain favourable environment conditions. In earlier reports of bacteria associated with fish diseases, the workers have made little efforts to differentiate between primary and secondary role of bacteria. In the present investigation, experimental infection trial were conducted with *A. hydrophila*, which was isolated from naturally EUS affected fishes. It was inoculated intraperitoneally in the fingerlings of *C. striatus*. The pathogenicity tests showed that *A. hydrophila* was pathogenic to fish. This observation is in agreement with findings of Pradhan and Pal (1990), Ali and Tamuli (1990), Singh *et al.*, (1991), Lio-Po *et al.*, (1992) and Karunasagar (1995). Pal and Pradhan (1990) have reported that *A. hydrophila* and *P. fluorescens* could kill the host fish more rapidly than other species of bacteria. *A. hydrophila* is found to be the most destructive pathogen causing infection even within eighteen hours after injection. This finding conforms with the work of Shankar (1995) who reported that the infection caused by *A. hydrophila* in *Cyprinus carpio* resulted in mass mortality. It has been observed that inoculation of *A. hydrophila* produced symptoms of EUS such as the appearance of small raised haemorrhagic areas on the body surface, erratic swimming movements, staying of fish near the surface layer

of water with head projecting out of it, lethargic condition and non-feeding tendency. In the present investigation, L_{D50} value was recorded at 2.98×10^3 cell/ml. the recorded fiducial lower and upper limits are 2.22×10^3 and 4.0×10^3 cells/ml, respectively. Chi-square value is found significant in experimental fishes as compared to control ones. The results of the present investigation reveals that *A. hydrophila* may be considered as potential pathogen for producing EUS in fishes.

REFERENCES

1. Ali, A and Tamuli, K., Isolation of an aetiological agent of Epizootic Ulcerative Syndrome. *Fishing Chimes*, 1990; 43-46.
2. Banu, A.N.H, Ialam, M.A, Chowdhary, M.B.R and Chandra K.J., Pathogenicity of *Aeromonas hydrophila* artificially infected to Indian major carp (*Cirrhinus mrigala* Ham). *Bangladesh J. Fish Res.*, 1999; **13**: 187-192.
3. Jhingran, A.G and Das, M.K, Epizootic Ulcerative Syndrome in fishes. Bull No. 65, CICERI, Barrackpore, 1990; 14.
4. Karunasagar, I., Sugumar, G and Karunasagar, I., Virulence characteristics of *Aeromonas* spp. Isolated from EUS affected in Diseases in Asian Aquaculture II. M. Shariff, J.R. Arthur and R.P Subasinge (eds), Fish Health Section, Asian Fisheries Society, Manila 1995; 307-314.
5. Lio-Po, G.D., Albright, L.J. and Alapide-Tendencia, E.V., *Aeromonas hydrophila* in the epizootic ulcerative syndrome (EUS) of Snakehead, *Ophiocephalus striatus* and experimental induction of dermomuscular necrotic lesion. Diseases in Asian Aquaculture, Vol, 1 (eds M. Shariff, R. Subasinghe and J. Arthur). Asia Fisheries Society, Manila, Phillipines. 1992; 461-474.
6. Mastan, S. A., Studies on bacterial fish pathogens in relation to the Epizootic Ulcerative Syndrome in fishes. *Ph.D. Thesis*. Submitted to Barkatullah University, Bhopal. 1998; 1-200
7. Mastan, S.A and Qureshi T.A, Sensitivity of bacteria isolated from EUS affected fishes against various antibiotics. *J. Ecotoxicol environ Monit.*, 1999; **9**(2): 149-153.
8. Mastan, S.A. and Qureshi T.A, : Effect of different antibiotics on EUS affected fish, *Channa striatus* (Bloch). *Environ Eco.*, 2001; **19**(1): 56-59.
9. Mastan, S.A and Qureshi T.A, : Role of bacteria in the Epizootic Ulcerative Syndrome of fishes. *J. Environ Biol.*, 2001; **22**(3): 49-56.

10. Mastan, S.A and Qureshi T.A; : Bacteriological and Histopathological Investigations on EUS affected *Macrognathus aculeatus*. *Asian J. Micro. Biotech En. Sc*;2001; **3**(2): 38-43.
11. Mastan, S.A and Qureshi T.A; : Experimentally, induced Epizootic Ulcerative Syndrome in *Labeo rohita* (Ham). *Indian J. Anim. Health*; 2002; **40**(2): 108-112.
12. Mastan, S.A and Qureshi T.A; : Observations on histological alterations in various tissues of EUS affected fish, *Channa striatus* (Bloch) *J. environ Biol.*; 2003; **24**(3): 405-410.
13. Mastan, S.A and Qureshi T.A 2004 : Bacteriological and Histopathological investigations on EUS affected *Puntius sarana* (Ham). *Asian J. Mic Biot. En. Sc.*, 2004; **7**(1): 115-120.
14. Mastan, S.A and Qureshi T.A; : Pathogenicity of *Aeromonas hydrophila* artificially injected in *Cirrhinus mrigala* (Ham). *Asian J. Mic. Biot Env. Sc.*, 2005; **7**(3): 48-51.
15. Pal, J. and Pradhan, K; Bacterial involvement in ulcerative condition in air breathing fish from India. *J. Fish Biol.*, 1990; **37**: 833-839.
16. Pradhan, k. and Pal, J.; Experimental induction of ulcer in the fish *Channa punctatus* by bacteria *Environ and ecol.*,1990; **8**: 812-815.
17. Qureshi, T.A, Prasad, Y., Mastan, S.A and Chauhan, R;. Investigations on the involvement of fungal and bacterial pathogens in EUS of fishes. *Biotech. Consortium.*, 2000; 36-42.
18. Shakila, L., Sheeba, M., Omprakasm and Manohar, L.;. Bacterial haemorrhagic Septicemia in *Cirrhinus mrigala* infected experimentally with *Aeromonas hydrophila* . *J. Enmviron Biol.*, 1999; **20**(1): 55-59.
19. Shankar, K.M.;. Monoclonal antibodies for serotyping, diagnosis and antigen characterization of *Aeromonas hydrophila* a pathogen of carp in Indian Fish Farma Progree Report, 1995; 1-2.
20. Shariff, M. and Subasinghe, R.;. Experimental induction of Epizootic Ulcerative Sundrome EUS. Proceeding of ODA Regional Seminar on Epizootic Ulcerative Syndrome. AAHRI, Bangkok, 1994; 279.
21. Singh, I. S. B., Philip, R., Maqbool, T.K., Ramesh, S., Harikrishnan, P. and Menon, M. R., Microbial involvement in the ulcerative disease of fishes of Inland waters of Kerela. Seminar on fish disease in backwater of Kerela, 19 Nov, Dessey Sastra Vedi, Thiruvananthapuram, Kerela, 1991.
22. Subasinghe, R.; Experimental induction of epizootic ulcerative syndrome in the presence of skin parasites, Proceeding of ODA Regional Seminar on Epizootic Ulcerative Syndrome. AAHRI, Bangkok. 1994; 271.