

## Frequency of Isolated Bacteria and Fungi from Bloodstream of 3212 Patients Hospitalized at Milad Hospital, Tehran, Iran 2009-2010

Gholamreza Rafiei<sup>1</sup>, Mohammad Rahbar<sup>2\*</sup>,  
Abbas Farhadian<sup>3</sup> and Rana Amini<sup>2</sup>

<sup>1</sup>Department of Biology, College of Sciences, Shiraz University, Shiraz, Iran.

<sup>2</sup>Department of Microbiology, Iranian Reference Health laboratory, Tehran, Iran.

<sup>3</sup>Department of Bacteriology, School of Medical Science, Tarbiat Modares University, Tehran, Iran.

(Received: 28 October 2010; accepted: 18 December 2010)

Bloodstream infections (BSIs) are one of most significant cause of mortality and morbidity worldwide, even in community-acquired or hospital-acquired infections. Unfortunately, until now there is not any comprehensive study to survey the frequency of pathogens isolated from BSIs in Iran. The aim of this study was to determine epidemiological features of nosocomial BSIs in the Milad hospital, Tehran, Iran.

This study is a retrospective study on survey of 3212 blood cultures during 2009 to 2010. Antimicrobial susceptibility testing was performed by Kirby-Bauer disk diffusion method as recommended by Clinical Laboratory Standard Institute (CLSI). All analysis was performed using SPSS 16.

Our study revealed that 66% of BSIs were caused by gram-negative bacteria, while 24.1% of BSI caused by gram positive bacteria, the rest of them (9.9%) was infected by yeast. Among gram negative bacteria, *Enterobacter cloacae* (26.4%) and *Escherichia coli* (24%) were more prevalent organisms, while among gram positive bacteria *Staphylococcus aureus* (47.8%) and Coagulase negative *Staphylococci* (CoNS) (16.8%) were more prevalent. In fact, this study indicates the importance of establishment of a comprehensive program in Iran or even in Middle East upon the antimicrobial resistance rate.

**Key words:** Bloodstream, Bacteria, Fungi, Tehran.

---

Nosocomial infections are among infections that become clinically evident after 48 hours of hospitalization<sup>1</sup>. Even among community-acquired or hospital-acquired

infections, nosocomial infections bloodstream infections (BSIs) are one of most significant cause of morbidity and mortality worldwide<sup>1,2</sup>. Since 1990s there were many plans, such as Surveillance and Control of Pathogens of Epidemiological Importance (SCOPE) and National Nosocomial Infection Surveillance (NNIS), to investigate the nosocomial and non-nosocomial infections in progressive countries<sup>3-6</sup>. Albeit in some cases each

---

\* To whom all correspondence should be addressed.  
Tel: (+9821) 66728112-66728113-66728121-66723346-66760526 Fax: (+9821) 66728121  
E-mail: rahbar\_reflab@yahoo.com

pathogen caused BSI separately<sup>7-13</sup>.

Unfortunately, until now there have not been any comprehensive epidemiologically program to survey the frequency of pathogens isolated from BSIs, and their antimicrobial susceptibility in Iran, similar to what have been analyzed in other studies (14). Although in some hospitals there were reports about the bacterial frequency and antimicrobial susceptibility<sup>15</sup>.

At this retrospective study we aim to demonstrate the epidemiological features of community-acquired and nosocomial BSIs in the Milad hospital as one of most prominent clinical centers in Iran.

### MATERIAL AND METHODS

This study was designed as a retrospective survey of 3212 blood culture specimens prepared during 2009 to 2010. As we have not any control over the collection of specimens, we excluded those blood stream isolates were a potential skin contaminants (e.g. *Diphtheroids*, *Bacillus* species, or *Micrococcus*) had been cultured, except the specimens which the same bacteria was isolated in two successive cultures (3). All bacterial and fungal isolates were microbiologically identified in the microbiology department of Milad central laboratory using standard biochemical identification methods<sup>16</sup>. Antimicrobial susceptibility testing was performed by Kirby-Bauer disk diffusion method and zone diameter were measured following CLSI criteria<sup>17</sup>. Quality controls methods were routinely in place at the microbiology department of Milad central laboratory. Owing to the high risk of BSI morbidity and mortality, bacterial isolates that showed intermediate susceptibility to an antimicrobial agent were categorized as resistant isolates for data analysis and presentation<sup>15</sup>. All analysis was performed using SPSS 16 software and  $\chi^2$ -test or Fisher's exact test.  $P < 0.05$  were considered statistically significant.

### RESULTS

Results obtained from 3212 blood cultures have been done during 2009-2010, showing that 371 (11.6%) of cases of patients were infected. The result of our study revealed that

53.4% of BSIs did occur in men. Approximately 121 cases (32.6%) of BSIs were derived from Intensive Care Units (ICU) such as General ICU, Emergency ICU, Open heart ICU, Pediatrics ICU and Neonates ICU, the number of infected cases with gram positive bacteria, gram negative bacteria or fungi were 35, 75 and 11, respectively.

Totally our results, as shown in table 1 data indicated that 66% of BSIs were related to gram negative bacteria, while 24.1% of infected blood culture was caused by gram positive bacteria, and the rest of them (9.9%) were infected by yeast. Among gram negative bacteria, *Enterobacter cloacae* and *Escherichia coli* were more prevalent showing a frequency of 26.4% and 23.6%, respectively and among gram positive bacteria *Staphylococcus aureus* with frequency of 47.8% and thereafter Coagulase-negative *Staphylococci* (CoNS) with frequency of 16.8% were more prevalent. Among fungi, nonalbicans *Candida* with a frequency of 81.1% was the most prevalent.

At this study, referencing to CLSI, we used 24 antibiotics for antibiogram testing (table 2). Investigation of gram positive bacteria antibiogram indicated that Vancomycin and Chloramphenicol were most effective antibiotics with 94.4% and 92% sensitive cases, respectively. All of the isolated *S. aureus*, CoNS and *Enterococcus faecalis* were susceptible to Vancomycin, while 55.6% of isolated *Enterococcus faecium* were resistant to Vancomycin (Vancomycin Resistant *Enterococci* VRE). Frequency of CoNS and *S. aureus* sensitivity for Chloramphenicol were 100% and 93%, respectively. We found that 54.1% of isolated *S. aureus* was methicillin resistant *S. aureus* (MRSA).

The isolated gram negative bacteria indicated that these bacteria were more sensitive to Imipenem (IPM) and secondly to Ciprofloxacin with frequency of 79.7% and 70.3%, respectively. *E. cloacae* and *E. coli* were sensitive to IPM with frequency of 97.9% and 97.6%, respectively. Results of antibiotic susceptibility pattern data for *Acinetobacter baumannii* indicated Colistin (100%) and Imipenem (42.9%) are the more effective agents.

It is noteworthy that during this study there were not any significant difference in antibiotic resistance in the first six month of the year compared to the second six month of the year (data is not showed).

**Table 1.** Frequency of isolated microorganisms from BSIs

Gram positive isolated bacteria	ICU	Non ICU	TOTAL
<i>Staphylococcus aureus</i>	20	23	43
<i>Enterococcus faecalis</i>	4	7	11
<i>Enterococcus faecium</i>	4	5	9
<i>Streptococcus viridans</i>	1	7	8
<i>Staphylococcus warneri</i>	-	5	5
<i>Staphylococcus pasteurii</i>	3	4	7
<i>Streptococcus pneumoniae</i>	-	2	2
<i>Streptococcus pyogenes</i>	1	-	1
<i>Streptococcus agalactiae</i>	-	1	1
<i>Streptococcus milleri</i>	1	-	1
<i>Staphylococcus capitis</i>	1	-	1
<i>Staphylococcus hominis</i>	-	1	1
Gram negative isolated bacteria	ICU	Non ICU	TOTAL
<i>Enterobacter cloacae</i>	6	59	65
<i>Escherichia coli</i>	8	50	58
<i>Acinetobacter baumannii</i>	29	10	39
<i>Stenotrophomonas maltophilia</i>	4	18	22
<i>Burkholderia cepacia</i>	4	16	20
<i>Pseudomonas aeruginosa</i>	8	5	13
<i>Klebsiella pneumoniae</i>	10	3	13
<i>Providencia rettgeri</i>	2	1	3
<i>Haemophilus influenzae</i>	-	2	2
<i>Enterobacter aerogenes</i>	-	2	2
<i>Citrobacter koseri</i>	-	2	2
<i>Acinetobacter</i> spp.	-	1	1
<i>Klebsiella ozaeana</i>	1	-	1
<i>Klebsiella planticola</i>	1	-	1
<i>Providencia stuartii</i>	1	-	1
<i>Sphingomonas</i> spp.	1	-	1
<i>Moraxella</i> spp.	-	1	1
Isolated fungi	ICU	Non ICU	TOTAL
<i>Candida</i> spp.	10	20	30
Yeast (No <i>Candida albicans</i> & SPP.)	-	5	5
<i>Candida albicans</i>	1	-	1

## DISCUSSION

Reviewing reports from other countries showed that BSIs occurred more frequently in men than in women<sup>18</sup>. Also data indicated proximally that only 30% of BSIs isolated from ICU are clinically and epidemiologically valuable for further studying<sup>19</sup>. Our data as some of other international studies indicated that the frequency of BSIs with gram negative bacteria is more prevalent than gram positive bacteria<sup>5,14</sup>.

At our study, among isolated gram positive bacteria, *S.aureus* and secondly CoNS

were more frequent, which was similar to the data achieved from other studies<sup>5,14</sup>. About frequency of gram negative bacteria, there were various reports, but mostly was mentioned as one of the important isolated bacteria –these reports mentioned *E.coli* as the first or the second most frequent isolated bacteria<sup>4,14,15</sup>. In the field of gram positive frequency our results were not consistent with other studies. The most recently studies indicated that most common gram positive pathogens causing BSI were coagulase-negative staphylococci. Other studies performed in our country also confirmed this finding. In a study

Table 2. Percentage of antibacterial susceptibility among gram positive and gram negative bacteria

	Gram positive bacteria						Gram negative bacteria						
	<i>S.aureus</i>	CoNS	<i>E. faecalis</i>	<i>E. faecium</i>	Others	<i>E.colocae</i>	<i>E.coli</i>	<i>A. baumannii</i>	<i>S. aeruginosa</i>	<i>B. maltophilia</i>	<i>P. cepacia</i>	<i>K. pneumonia</i>	Others
AM <sup>1</sup>	10.3	7.7	90.9	22.2	80.0	4.6	10.7	2.8	0.0	0.0	0.0	0.0	7.1
AN <sup>2</sup>	45.5	75.0	NT	NT	NT	51.7	86.0	28.6	22.7	63.2	30.8	10.0	61.5
AZM <sup>3</sup>	44.1	0.0	NT	NT	84.6	NT	NT	NT	NT	NT	NT	NT	NT
C <sup>4</sup>	95.2	100	81.8	66.7	100	52.5	72.7	2.6	23.8	15.0	0.0	53.8	42.9
CAZ <sup>5</sup>	NT	NT	NT	NT	NT	37.5	69.0	2.8	18.2	5.0	23.1	7.7	40.0
CC <sup>6</sup>	14.6	7.7	NT	NT	38.5	NT	NT	NT	NT	NT	NT	NT	NT
CF <sup>7</sup>	NT	NT	NT	NT	NT	4.8	52.5	0.0	0.0	0.0	0.0	0.0	7.7
CP <sup>8</sup>	47.6	28.6	NT	NT	77.8	92.3	74.6	23.1	100	75.0	69.2	30.8	66.7
CRO <sup>9</sup>	32.6	14.3	NT	NT	90.0	42.2	67.8	5.3	0.0	20.0	0.0	15.4	40.0
CS <sup>10</sup>	NT	NT	NT	NT	NT	100	100	100	NT	NT	NT	100	NT
CT <sup>11</sup>	NT	NT	NT	NT	NT	45.3	69.5	5.6	0.0	5.3	0.0	15.4	57.1
CTX <sup>12</sup>	NT	NT	NT	NT	NT	38.7	67.2	0.0	0.0	10.0	0.0	15.4	46.7
CZ <sup>13</sup>	NT	NT	NT	NT	NT	3.3	50.0	0.0	0.0	0.0	0.0	0.0	8.3
E <sup>14</sup>	42.9	0.0	9.1	22.2	83.3	NT	NT	NT	NT	NT	NT	NT	NT
Gm <sup>15</sup>	45.9	27.3	NT	NT	NT	53.1	79.3	41.0	31.8	57.9	30.8	7.7	53.8
IPM <sup>16</sup>	NT	NT	NT	NT	NT	97.9	97.6	42.9	0.0	92.9	44.4	85.7	87.5
OX <sup>17</sup>	54.1	50.0	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT
P <sup>18</sup>	2.4	0.0	18.2	0.0	72.7	NT	NT	NT	NT	NT	NT	NT	NT
RA <sup>19</sup>	57.1	66.7	0.0	0.0	85.7	NT	NT	NT	NT	NT	NT	NT	NT
SXT <sup>20</sup>	85.7	15.4	NT	NT	22.2	49.2	70.2	15.4	95.5	63.2	12.5	30.8	40.0
TE <sup>21</sup>	25.6	35.7	18.2	22.2	58.3	30.2	57.6	12.8	0.0	63.2	23.1	23.1	35.7
TOB <sup>22</sup>	NT	NT	NT	NT	100	47.5	62.2	32.4	10.5	35.3	23.1	7.7	50.0
V <sup>23</sup>	100	100	100	44.4	100	NT	NT	NT	NT	NT	NT	NT	NT
XM <sup>24</sup>	NT	NT	NT	NT	NT	4.9	33.3	0.0	0.0	0.0	0.0	0.0	0.0

1. Ampicilin, 2. Amikacin, 3. Azithromycin, 4. Chloramphenicol, 5. Cefazidime, 6. Clindamycin, 7. Cephalotin, 8. Ciprofloxacin, 9. Ceftriaxone, 10. Colistin, 11. Ceftizoxime, 12. Cefotaxime, 13. Cefazolin, 14. Erythromycin, 15. Gentamycin, 16. Imipenem, 17. Oxacillin, 18. Penicillin, 19. Rifampin, 20. Trimethoprim Sulfamethoxazol, 21. Tetracycline, 22. Tobromycin, 23. Vancomycin, 24. Cefuroxime NT: Not Tested

done by Rahbar *et al* showed that Gram-positive cocci, included coagulase-negative staphylococci, *Staphylococcus aureus*, *Streptococcus pneumoniae* and other Gram-positive cocci, accounted for 42.3% of isolates. Gram-negative bacilli were responsible for another 42.3% of isolates. One reason for controversy among these finding could be due to number of blood culture taken for each patient, unfortunately in our study only one blood specimens per patient had been taken cultured. We had many episodes of Cones isolates but because of probability contamination we excluded these cases from our study. With regard to the susceptibility of *S.aureus* to Vancomycin (100% sensitive) our report about, it completely matched the data reported from Europe (SENTRY) (4) but it was different than previous report from one of the Iran medical center's that they had isolated *S.aureus* resistant to Vancomycin with frequency of 21%. This finding maybe related to admission of antibiotic before sampling or in methods of identification of antimicrobial susceptibility in previous study. At this study our finding about *E.faecium* and its susceptibility to Vancomycin resemble the data reported by SENTRY study<sup>4</sup>. Our achievement about the frequency of susceptibility to Chloramphenicol for CoNS and *Enterococcus- spp.* was similar to data published in USA, Canada and Latin America but the susceptibility of *S.aureus* to Chloramphenicol in our finding was more prominent than previous studies<sup>14</sup>.

Unfortunately, until now there has not been not any coordinated survey about frequency of isolated bacteria from BSIs in clinical centers in Iran, so we believe that this study can be used as the base of a cohort study to highlight epidemiological trend and frequency of bacteria and its antimicrobial susceptibility pattern in Iran or maybe even in Middle East. This cohort study not only can be used as a source to recognize local infections for some of special important infections such as *Acinetobacter spp.* in order to prevent extension of multidrug resistance bacteria but also it can be used to decrease technical deviation and personal errors in medical laboratories (9,10,12). Perhaps, the reason of differences occurred in significance of antimicrobial resistant pattern is due to the deficiencies in specimen collection and the duration of the study(13, 14).

## CONCLUSIONS

In fact, our study indicates that blood cultures, specimen collection and the antimicrobial drug resistance should be performed by standard methods. Surveillance studies must be established in the country and the data should be evaluated periodically. Certainly these data could be used for appropriate use of antibiotic therapy and decrease the morbidity and mortality ratio caused by nosocomial infections.

## REFERENCES

1. McGregor C. J., Rich E.S., Harris D. A., Perencevich N. E., Osih R., Lodise P. T., Miller R. R., and Furuno P. J., A Systematic Review of the Methods Used to Assess the Association between Appropriate Antibiotic Therapy and Mortality in Bacteremic Patients. Inappropriate Therapy for Bacteremia. *CID*; 2007; **45**: 329-337.
2. Zaragoza R., Artero A., Camarena J.J., Sancho S., Gonzales R. and N. J.M., The influence of inadequate empirical antimicrobial treatment on patients with bloodstream infection in an intensive care unit. *Clinical Microbiology and Infection*, 2003; **9**: 413-418.
3. Wisplinghoff H., Bischoff T., Tallent M.S., Seifert H., Wenzel P. R. and Edmond B. M., Nosocomial Bloodstream Infections in US Hospitals: Analysis of 24,179 Cases from a Prospective Nationwide Surveillance Study. *Clinical Infectious Diseases*; 2004; **39**: 309-17.
4. Fluit C. A., Jones E. M., Schmitz J. F., Acar J., Gupta R., Verhoef J. and the SENTRY Patients Group; Antimicrobial Susceptibility and Frequency of Occurrence of Clinical Blood Isolates in Europe from the SENTRY Antimicrobial Surveillance Program, 1997 and 1998. *Clinical Infectious Diseases*; 2000; 454-460.
5. Decusser J.-W., Pina P., Delalande C., Pangon B., Courvalin P., Allouch P., and the ColBVH study group; Frequency of isolation and antimicrobial susceptibility of bacterial pathogens isolated from patients with bloodstream infections: a French prospective national survey. *Journal of Antimicrobial Chemotherapy*. 2003; **51**: 1213-1222.
6. Karlowsky J. A., Jones M. E., Draghi D. C., Thornsberry C., Sahn D. F. and Volturo G. A.; Prevalence and antimicrobial susceptibilities of bacteria isolated from blood cultures of

- hospitalized patients in the United States in 2002. *Annals of Clinical Microbiology and Antimicrobials*. 2004; **3**: 4.
7. Halim T. Y., Song K. W., Barnett M. J., Forrest D. L., Hogge D. E., Nantel S. H., Nevill T. J., Shepherd J. D., Smith C. A., Sutherland H. J., Toze C. L., and Lavoie J. C., Positive impact of selective outpatient management of high-risk acute myelogenous leukemia on the incidence of septicemia. *Annals of Oncology*. 2007; **18**: 1246-1252.
  8. Bennett K., Sharp S. E., Rapid Differentiation of methicillin-resistant *Staphylococcus aureus* and methicillin-susceptible *Staphylococcus aureus* from blood culture by use of direct ceftoxitin disk diffusion test. *Journal of Clinical Microbiology*, 2008; 3836-3838.
  9. Wisplinghoff H., Edmond M. B., Pfaller M. A., Jones R. N., Wenzel R. P., and Seifert H., Nosocomial Bloodstream Infection Caused by Acinetobacter Species in United States Hospitals: Clinical Features, Molecular Epidemiology, and Antimicrobial Susceptibility. *Clinical Infectious Diseases*; 2000; **31**: 690-7.
  10. Brito D. D., Oliveira E. J., Abdallah V. O. S., Darini A. L. C. and Filho P. P. G., An Outbreak of *Acinetobacter baumannii* Septicemia in a Neonatal Intensive Care Unit of a University Hospital in Brazil. *The Brazilian Journal of Infectious Diseases*; 2005; **9**(3): 301-309.
  11. Velasco E., yington R., Martins C. S. A., Schirmer M., Dias L. C. M. and Gocalves V. M. S., Bloodstream infection surveillance in a cancer centre: a prospective look at clinical microbiology aspects. *Clin Microbiol Infect*; 2004; **10**: 542-549.
  12. Sunenshine R. H., Wright M-O., Margaskis L. L., Harris A. D., Song X., Hebden J., Cosgrove S. E., Anderson A., Carnell J., Jernigan D. B., Kleinbaum D. G., Perl T. M., Standiford h. C. and Srinivasan A., Multidrug-resistant *Acinetobacter* infection Mortality Rate and Length of Hospitalization. *Emerging Infectious Diseases*. 2007; **13**(1): 97-103.
  13. Osih R. B., McGregor J. C., Rich S. E., Moore A. C., Furuno J. P., Prenevich E. N., and Harris A. D. Impact of Empiric Antibiotic Therapy on outcomes in Patients with *Pseudomonas aeruginosa* Bacteremia. *Antimicrobial Agents and Chemotherapy*, 2007; 839-844.
  14. Diekema D. J., Pfaller M. A., Jones R. N., Doern G. V., Kugler K. C., Beach M. L., Sader H. S. and the SENTRY participants Group. Trends in antimicrobial susceptibility of bacterial pathogens isolated from patients with bloodstream infections in the USA, Canada and Latin America. *International Journal of Antimicrobial*. 2000; 257-271.
  15. Mamishi S., Pourakbari B., Ashtiani M. H., Hashemi F. B., Frequency of isolation and antimicrobial susceptibility of bacteria isolated from bloodstream infections at Children's Medical Center, Tehran, Iran, 1996-2000. *International journal of Antimicrobial*. 2005; 373-379.
  16. Reisner S. B., Woods G. L., Thomason R. P., Laron D. H., Garsia L. S., Shimuzu R. Y., Specimen collection. In: Murray P.R., Baron E. J., Tenover F. C., Tenover R. H., editors. *Manual of clinical microbiology*. 7th ed. Washington, DC: American Society for Microbiology; 1999; 64-76.
  17. Clinical and Laboratory Standard Institute. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. Approved Standard M7-A4. Wayne 2007.
  18. Hill P. C., Onyeama C. O., Ikumapayi U. NA., Secka O., Ameyaw S., Simmonds N., Donkor S. A., Howie S. R., Tapgun M., Corrah T. and Adegbola R. A., Bacteraemia in patients admitted to an urban hospital in West Africa. *BioMedCentral Infectious Diseases*, 2007; **7**: 2.
  19. Ibrahim E. H., Sherman G., Ward S., Fraser V. J., and Kollef M., The Influence of Inadequate Antimicrobial Treatment of Bloodstream Infections on Patient Outcomes in the ICU Setting. *Chest*; 2000; **118**: 146-155.
  20. Gaidelyte A., Vaara M. and Bamford D. H., Bacteria, Phages and Septicemia. *PLoS ONE* 2(11): e1145.doi:10.1371/journal.pone.0001145. Rahbar M, Gra-Agaji R, Hashemi S. Nosocomial blood stream infections in Imam Khomeini Hospital, Urmia, Islamic. 2007.