In vitro Anti-staphylococcal Potential of Endophytic Fungi from *Aegle marmelos*

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Staphylococcus aureus is the third most dreaded human pathogen which is getting refractory to current armamentarium of antimicrobial drugs. Endophytic fungi have been recognized as a fountainhead of novel bioactives for human therapeutic intervention and drug development. In the present investigation we report isolation and screening of endophytic fungi isolated from Aegle marmelos inhabiting conserved forest area in Western Ghats, India. A total of 25 endophytic fungi representing 10 genera were isolated by adopting a standardized protocol. Fusarium species exhibited maximum colonization (28%), followed by Aureobasidium (12%) and Lasiodiplodia species (8%). In vitro screening of culture broth of these fungi using a standardized anti-microbial assay exhibited activity only in one isolate #1005AMLBRT. Subsequently ethyl acetate extracts of the culture broths revealed that 15 isolates did not possess any antistaphylococcal potential. However the best anti-staphylococcal potential was found in ethyl acetate extract of #1005AMLBRT. #1005AMLBRT was found to be a new species of Alternaria named as Alternaria marmelos based on phylogenetic and morphological studies. Further studies on Alternaria marmelos are warranted for production, isolation and characterization of the anti-staphylococcal principle.

> **Keywords:** Endophytic fungi, Anti- Staphylococcal activity, Fermentation broth, *Alternaria, Aegle marmelos.*

Staphylococcus aureus has been recently identified as the third most dreaded pathogen responsible for mortality and morbidity in hospitals and community due to its refractory behaviour¹. S. aureus is an opportunistic pathogen which causes soft tissue and skin infections, respiratory infections, osteomyelitis, endocarditis and pneumonia². Patients with prosthetic devices are also found to be prone to resistant Staphylococcal infections apart from those who have undergone invasive surgical procedures³. Methicillin resistant Staphylococcus aureus (MRSA) and its resistant clones are responsible for nosocomial infections as well as community epidemics⁴⁻⁶. Currently MRSA and its variants are posing a critical situation for clinicians since they are frequently getting refractory to current armamentarium of antibiotics^{1,7}. Accordingly it is imperative to explore for new anti-MRSA agents to treat chronic staphylococcal infections.

Endophytic fungi largely inhabit as biotopes namely in higher plants and therefore have been considered as wellspring of compounds exhibiting diverse activities like anti-microbial, antifungal, anti-tumor, anti-oxidant etc. Endophytic fungi are ubiquitous in healthy tissues amongst all plants and have been accepted a rich source of bioactive compounds^{8,-10}. Metabolites produced by fungal endophytes could be used as an arsenal for pathogenic bacteria as they possess better biosynthetic abilities presumably as a result of their

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gene recombination with the host genes while residing within the healthy plant tissue^{11, 12}. There are several reports on the broad spectrum antimicrobial activity of fungal endophytes from medicinal and higher plants¹²⁻¹⁶ but limited experimental information exists on emphasis of single pathogenic bacteria vis. *Helicobacter pylori⁵*, *Mycobacterium tuberculosis*¹⁷, *Staphylococcus aureus* etc.

Aegle marmelos or Bael (Family, Rutaceae) is a traditional medicinal plant possessing a moderate size and aromatic nature. Bael has enormous traditional uses against various diseases and many bioactive compounds have been identified and isolated from this plant¹⁸. Traditionally the plant has been used for treating intermittent fever, intestinal ailments, fertility control and treatment after childbirth and fish poison¹⁹. The fruit of Aegle marmelos possess antidiarrhoel and anti-dysentery properties leading to its inclusion in the British Pharmacopeia²⁰. Further studies have indicated that the roots of Aegle marmelos also exhibit anti-diarrheal properties, methanolic extract of fruit possess a very strong activity against multidrug resistant Salmonella *typhi*^{21, 22}.

In the present paper we report the evaluation of fermentation extracts of endophytic fungi of the medicinal plant *Aegle marmelos* for its anti-staphylococcal potential and identification of the isolate exhibiting potential anti-staphylococcal activity.

MATERIALS AND METHODS

Plant Sample collection

The plant samples were collected from the heart of conserved rain forest area geographically located at 11°362 183 N 76°042 593 E and 11°592 383 N 77°82 263 E in the Western Ghats region of India during rainy season after seeking due permission from Ministry of Environment and Forests, Govt. of India. The samples were properly sealed in sterilized sample pouches and stored at 4°C till further use.

Isolation of endophytic fungi

For isolation of the endophytic fungi, adult and healthy leaves and stem of *Aegle marmelos* were surface sterilized by aseptically immersing in 1 % sodium hypochlorite solution for

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2-3 minutes followed by dipping in 70 % ethanol for 1-2 min. and finally by washing with 30% ethanol. The surface sterilized samples were then cut into 1 mm pieces with the help of a sterile blade. These 1 mm pieces were inoculated on PDA plates containing 50 μ g/mL of chloramphenicol with the ventral side facing media surface. Maximum of eight pieces were inoculated in single plate. The plates were incubated at 26±2°C for maximum of 25 days¹³. The cultures which appeared on the plate were subsequently transferred to PDA plates containing no antibiotic to obtain pure culture. Endophytes were numbered and stored on PDA slants at 4°C. **Fermentation**

Mycelial plugs of 5mm diameter of 7-days old fungal culture was inoculated in 50 ml presterilized Potato Dextrose Broth (PDB), pH 5.1 (Hi Media) in Erlenmeyer flasks (Schott Duran). The flasks were incubated at shaker at $26^{\circ}\pm 2^{\circ}$ C, 120 rpm for 12 days for production of secondary metabolites²³. After 12 days broth was separated from mycelia by filtration. Filtration was carried out aseptically using a muslin cloth and subsequently through Whatman paper 4²³. Solvent fractionation of the cell free filtrates was carried out using Ethyl acetate. The solvent fraction of different fungal fermentation broths so obtained were freeze dried to obtain residues for testing their antimicrobial activity.

Test Microorganisms

Standard and clinical isolates of *Staphylococus aureus* were used as test organism. S. aureus NCTC 6571 was used a standard isolate while *S. aureus* G3 (Pus Culture, MRSA and VRSA) *S.aureus* G26 (Burn culture) were the clinical isolates tested. Clinical isolates were collected from Govt. Rajindra Medical College, Patiala.

In vitro antimicrobial activity

Agar Well Diffusion Assay, Wells of 5 mm were scooped with the help of pre sterilized cork borer on MH agar (Hi Media) plates to provide a depth of 4 to 5 mm. 30 μ l (Stock of 6.4 mg/ml of each residue) of the cell free filtrate / solvent extract in DMSO was dispensed in the wells. Subsequently solvent along with DMSO used for the dissolution of the residues was also loaded as the control and allowed to diffuse for 15 min. Thereafter wells were sealed with molten MH agar. Finally after, 15 min. the plate was swabbed with 18 24 hrs old 0.5 McFarland adjusted culture of the test isolate. Antibacterial activity was determined by measuring halo formation. All the tests were performed in triplicates²⁴⁻²⁵.

DNA extraction and PCR amplification potential anti-staphylococcal exhibiting fungal endophyte

The fungal DNA was extracted from 3-4 day old culture grown on PDA using the Wizard[®] Genomic DNA purification kit (Promega, USA). 2-3 discs (0.5 mm) of cultured mycelia were cut from 3-4 day old culture and crushed in liquid nitrogen to a fine powder. Further DNA extraction was carried out according to the instructions of kit manufacturer.

The ITS1, 5.8S, ITS2 rDNA sequence was amplified using a Thermocycler (My Cycler, Bio-Rad Laboratories, Inc.) PCR reaction was carried out by using the primers ITS1 (5' TCC GTA GGT GAACCT GCG G 3') AND ITS 4 (5'TCC TCC GCT TAT TGA TAT GC 3'). The PCR reaction was performed in a 25µl reaction mixture containing 1µl of extracted fungal DNA, 10 µM of each primer (ITS1 and ITS4), 2.5mM of dNTP (Bangalore GeNei), 25 mM MgCl₂ (Bangalore GeNei), 1.5 U of Taq DNA Polymerase (Bangalore GeNei) in 10 X Taq buffer (Bangalore GeNei). The PCR cycling conditions consisted of initial denaturation at 96°C for 5 min followed by 39 cycles of 95°C for 45 sec, 60°C for 45 sec, 72°C for 45 sec followed by final extension at 72°C for 5 mins. The PCR products were examined using gel electrophoresis in a 1.5 % agarose gel dissolved in 1X TAE buffer at 40V for 1.30 hr. Gel imaging was performed under UV light in Bio- Rad Gel documentation System using Quantity-1-D analysis software. An approximate 500 bp PCR product was purified by using the Wizard[®] SV Gel and PCR clean up system kit (Promega, USA). Purified PCR products were sequenced (Xcelris Genomics, Xcelris Labs Ltd. Ahemadabad, Gujarat, India) by using 96 capillary high through put sequencer (ABI 3730 XL). Phylogenetic analysis

The ITS/5.8 rRNA sequences obtained from the respective primers were aligned and the consensus sequence was deposited in GenBank NCBI database under the accession numbers JN400741. Further the consensus sequence was subjected to a NCBI-BLAST search to verify the identity of #1005AMLBRT. The sequences were edited with BioEdit 5.0.6 and aligned using MAFFT v 6.240 with other sequences obtained from GenBank. The alignment acquired from MAFFT was submitted to TreeBASE (http,//www.treebase.org) and obtained the submission ID of 11798. GARLI 2.0 (26) was used to perform the maximum likelihood analysis with default parameters except that the number of searches was brought to 5. The resultant best tree having the lowest likelihood ratio was selected and edited in MEGA 4.0. Branch support was estimated by performing 100 bootstrap replicates (27) in GARLI. The resulting trees were fed into PAUP version 4b10²⁸ to obtain a majority rule consensus tree. Bayesian posterior probabilities (PP) for each internodes were calculated with a Metropolis-coupled Markov Chain Monte Carlo (MCMC) sampling method as implemented in MrBayes version 3.1²⁹. Six simultaneous Markov chains were run for one million generations (resulting 10K total trees). The first 2,000 trees were discarded and the remaining 8,000 were used for calculating PP in the majority rule consensus rule tree. These analyses were repeated five times starting from different random trees to ensure trees from the same space were being sampled during each analysis.

RESULTS AND DISCUSSION

25 fungal isolates were recovered from the leaves, stem and internal stem tissue of Aegle marmelos representing 10 endophytic taxa. The distribution of the endophytic fungus varied, maximum isolates were recovered from the internal tissue of the stem. In all six Fusarium species were isolated, two each from leaves, stems and internal stem tissue. One Alternaria species was reported from the leaves of Aegle marmelos. Three endophytic Aureobasidium were also isolated from internal tissue of the stem of Aegle marmelos. (Table 1). 2 isolates of Lasiodiploida sp. were also isolated from internal tissue of the stem of Aegle marmelos. Other isolates included one each of Sphaeropsis sp., Barriopsis sp., Cunninghamella sp. and Penicillium sp. All the cultures were identified on the basis of morphological and cultural characteristics30.

In vitro Anti-Staphylococcal assay of spent broth

Spent PDB of all these fungal isolates were subjected to screening for their antistaphylococcal potential against a panel of three organisms comprising of control and clinical

isolates of *Staphylococci* exhibiting Methicillin and Vancomycin resistance. Only one spent broth of #1005 AMLBRT exhibited antimicrobial activity in the in vitro agar well diffusion assay with zone sizes of 13 mm, 14mm and 15mm respectively against *S. aureus* NCTC6571, Sau G3 and SauG26 respectively (Fig.1).

In vitro assay of ethyl acetate fraction of fungal spent broth for Anti-Staphylococcal assay

Highest broad spectrum antistaphylococcal activity was exhibited by ethyl acetate fraction of spent broth of #1005 AMLBRT (Fig 2). It was followed by #6 AMLWLS> #1079AMSTITYEL > # 9AMLBRT (Fig.3). The diameter of inhibition zone against the test isolates ranged between 18.0-26.7 mm, 17.0-21.7 mm, 14.7-19.7 mm and 12.3-18 mm respectively for #1005 AMLBRT, #6AMLWLS, #1079 AMSTITYEL and #9AMLBRT. 15 endophytic fungal isolates did not exhibit any inhibitory activity in the ethyl acetate fraction of their spent broth against any of the test Staphylococcal isolates. Ethyl acetate extracts of fermentation broth of fungal endophytes from Mallus haliana were found to possess broad spectrum anti-bacterial activity. However *Alternaria brassicicola* ML-P08 was selected from them as it possessed the strongest activity against all the isolates in the test panel³¹.

Phylogenetic identification of #1005 AMLBRT

As #1005 AMLBRT exhibited potential anti-staphylococcal activity, rDNA analysis was carried out using consensus sequence (JN400741) from genomic DNA after amplification of ITS1 and ITS4 primers followed by BLASTN. The BLASTN report revealed close similarity of #1005 AMLBRT to the genus Alternaria (Table 2). Subsequently a data matrix was obtained after aligning 24 BLAST sequences with the consensus sequence of endophyte #1005AMLBRT (JN400741). Maximum likelihood analysis using GARLI 2.0 was conducted using the aligned data matrix comprising of a total of 1023 characters of which 907 (88.7%) were constant, 80 (7.8%) un-informative variable and 36 (3.5%) parsimony informative. The best tree obtained with lowest likelihood ratio of -2155.2073 has been represented in Fig.4. Bootstrap values $(\geq 90\%)$ are based on 100 replicates while PP values are presented in brackets. The branch support is

S.	Endophytic fungi Cultures isolated from different part plants					
No.		Leaf	Stem	Internal tissue of Stem	No.	
1	Fusarium sp.	# 9AMLBRT, #6 AMLWLS	#9(b)AMSTYEL, #7 AMSTYEL	#1070 AMSTITYEL, #1022 AMSTITYEL, #1007 AMLBRT	07	
2	Alternaria sp.	#1005 AMLBRT	-	-	01	
3	Penicillium sp.	-	-	#1011 AMSTITYEL	01	
4	Aureobasidium sp.	-	-	#23(b) AMSTYEL, #11 AMBAWLS, #23 AMSTYEL	03	
5	Lasiodiplodia sp.			#1079 AMSTITYEL, #1104 AMSTITYEL	02	
6	Sphaeropsis sp.			#1003 AMSTITYEL	01	
7	Barriopsis sp.			#1111 AMSTITWLS	01	
8	Cunninghamella sp.			#1032 AMSTITYEL	01	
9	Mycelia-Sterilia			#16 AMLWLS, #1088 AMSTITWLS, #20 AMSTYEL,	03	
10	Unidentified			#1103 AMSTITYEL, #1095 AMSTITWLS, #1082 AMSTITWLS, #32 AMSTYEL, #18 AMSTYEL,	05	

Table 1. Endophytic fungi isolated from leaves and Stem of Aegle marmelos from western Ghats

considered wherein the PP values are higher than 0.95 in the MCMC analysis. The phylogenetic tree reveals the clustering of the characters of #1005 AMLBRT with *Alternaria palandui* (DQ 323687) exhibiting 98% bootstrap values and a posterior probability of 0.98 in one clade.

Morphological studies

Morphological identification #1005 AMLBRT culture was based on the colony or hyphal morphology of the culture on potato dextrose agar, characteristics of spores, discernible reproductive structures. #1005 AMLBRT culture on potato dextrose agar (PDA) exhibited a colony 6 cm diameter after 5-7 days. Colony development on PDA is concentric with obvious rings of sporulation influenced by light/dark cycle. The colony on PDA is so dense as to hide the agar surface. Sporulation is very abundant. Culture medium gradually turns grayish to olivaceous and the substratum turned orange (Fig 5a). The size of a mature conidium body ranges to approximately $35-80 \ \mu m \ x \ 12-19 \ \mu m$. It has a $5-6 \ transverse \ septa$ and one longiseptum in $3 \ or \ 4^{th} \ transverse \ segment$ and is dull brown in colour (Fig 5b).

Molecular tools indicate that #1005 AMLBT is related to Alternaria palandui but morphological studies (macroscopic as well as microscopic) do not confer to the molecular studies. The branch length of #1005 AMLBRT is significantly longer than *Alternaria palandui* indicating molecular difference between the two species. Alternaria palandui does not have growth rings of sporulation influenced by light / dark cycle; the conidial colour is moderate to dark brown with a size of 35-55 x 7-9 µm. Further Alternaria palandui is epiphytic and pathogenic to Allium species. Previously endophytic existence of Alternaria alternata in leaves of Aegle marmelos has been reported purely on morphological and classical taxonomical studies³²

S. No	Accession	Description	Max Score	Query coverage	E value	Max. Identity
1	DQ156344	Alternaria brassicae strain B	897	100%	0	100%
2	AY762949	Alternaria triticimaculans strain EGS 40-150	863	99%	0	97%
3	FJ755199	Alternaria tenuissima strain CZ075A	856	37%	0	96%
4	DQ323687	Alternaria palandui isolate Alt14	879	95%	0	96%
5	AB369424	Alternaria longipes isolate, GL1YS4	875	97%	0.0	96%
6	FR846400	Alternaria compacta strain 15	897	98%	0	95%
7	EU520097	Marssonina mali isolate NW195	863	96%	0	95%
8	AF314583	Alternaria pomicola strain PL1	897	100%	0	94%
9	AF314579	Alternaria citri strain AC2	897	100%	0	94%
10	AY154683	Alternaria mali	901	38%	0	94%
11	AY154712	Alternaria tenuissima strain IA287	901	38%	0	94%
12	AY751457	Alternaria longipes strain EGS30-033	901	99%	0	94%
13	FJ869872	Alternaria brassicae	901	99%	0	94%
14	GQ121322	Alternaria alternata isolate IEIHBT	901	66%	0	94%
15	AY923859	Alternaria aff. dianthicola RHR2	897	100%	0	94%
16	AB470908	Bionectria ochroleuca isolate, X12	899	98%	0.0	94%
17	EF432256	Phomopsis sp. G26	901	89%	0	94%
18	EU 520078	Alternaria gaisen isolate NW576	899	98%	0.0	94%
19	AF455448	Alternaria alternata wb398isolate	901	96%	0.0	94%
20	AF314578	Alternaria abutilonis strain QMT	897	100%	0.0	94%
21	AY154683	Alternaria mali	901	38%	0.0	94%
22	Y17066	Alternaria infectoria isolate 4A	666	88%	0.0	90%
23	Y17069	Alternaria solani isolate ICMP 6519-79	697	99%	0.0	89%
24	Y17086	Alternaria linicola	699	99%	0.0	89%
25	AF229464	Alternaria crassa strain DGG Acr1	702	99%	0.0	89%

Table 2. Similarity pattern of ITS sequence of #1005AMLBRT with genus Alternaria based on BLASTN



(a) S. aureus NCTC6571

(b) S. aureus G3



Fig.1. In vitro anti-staphylococcal assay of whole spent broth of endophytic fungal isolates from Aegle marmelos



(a) S. aureus NCTC6571

(b) S. aureus G3

(c) S. aureus G26

Fig.2. In vitro anti- staphylococcal assay of ethyl acetate fraction from spent broth of #1005 AMLBRT an endophytic fungus from Aegle marmelos

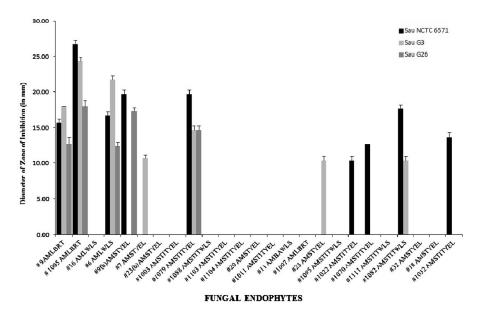


Fig. 3. *In vitro* anti- staphylococcal activity of ethyl acetate fractions of spent broth of endophytic fungi J PURE APPL MICROBIO, **6**(4), DECEMBER 2012.

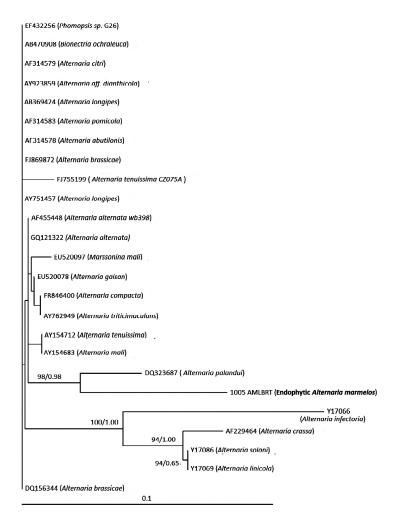


Fig.4. A maximum likelihood (ML) tree generated based ITS and 5.8 S rDNA sequence data (-lnL = 2155.2073). Bootstrap values (equal to or above 90%) based on 100 replicates are shown on the upper branches followed by Posterior probability values from MCMC analyses hyphen

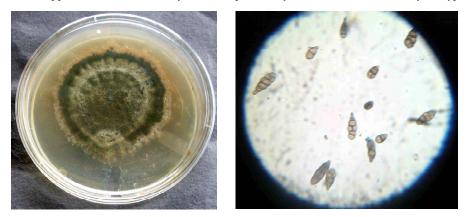


Fig. 5. Morphological features of #1005 AMLBRT (a) 10 day old culture on PDA plate (b) Conidia of #1005AMLBRT at 400x magnification

however phylogenetic studies are necessary to further substantiate identification of endophytic microflora more conclusively.

The present Alternaria isolate #1005AMLBRT from leaves of Aegle marmelos is a new species based on morphological as well as molecular data indicating its non- similarity to Alternaria palandui (Fig 4). #1005AMLBRT is thus named as Alternaria marmelos. Further the production of anti-staphylococcal principles Alternaria elaborated by marmelos (#1005AMLBRT) substantiate the fact that it has overcome the antifungal phytochemicals present in the host plant Aegle marmelos leaves³³. Xanalteric acids I and II have been isolated from an endophytic Alternaria isolate from mangrove plant Sonneratia alba and have been found to possess a weak anti-staphylococcal activity (34). Anti-staphylococcal activity has also been reported from Alternaria alternata existing as an endophyte in Coffea arabica³⁵. Endophytic Alternaria has also been a potential source of antifungal agents to overcome the fungi Plasmopara viticola responsible for downy mildew in grapevine (36). The present study highlights the immense need explore endophytic fungi from medicinal plants and their screen them for their antibiotic potential in developing new drugs. Alternaria species has proved to be resources of newer compounds when it exists as an endophyte. Only one endophytic fungi from bark of Aegle marmelos namely Bartalinia robillardoides Tassi has been found to produce taxol in the free fermentation medium³⁷ till date.

CONCLUSION

Thus the endophyte *Alternaria marmelos* (#1005AMLBRT) from *Aegle marmelos* exhibits potential for further investigations to isolate and characterize the secondary metabolites/bioactive compounds from ethyl acetate extract to overcome multidrug resistant *Staphylococcus aureus*.

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REFERENCES

- 1. Saxena, S. and Gomber, C. Surmounting antimicrobial resistance in the Millennium Superbug, *Staphylococcus aureus*. *Central European Journal of Medicine*, 2010; **5**(1): 12-29.
- 2. Peacock, S.J., de Silva, I., Lowy, F.D. What determines the nasal carriage of *Staphylococcus aureus? Trends Microbiol.*, 2001; **9**: 605-610.
- Fang, G., Keys, T.F., Gentry, L.O., Harris, A.A., Rivera, N., Getz, K. Prosthetic valve endocarditis resulting from nosocomial bacteremia. A prospective, multicenter study. *Ann. Intern. Med.*, 1993; 119(7): 60-67.
- 4. Cookson, B.D. Methicillin-resistant Staphylococcus aureus in the community, new battlefronts, or are the battles lost? Infect. Control Hosp. Epidemiol., 2000; **21**: 398–403
- Parker, M.T., Hewitt, JH. Methicillin resistance in *Staphylococcus aureus*, *Lancet*, 1970; 1: 800-804
- Liu, C. and Chambers, H.F. Staphylococcus aureus with Heterogeneous Resistance to Vancomycin, Epidemiology, Clinical Significance, and Critical Assessment of Diagnostic Methods. Antimicrob. Agents and Chemother., 2003; 47(10): 3040-3045.
- Chopra, I. Antibiotic resistance in Staphylococcus aureus, Causes, concerns and cures? Exp. Rev.Ant. Infect. Ther., 2003, 1(1): 45-55
- 8. Tan, R.X. and Zou, W, X. Endophytes, a rich source of functional metabolites. *Natural Product Reports*, 2001; **18**: 448-459.
- Schulz, B., Boyle, C., Draeger, S. and Römmert, A.K. Endophytic fungi, a source of novel biologically active secondary metabolites. *Mycological Research*, 2002; 106: 996-1004.
- Strobel, G. and Daisy, B. Bioprospecting for microbial endophytes and their natural products. *Microbiology and Molecular Biology Review*, 2003; 67: 491-502.
- 11. Li, Y., Song, Y.C., Liu, Y.M., Tan, R.X. Anti-

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helicobacter pylori substances from endophytic fungus. *World J Microbiol Biotechnol.*, 2005; **21**: 553-558.

- Liu, X., Dong, M., Chen, X., Jiang, H., Lv, X. and Zhow, J. Antimicrobial activity of an endophytic *Xylaria sp.* YX-28 and identification of its antimicrobial compound 7-amino- 4methyl coumarin. *Appl. Microbial Technol.*, 2008; **78**: 241-247.
- Strobel, G., Daisy, B., Castillo, U. and Harper, J. Natural products from microorganisms. J. Nat. Prod., 2004; 67(2): 257-268
- Phongpaichit, S., Rungjindamai, N., Rukachaisirikul, V. and Sakayaroj, J. Antimicrobial activity in cultures of endophytic fungi isolated from *Garcinia* species. *FEMS Immunol. Microbiol.*, 2006; 48: 367-372.
- Kharwar, R.N., Verma, V.C., Kumar, A., Gond, S.K., Harper, J.K., Hess, W.M., Lobkovosky, E., Ma, C., Ren, Y. and Strobel, G.A. Javaerin, an antibacterial napthaquinone from endophytic fungus of Neem, *Chloridium* sp. *Curr. Microbiol.*, 2009; 58: 233-238.
- Selvanathan, S., Indrakumar, I., Muthumary, J. Biodiversity of endophytic fungi isolated from Calotropis gigantea(L.) R.Br. Recent Research in Science & Technology, 2011; 3(4): 94-100.
- de Souza, J.V.B., Lima, A.M., Martins, E.S.J., Salem, J.I.Anti-mycobacterium activity from culture filtrates obtained from the dematiaceous fungus C10. *Journal of Yeast and Fungal Research*, 2011; 2(3): 39-43.
- Maity P, Hansda D, Bandopadhyaya U, Mishra DK.. Biological activities of crude extracts and chemical constituents of bael(*Aegle marmelos*). *Ind. J Exp. Biol.* 2009; 47: 849-861.
- Bsu Da, Sen, R. Alkaloids and coumarins from root bark of *Aegle marmelos*. *Phytochemistry*, 1974; 13: 2329-2330.
- 20. Chopra, R. (1982) Indigenous drugs of India. Academic Publishers. Calcutta.
- Mazumdar, R., Bhattacharya, S., Mazumdar, A., Pattnaik, A.K., Tiwary, P.M., Chaudhary, S. Antidiarrhoeal evaluation of *Aegle marmelos* (Correa) Linn. root extract", *Phytother. Res.*, 2006; 20: 82-84.
- Rani, P. and Khullar, N. Antimicrobial evaluation of some medicinal plants for their anti-enteric potential against multidrug resistant *Salmonella typhi. Phytother. Res.*, 2004, 18(8): 670-673.
- 23. Rodrigues, K.F., Manfred, H. and Christa,W. Antimicrobial activity of secondary metabolites produced by endophytic fungi from *Spondias mombin. J. Basic Microbiol.*, 2000; **40**(4): 261-267.

- Das, K., Tiwari, R.K.S. and Shrivastava, D.K. Techniques for evaluation of medicinal plant products as antimicrobial agent, Current methods and future trends. J. Med. Pl. Res., 2010; 4(2): 104-111.
- 25. Heatley, N.G. Method for the assay of penicillin. *Biochem. J.*, 1944; **38**: 61-65.
- 26. Zwickl, D.J., Genetic algorithm approaches for the phylogenetic analysis of large biological sequence datasets under the maximum likelihood criterion. Ph.D. dissertation, The University of Texas at Austin 2006.
- Felsenstein, J. Confidence limits on phylogenies, An approach using the bootstrap. *Evolution*, 1985; **39**: 783-791.
- Swofford, D.L. (2003). PAUP*, Phylogenetic analysis using parsimony (and other methods) 4.0 b10. Sinauer Associates, Sunderland, MA.
- 29. Huelsenbeck, J.P. and Ronquist, F. MRBAYES, Bayesian inference of phylogeny. *Bioinformatics*, 2001; **17**: 754-755.
- Barnett, H.L. and Hunter, B.B. (1998) Illustrated genera of imperfect fungi. 4th edn. Macmillan, Publ Co, NewYork
- Gu, W. Bioactive metabolites from *Alternaria* brassicicola ML-P08, an endophytic fungus residing in *Malus hallianna*. World J Microbiol. Biotechnol., 2009, 25:1677-1683
- Gond, S.K., Verma, V.C., Kumar, A., Kumar, V., Kharwar, R.N. Study of endophytic fungal community from different parts of *Aegle* marmelos Correae (Rutaceae) from Varanasi (India) World J Microbiol Biotechnol., 2007; 23:1371–1375
- Pitre, S., Srivastava, S.K. Pharmacological, microbiological and phytochemical studies on the roots of *Aegle marmelos*. *Fitoterapia.*,1987; 58: 197.
- 34. Kjer, J., Wray,V., Edrada Ebel, R., Ebel, R., Pretsch, A., Lin, W., Proksch, P. Xanalteric acids I and II and related compounds from endophytic *Alternaria* sp. isolated from the Mangrove plant *Sonneratia alba. J.Nat. Prod.*, 2009; 72(11): 2053-2057
- Fernandes, M.D.R.V., Silva, T.A.C., Pfenning, L.H. Biological activities of the fermentation extract of the endophytic fungus *Alternaria alternata* isolated from *Coffea arabica* L. *Brazilian J. Pharmaceutical Sciences*, 2009, 45(4): 677–685.
- 36. Musetti, R., Vecchione, A., Stringher, L., Borselli, S., Zulini, L., Marzani, C., D'Ambrosio, M., di Toppi, L.S. and Pertot, I. Inhibition of sporulation and ultrastructural alterations of grapevine downy mildew by the

endophytic fungus *Alternaria alternata*. *Phytopathology*, 2006, 96: 689-698.

37. Gangadevi, V. and Muthumary, J. Taxol, an anticancer drug produced by an endophytic

fungus *Bartalinia robillardoides* Tassi isolated from a medicinal plant *Aegle marmelos* Correa ex Roxb. *World J Microbiol. Biotechnol.*, 2008: **24**: 717-724.