

Safety Aspects of *Lactococcus lactis* as a Dairy Starter

Umar Farooq*, Xiaoming Liu and Hao Zhang

State Key Laboratory of Food Science and Technology, School of Food Science and Technology, Jiangnan University, Lihu Road 1800, Wuxi 214122, Jiangsu, China.

(Received: 04 February 2012; accepted: 02 April 2012)

Lactococcus lactis owing to its completely sequenced genome and biochemical characteristics is a major dairy starter used in dairy product processing. Safety of any starter is dependent on its unequivocal identification at the genus, species, and strain level along with its potential ability to transfer antibiotic resistance genes, as functional effects is strain specific. Prior to incorporating new strains into products their efficacy should be carefully assessed, and a case by case evaluation as to whether they share the safety status of traditional food-grade organisms should be made.

Key Words: *Lactococcus lactis*; Dairy starter; Taxonomy; Antibiotic resistance; Safety aspects.

Lactococcus lactis is by far the most extensively studied lactic acid bacterium, used as a primary component of starter cultures in the dairy industry, especially in the manufacture of cheese.¹ The species are commonly isolated from plant material, but the most recognized habitat is dairy products. Isolates identified as gram positive, catalase negative, homofermentative, microaerophilic cocci, and exclusively produce L (+) lactic acid. Unlike many members of *Streptococcus thermophilus*, *Lactococcus lactis* species do not grow in long chains, mostly in pairs or in short chains, depending on growth conditions appears ovoid with typically 0.5 - 1.5 μM in length and unable to grow at 45 °C, pH 9.6 or in 6.5% NaCl broth.²

L. lactis being first lactic acid bacterium with completely sequenced genome^{3,4}, is an excellent microorganism for advanced analysis. Over the last decade, novel insights into metabolism and function of *Lactococcus lactis* has sparked renewed interest for application of a new generation of starter cultures for various dairy products, fermented foods, probiotics, and even live vaccines^{5,6}. However, it also gives rise to concern because of the potential transfer of antibiotic resistance and their role in human infections. The evaluation of safety aspects of such improved or novel strains that are to be used in possible future applications is an important issue⁷. Thus an up to date analysis of safety aspects of *Lactococcus lactis* is needed to ensure consumer safety.

Function of *Lactococcus lactis* as a dairy starter

Lactococcus lactis isolates are the most important industrial dairy starter in the manufacture of a wide range of fermented products, including sour cream, butter, fresh and soft cheeses and various hard and semi hard cheeses composed of single or multiple strains with or without other lactic acid bacteria⁸.

* To whom all correspondence should be addressed.
Tel : +86-510 -85329093; Fax : +86-510 -85912155
E-mail: zhanghao@jiangnan.edu.cn

The function of *Lactococcus lactis* is twofold; on one hand, they produce lactic acid at an appropriate rate, usually through fermentative degradation of the sugars present in the raw materials. It results in lowering of the pH that makes the medium inhospitable for most spoilage and/or pathogenic organisms and contribute to extend the shelf life span of the food. In addition, Nisin produced by certain species of *Lactococcus lactis* is a 34-amino acid peptide and is one of the well characterized lantibiotics containing one lanthionine and four β -methylanthionine cyclic structures⁹. Since Nisin has strong antimicrobial activity against gram positive bacteria, it has been widely used as most effective food preservative^{10,11}. On the other hand, these bacteria make important contributions to development of texture by producing exopolysaccharides¹² and to the development of cheese flavour, initiating proteolysis during ripening which is believed to be result from the action of enzymes from the starter cultures¹³ as acid accumulation changes the rheological and organoleptical properties of the product, a process that is complemented by the production and, in some instances, the secretion of hydrolytic enzymes, mainly proteinases, peptidases, amino acid hydrolases and, to a lower extent, lipases and esterases¹⁴.

Safety aspects

The present review is limited to two important safety aspects.

Taxonomy and identification tools as the basis of safety evaluation

The safety of a novel starter is dependent on its unequivocal identification at the genus, species, and strain level; as functional and technological aspects is strain specific and to avoid the inclusion of potentially pathogenic microorganisms in commercial products. *Lactococcus lactis* were originally classified under the genus *Streptococcus*, but in 1985, Schleifer et al., provided evidence, based on chemotaxonomic studies confirmed by 16s rRNA sequencing, to reclassify some species of the genera *Streptococcus* (Lancefield group N lactic streptococci) and *Lactobacillus*, that justifies conferring genus *Lactococcus*². The *Lactococcus* genus is constituted by six species, *Lactococcus chungangensis*, *L. garvieae* (formerly *E. serolicida*), *L. piscium*, *L. plantarum*, *L.*

raffinolactis, (formerly *S. raffinolactis*) and *L. lactis*^{15,16,17}. The *Lactococcus lactis* specie is further differentiated into three subspecies, namely *L. lactis subsp. cremoris*, *L. lactis subsp. hordniae* (formerly *Lactobacillus hordniae*) and *L. lactis subsp. Lactis*. However *L. lactis subsp. cremoris* and *L. lactis subsp. lactis* have been found more important in dairy applications while *L. lactis subsp. hordinae* has no relevance in fermented food production¹⁸. A combination of phenotypic traits and genotypic information has been used to investigate microbial diversity within *Lactococcus lactis* sub species¹⁹. *L. lactis subsp. cremoris* is distinguished from *L. lactis subsp. lactis* by its inability to (a) grow at 40°C, (b) grow in 4% NaCl, (c) hydrolyze arginine, (d) ferment ribose and (e) grow at pH 9.2²⁰.

The *lactis* and *cremoris* subspecies of *L. lactis* have also been shown to be genetically distinct by highly discriminatory molecular methods, DNA-DNA homology studies, the polymorphism of the 16S-23S rDNA spacer region²¹, multiple locus micro satellite analysis²², comparison of 16S rRNA sequences²³, PCR-DGGE (Denaturing Gradient Gel Electrophoresis) analyses²⁴ and comparative genome hybridization using multi-strain arrays.²⁵

The diversity within the species has recently been re-evaluated with molecular analyzes, confirming that phenotypic and genotypic diversity are not coherent¹⁸. Currently, a clear example is commonly worldwide used laboratory strains MG1363; it displays many of *subsp. lactis* phenotypic traits, but it is usually referred to as *subsp. cremoris* due to its genetic similarity to *subsp. cremoris*²⁶. Moreover, It is clear that strains used as starter in dairy industry should be identified using molecular methods and up-to-date taxonomical nomenclature and comparison of the results obtained by using different molecular methodologies (polyphasic approach) is the best way to establish strain identity. Therefore in future, the genetic based nomenclature will probably overcome the classical and phenotype-based one²⁷, generating a dichotomy in taxonomic procedure of identification of strains at the sub species level.

Safety consideration of antibiotic resistance in *Lactococcus lactis*

The presence of antibiotic resistance

genes in the lactic acid bacteria genomic content is not a safety concern in itself, as long as the genes are not mobilized and transferred to other bacteria²⁸. Thus, it is of great interest to investigate whether *Lactococcus* can act as reservoirs for antibiotic resistance genes, from which they could be spread to opportunistic or pathogenic bacteria. First strong evidence that antibiotic resistance can be spread in a food environment was found by Perrreten et al.²⁹ who clearly observed genes conferring resistance for streptomycin, tetracycline and chloramphenicol in *L. lactis subsp. lactis* K214 strain isolated from a raw-milk soft cheese, encoded by three different genes, located in a multi-antibiotic resistance plasmid, and these genes were almost identical to others previously found in *Staphylococcus aureus* and *Listeria monocytogene*³⁰. Since then, many genes coding for proteins presenting resistance to several antibiotics, mainly tetracycline and erythromycin, have been exemplified in *Lactococcus lactis*³¹, and transfer from *Lactococcus* to other bacteria, including Gram-positive pathogens, as well as the inheritance of resistance genes, has been demonstrated³². Further studies are needed to elucidate whether specific virulence factors are carried and expressed by the *L.lactis* clinical isolates, conferring to these strains the specific ability to cause infection in humans.

In spite of this, rare cases of human infection were reported, however, all of which concerned immuno suppressed patients^{33, 34}. Yet, many open questions regarding the safety of starters remain. Thus, future, *in vivo* experiments should shed some light on the transfer events occurring from, via, or to starters. It is also difficult to assess what level of gene transfer, if any, may be considered acceptable by the community and also significant reason to select strains lacking the potential to transfer genetic determinants of antibiotic resistance. There is little basis for scientific regulation of strains with intrinsic resistance, as little is known about the levels of intrinsic resistance in current starters and should be carefully investigated.

It is evident that potential ability of starter strains to transfer antibiotic resistances to pathogenic bacteria in the food and gut environment should be considered in the safety assessment. Also, it appears that the

gastrointestinal tract may comprise a more favorable environment for antibiotic resistance transfer than conditions provided *in vitro*³⁵.

CONCLUSION

Despite the excellent overall safety record of *Lactococcus lactis* in human, they should be used with caution in certain specific patient groups particularly critically patients, those with immune deficiency and patient groups with increased risk for bacterial translocation due to disturbed intestinal mucosal barrier function. Taken together, the beneficial aspects of *Lactococcus lactis* clearly outweigh its rare sepsis.

Lactococcus lactis is Generally Recognized As safe (GRAS) microorganism. However, fundamental and applied clinical research is still needed to optimally explore its potential as functional starter cultures in the existing production technology to obtain quantitative and qualitative data and the host strains used for those purposes should be composed solely of DNA food-grade organism and devoid of any antibiotic resistance markers. Otherwise, they may potentially transfer to intestinal microflora in humans, there by compromising human antibiotic therapy.

ACKNOWLEDGMENTS

This work was supported by the National Science Fund for Distinguished Young Scholars (No. 31125021), the National High Technology Research and Development Program of China (863 Program No. 2011AA100901, 2011AA100902), the Key program of National Natural Science Foundation of China (No. 20836003), the National Basic Research Program of China (973 Program No. 2012CB720802), the National Science and Technology Pillar Program (No. 2010CB0070311), the 111 project B07029, and Fundamental Research Funds for the Central Universities (JUSRP111A31 and JUSRP31103).

REFERENCES

1. Hejnowicz, M. S., Golebiewski, M., & Bardowski, J. Analysis of the complete genome sequence of the *lactococcal* bacteriophage bIBB29. *Int. J. of Food Microbiol.* 2009; **131**, 52-61.

2. Casalta, E., & Montel, M. C. Safety assessment of dairy microorganisms: the *Lactococcus* genus. *Int. J. of Food Microbiol.* 2008; **126**, 271-273.
3. Bolotin, A., Wincker, P., Mauger, S., Jaillon, O., Malarme, K., Weissenbach, J., Ehrlich, S. D., & Sorokin, A. The complete genome sequence of the lactic acid bacterium *Lactococcus lactis* ssp. *lactis* IL1403. *Gen. Res.*, 2001; **11**, 731-753.
4. Siezen, R. J., Bayjanov, J., Renckens, B., Wels, M., van Hijum, S. A., Molenaar, D., & van Hylckama Vlieg, J. E. Complete genome sequence of *Lactococcus lactis* subsp. *lactis* KF147, a plant-associated lactic acid bacterium. *J. of Bacteriol.* 2010; **192**, 2649-2650.
5. Hugenholtz, J. The lactic acid bacterium as a cell factory for food ingredient production. *Int Dairy J.* 2008; **18**, 466-475.
6. Gao, Y., Lu, Y., Teng, K. L., Chen, M. L., Zheng, H. J., Zhu, Y. Q., & Zhong, J. Complete Genome Sequence of *Lactococcus lactis* subsp. *lactis* CV56, a Probiotic Strain Isolated from the Vaginas of Healthy Women. *J. of Bacteriol.* 2011; **193**, 2886-2887.
7. Klare, I., Konstabel, C., Werner, G., Huys, G., Vankerckhoven, V., Kahlmeter, G., Hildebrandt, B., Muller-Bertling, S., Witte, W., & Goossens, H. Antimicrobial susceptibilities of *Lactobacillus*, *Pediococcus* and *Lactococcus* human isolates and cultures intended for probiotic or nutritional use. *J. of Antimic. Chemo.* 2007; **59**, 900-912.
8. Delorme, C. Safety assessment of dairy microorganisms: *Streptococcus thermophilus*. *Int. J. of Food Microbiol.* 2008; **126**, 274-277.
9. Reviriego, C., Fernández, A., Horn, N., Rodríguez, E., Marín, M. L., Fernández, L., & Rodríguez, J. M. Production of pediocin PA-1, and coproduction of nisin A and pediocin PA-1, by wild *Lactococcus lactis* strains of dairy origin. *Int. Dairy J.* 2005; **15**, 45-49.
10. Alegria, A., Delgado, S., Roces, C., Lopez, B., & Mayo, B. Bacteriocins produced by wild *Lactococcus lactis* strains isolated from traditional, starter-free cheeses made of raw milk. *International Journal of Food Microbiology*. 2010; **143**, 61-66.
11. Mitra, S., Chakrabarty, P. K., & Biswas, S. R. Potential production and preservation of dahi by *Lactococcus lactis* W8, a nisin-producing strain. *LWT-Food Sci. and Tech.* 2010; **43**, 337-342.
12. Ruas-Madiedo, P., Alting, A. C., & Zoon, P. Effect of exopolysaccharides and proteolytic activity of *Lactococcus lactis* subsp. *cremoris* strains on the viscosity and structure of fermented milks. *Int. D. J.* 2005; **15**, 155-164.
13. Smit, B. A., van Hylckama Vlieg, J. E., Engels, W. J., Meijer, L., Wouters, J. T., & Smit, G. Identification, cloning, and characterization of a *Lactococcus lactis* branched-chain alpha-keto acid decarboxylase involved in flavor formation. *Appl. And Env. Microbiol.* 2005; **71**, 303-311.
14. Ayala-Hernández, I., Hassan, A. N., Goff, H. D., & Corredig, M. Effect of protein supplementation on the rheological characteristics of milk permeates fermented with exopolysaccharide-producing *Lactococcus lactis* subsp. *cremoris*. *Food Hydrocoll.* 2009; **23**, 1299-1304.
15. Schleifer, K. H., Kraus, J., Dvorak, C., Renate Kilpper-Bälz, Collins, M. D., Fischer, W. Transfer of *Streptococcus lactis* and related *Streptococci* to the *Lactococcus* genus. *Sys. and Appl. Microbiol.* 1985; **6**, 183-195.
16. Schleifer, K. H., Kilipper-Balz, R. Molecular and chemotaxonomic approaches to the classification of *streptococci*, *enterococci* and *lactococci*: a review. *Sys. and Appl. Microbiol.* 1987; **6**, 1-19.
17. Tanigawa, K., Kawabata, H., & Watanabe, K. Identification and typing of *Lactococcus lactis* by matrix-assisted laser desorption ionization-time of flight mass spectrometry. *Appl. and Env. Microbiol.* 2010; **76**, 4055-4062.
18. Rademaker, J. L., Herbet, H., Starrenburg, M. J., Naser, S. M., Gevers, D., Kelly, W. J., Hugenholtz, J., Swings, J., & van Hylckama Vlieg, J. E. Diversity analysis of dairy and nondairy *Lactococcus lactis* isolates, using a novel multilocus sequence analysis scheme and (GTG)₅-PCR fingerprinting. *Appl. And Env. Microbiol.* 2007; **73**, 7128-7137.
19. Siezen, R. J., Starrenburg, M. J., Boekhorst, J., Renckens, B., Molenaar, D., & van Hylckama Vlieg, J. E. Genome-scale genotype-phenotype matching of two *Lactococcus lactis* isolates from plants identifies mechanisms of adaptation to the plant niche. *Appl And Env. Microbiol.* 2008; **74**, 424-436.
20. Psoni, L., Kotzamanidis, C., Yiangou, M., Tzanetakis, N., & Litopoulou-Tzanetaki, E. Genotypic and phenotypic diversity of *Lactococcus lactis* isolates from Batzos, a Greek PDO raw goat milk cheese. *Int. J. of Food Microbiol.* 2007; **118**, 211-220.
21. Blaiotta, G., Pepe, O., Mauriello, G., Villani, F., Andolfi, R., & Moschetti, G. 16S-23S rDNA intergenic spacer region polymorphism of *Lactococcus garvieae*, *Lactococcus raffinolactis* and *Lactococcus lactis* as revealed by PCR and

- nucleotide sequence analysis. *Sys. and Appl. Microbiol.*2002; **25**, 520-527.
22. Quenee, P., Lepage, E., Kim, W. S., Vergnaud, G., & Gruss, A. Minisatellite polymorphism as a tool to distinguish closely related *Lactococcus lactis* strains. *FEMS Microbiol. Lett.*2005; **248**, 101-109.
 23. Van Hylckama Vlieg, J. E. T., & Hugenholtz, J. Mining natural diversity of lactic acid bacteria for flavour and health benefits. *Int.Dairy J.*2007; **17**, 1290-1297.
 24. Giannino, M. L., Marzotto, M., Dellaglio, F., & Feligini, M. Study of microbial diversity in raw milk and fresh curd used for Fontina cheese production by culture-independent methods. *Int. J. of Food Microbiol.*2009; **130**, 188-195.
 25. Siezen, R. J., Bayjanov, J. R., Felis, G. E., van der Sijde, M. R., Starrenburg, M., Molenaar, D., Wels, M., van Hijum, S. A., & van Hylckama Vlieg, J. E. Genome-scale diversity and niche adaptation analysis of *Lactococcus lactis* by comparative genome hybridization using multi-strain arrays. *Micro. Biotech.*2011; **4**, 383-402.
 26. Wegmann, U., O'Connell-Motherway, M., Zomer, A., Buist, G., Shearman, C., Canchaya, C., Ventura, M., Goesmann, A., Gasson, M. J., Kuipers, O. P., van Sinderen, D., & Kok, J. Complete genome sequence of the prototype lactic acid bacterium *Lactococcus lactis* subsp. *cremoris* MG1363. *J. of Bacteriol.*2007; **189**, 3256-3270.
 27. Kok, J., Buist, G., Zomer, A. L., van Hijum, S. A., & Kuipers, O. P. Comparative and functional genomics of *lactococci*. *FEMS Micro. Rev.*2005; **29**, 411-433.
 28. EFSA. Introduction of a Qualified Presumption of Safety (QPS) approach for assessment of selected microorganisms referred to EFSA. *The EFSA J.*2007; **587**, 1-16. .
 29. Perreten, V., Schwarz, F., Cresta, L., Boeglin, M., Dasen, G., & Teuber, M. Antibiotic resistance spread in food. *Nature.*1997; **389**, 801-802.
 30. Chopra, I., & Roberts, M. Tetracycline antibiotics: mode of action, applications, molecular biology, and epidemiology of bacterial resistance. *Micro. and Molec. Bio. Rev.*2001; **65**, 232-260.
 31. Ammor, M. S., Florez, A. B., van Hoek, A. H., de Los Reyes-Gavilan, C. G., Aarts, H. J., Margolles, A., & Mayo, B. Molecular characterization of intrinsic and acquired antibiotic resistance in lactic acid bacteria and *bifidobacteria*. *J. of Molec. Microbiol. and Biotec.*2008; **14**, 6-15.
 32. Aquilanti, L., Garofalo, C., Osimani, A., Silvestri, G., Vignaroli, C., & Clementi, F. Isolation and molecular characterization of antibiotic-resistant lactic acid bacteria from poultry and swine meat products. *J. of Food Prot.*2007; **70**, 557-565.
 33. Florez, A. B., Ammor, M. S., & Mayo, B. Identification of tet(M) in two *Lactococcus lactis* strains isolated from a Spanish traditional starter-free cheese made of raw milk and conjugative transfer of tetracycline resistance to *lactococci* and *enterococci*. *Int. J. of Food Microb.*2008; **121**, 189-194.
 34. Zechini, B., Cipriani, P., Papadopoulou, S., Di Nucci, G., Petrucca, A., & Teggi, A. Endocarditis caused by *Lactococcus lactis* subsp. *lactis* in a patient with atrial myxoma: a case report. *Diagnostic Microbiol and Infect. Dis.*2006; **56**, 325-328.
 35. Feld, L., Schjorring, S., Hammer, K., Licht, T. R., Danielsen, M., Krogfelt, K., & Wilcks, A. Selective pressure affects transfer and establishment of a *Lactobacillus plantarum* resistance plasmid in the gastrointestinal environment. *J. of Anti. Chemo.*2008; **61**, 845-852.