

Water Quality Deterioration and its Socio-economic Implications

Sibanda Timothy and Okoh I. Anthony*

Applied and Environmental Microbiology Research Group (AEMREG),
Department of Biochemistry and Microbiology, Faculty of Science and Agriculture,
University of Fort Hare. Private Bag X1314, Alice 5700, South Africa.

(Received: 29 September 2012; accepted: 10 November 2012)

From ancient times, people have chosen to live close to water sources, settling in river valleys. Improved water supply and water resources management boosts countries' economic growth and contributes greatly to poverty eradication. An adequate supply of safe drinking water is one of the major prerequisites for a healthy life but, because of surface water pollution, waterborne disease became, and still is, a major cause of death in many parts of the world, particularly in children. Rapid urbanisation has exacerbated surface water pollution by increasing point pollution and non-point source pollution entering surface waterbodies. Physico-chemical and microbiological properties of water are used to assess water quality as they give a good impression of the status, productivity and sustainability of waterbodies. Water pollution control has been a matter of public concern for more than a century. Currently, human beings and natural ecosystems in many river basins suffer from debilitating effects of water pollution. Hence, development of better water conservation practices and policies are critical to the sustenance of our water both in terms of quantity and quality to ensure protection of public health.

Key words: Freshwater, Water quality, Pollution, Public health, Physico-chemical, Microbiological.

From ancient times, people have chosen to live near water, settling in river valleys, beside lakes, or along coastlines ¹. Without freshwater of adequate quantity and quality, sustainable development will not be possible ². Water quality reflects the composition of water as affected by natural causes and man's cultural activities expressed in terms of measurable quantities and related to intended water use ². Worldwide, waterbodies are the primary dumpsites for disposal of waste, especially effluents from industries located near them³. Effluents from industries harbour toxic contents, capable of altering the physical, chemical and biological nature of the

receiving waterbodies ^{4,5}. First to be degraded by such waste is the physical quality of the water, while the biological degradation becomes evident later in terms of number, variety and organisation of the living organisms in the water ⁶. DWAF ⁷ defined water pollution as the alteration of the properties of a water resource so as to make it, among others, "harmful or potentially harmful to the welfare, health or safety of human beings". Much of the current concern with regards to environmental quality is focused on water because of its importance in maintaining human health and health of the ecosystem ⁸.

Surface water stress and vulnerability

Water pollution occurs when unwanted or toxic substances are accidentally or intentionally introduced into waterbodies in quantities which affect the resource in providing its services, which include domestic use, irrigation, navigation, recreational and life support (ecological) functions

* To whom all correspondence should be addressed.
E-mail: aokoh@ufh.ac.za

⁹. Water stress and vulnerability are linked, since pollution reduces the volume of water available for human use ¹. Thus, preventing pollution is among the most cost-effective means of increasing water supplies.

Rapid urbanisation has exacerbated surface water pollution by increasing point source pollution and non-point source pollution entering surface waterbodies ¹⁰. Both the withdrawal of surface water for human use and economic activities and its subsequent discharge back into surface water resources as effluents can affect the ability of aquatic ecosystems to survive ¹⁰. Such effluents have been reported to also include antibiotics and other pharmaceutical compounds ¹¹. The occurrence of antibiotics in aquatic environments is of ecotoxicological concern because of potential ecosystem alteration ¹². Prolonged exposure to low doses of antibiotics leads to the selective proliferation of resistant bacteria, which could transfer the resistance genes to other bacterial species ¹³.

Water quality problems and their effects are different in type and magnitude in developed and developing countries, particularly those stemming from microbial and pathogen content ¹⁴. Currently, human beings and natural ecosystems in many river basins suffer from water scarcity. In global-scale assessments, basins with water stress are defined either as having a per capita water availability below 1,000m³/yr (based on long-term average runoff) or as having a ratio of withdrawals to long-term average annual runoff above 0.4 ¹⁴. Populations living in such severely stressed basins are estimated to range from 1.4 billion to 2.1 billion¹⁴.

Indicators of surface water pollution

Reporting on the endemic water pollution problem in Zimbabwe, The Herald ⁹ had this to say, "The presence of pollutants in water is primarily perpetrated by human activities. These pollutants are either discharged directly into rivers or are carried into streams and rivers by surface runoff, leading to serious deterioration of water quality. Water pollution can be categorised into four broad categories viz: organic pollution which occurs when excess of organic matter, such as manure or sewage, enters the water; toxic pollution which occurs when a chemical pollutant that is not a natural component of an aquatic ecosystem is

introduced; thermal pollution which occurs when water is used as a coolant near a power or industrial plant and then returned to the aquatic environment at a higher temperature than it was originally and ecological pollution which takes place when chemical pollution, organic pollution or thermal pollution are caused by nature rather than by human activity." The presence of pollutants belonging to any of the above categories can be ascertained by periodically assessing the physicochemical and microbiological properties of in-stream water ¹⁵.

Physicochemical parameters

Dissolved constituents of waterbodies are often determined as a major component for baseline limnological studies ¹⁶. Physico-chemical properties of water are used to assess water quality as they give a good impression of the status, productivity and sustainability of such waterbodies¹⁷. Physico-chemical determinants of pollution in rivers, lakes, and oceans include temperature, pH, total dissolved solids (TDS), turbidity, electrical conductivity (EC), biological oxygen demand (BOD), chemical oxygen demand (COD), dissolved oxygen (DO), heavy metals and nutrients amongst others ^{15; 18}. These pollutants originate either from point or non-point sources ¹. DO is the most important factor in the assessment of water quality and is vital for aquatic life ¹⁸ while temperature is the most important physical variable affecting the metabolic rate of aquatic microorganisms ^{19; 20} as well as the chemical reactions in water, thereby determining the solubility of gases (including oxygen) and imparting taste and odour to the water ²¹. Healthy freshwater bodies are characterised by a DO concentration of at least 5 mg/L ²². Surface water temperature is directly affected by changes in ambient air temperatures and indirectly by the inflow of water of a different temperature ²³, characteristic of discharge of large volumes of water from industrial plants. The pH of natural waters range from less than 4 to greater than 12, but usually falls between 6 and 9 for unpolluted river systems ²⁴. pH values above and below this range are indicative of water pollution ¹⁸; and could give rise to toxic effects, largely as a result of disturbances in internal ion homeostasis ²³. High concentrations of dissolved phosphate may lead to osmotic stress, as is the case with high nitrate concentrations ²⁵. Even

though trace quantities of phosphorous are naturally present in surface waters ²¹, higher concentrations of phosphate could be indicative of pollution from domestic waste and agricultural runoff, and may lead to eutrophication, which has drastic economic, social and ecological consequences ²¹. High levels of organic pollution can also result in low DO and high BOD and COD concentrations ²⁶. High turbidity is harmful to aquatic organisms since it can cause anaerobic conditions, interfere with respiration in aquatic fauna and also reduce light penetration, hindering photosynthesis and natural aquatic life ²⁷. EC is directly related to TDS in water and its value becomes greater with increasing degree of pollution ²⁸.

Faecal indicator bacteria

Faecal indicator bacteria (FIB) have been used for many years to determine the quality and safety of surface and ground waters ^{29; 30}. Bacterial groups classified as FIB include the total coliforms (TC), faecal coliforms (FC) and enterococci (synonymously used as faecal streptococci) ³¹. Faecal streptococci have been suggested as the recommended indicator for salt water while either faecal streptococci or *Escherichia coli* can be used for monitoring freshwaters ³¹. Faecal streptococci are widely accepted as useful indicators of faecal pollution in natural aquatic ecosystems because they show a close relationship with gastrointestinal symptoms associated with bathing in marine and freshwater environments ^{32; 33} while their persistence patterns are also similar to those of potential water-borne pathogenic bacteria ³⁴.

FC (also known as thermotolerant coliforms) include strains of the genera *Klebsiella* and *Escherichia* ³⁵. It has been suggested that for the purpose of sanitary water testing, *E. coli* should be used as an indicator of faecal pollution since it possesses a more direct and closer relationship with homeothermic faecal pollution ³⁶. However, *E. coli* has been detected in some pristine areas ³⁷ and has also been associated with regrowth in drinking water distribution systems ³⁸. The presence of these bacteria in surface waters is thought to indicate that pathogenic organisms such as *Salmonella* spp., *Shigella* spp. and hepatitis A may also be present ³⁰. Polo *et al.* ³⁹ reported incidences of serotypes of *Salmonella* spp. isolated from freshwater sources in Spain which were identical to serotypes found in clinical

samples, a case that underlines the connection between water quality and public health. However, epidemiological studies in warm tropical waters demonstrate the lack of a strong relationship between faecal indicators and health outcomes, in part, due to the inappropriate nature of *E. coli* or faecal streptococci as indicators of waterborne pathogens in these recreational waters ³¹. Alternatively, spores of *Clostridium perfringens* have been proposed as a useful indicator when fresh faecal contamination is being investigated ⁴⁰ and more importantly, as suitable indicators for parasitic protozoa and viruses in sewage-impacted waters ⁴¹. Bacteriophages have also been suggested as indicators specific for human sewage, and more specifically *Bacteroides fragilis* phages which appear to survive in a similar manner to that of human enteric viruses under a range of conditions ⁴².

Contamination of surface waters with faecally derived bacteria can occur through point sources like sewage effluents and non-point sources such as agricultural and urban run-off ⁴³. Sewage effluents contain a wide variety of pathogenic microorganisms that may pose a health hazard to the human population when discharged into recreational waters ^{31; 44}. The density and variety of these pathogens are related to the size of the human population, the seasonal incidence of the illness, and dissemination of pathogens within the community ⁴⁵. Studies also prove that bathers can be a significant source of pathogenic microorganisms, even in cases where there is no faecal pollution from the outside ³¹. Storm events and recreational activity also cause the re-suspension of FIBs resulting in a rapid increase in the load of pathogens in the water phase ⁴³. The U.S. Environmental Protection Agency ⁴⁶ reported that 35% of impaired rivers and streams were polluted by FIB which could indicate the presence of enteric pathogens.

Water-borne enteric viruses

Faecal matter of patients suffering from virally-induced gastroenteritis contains high concentrations of human enteric viruses which, if not inactivated during wastewater treatment processes may contaminate surface water sources for drinking water, recreational activities, aquaculture and irrigation ^{47; 48; 49; 50; 51}. Human enteric viruses include the families *Picornaviridae*

(these are enteroviruses like poliovirus, coxsackievirus, and echovirus), *Adenoviridae*, *Caliciviridae* (norovirus, calicivirus), *Astroviridae*, and *Reoviridae* (reovirus, rotavirus)⁵². They cause a wide range of diseases which include epidemic gastroenteritis, meningitis, ocular and respiratory infection, paralysis, myocarditis and hepatitis⁵³.

Enteric viral pathogens have been shown to be present in environmental waters even when bacterial indicators are absent^{54; 55; 56}. Studies focusing on non-enteric viruses like bacteriophages, viruses infecting algae, protozoa, fish and vascular plants⁵⁷ have found that the dynamics of these viruses are linked to the dynamics of potential host cells (primarily bacteria and algae)^{58; 59}. The abundance of these viruses has been shown to peak following an increase in host cell abundance^{60; 61}. However, enteric viruses are obligate intracellular host-specific parasites which cannot grow or survive for extended periods in environmental waters outside their host. These viruses are very small, ranging from 20-70 nm in diameter and consist of a nucleic acid genome surrounded by a protein capsid and, in some cases, a lipoprotein envelope⁶². Survival and/or persistence of enteric viruses in the environment is strongly linked to various environmental factors like exposure to ultra-violet radiation, temperature^{63; 64; 65}, adsorption to particulate material^{66; 67} and salinity^{68; 69; 70}. Their susceptibility to the different environmental factors may, however, vary profoundly within the virosphere as viruses are capable of developing resistance mechanisms to survive in harsh habitats including hypersaline waters and hot springs or hydrothermal vents^{71; 72; 73; 74}.

Viral persistence in tropical freshwater environment

While specific viruses or strains of viruses are not always present in a community at any one time, representatives of the large groups are, however, generally present on most occasions⁶². Enteric viruses may be found in high numbers in domestic wastewater, their numbers generally varying with the level of virus infection in the community⁷⁵. Wastewater treatment processes that do not include a disinfection step are often inefficient in removing viruses⁶². In previous studies, levels of human viral

contamination in sewage and wastewater treatment plants were analysed^{76; 77; 78; 79; 80; 81}. All these studies reported high concentrations of viruses in sewage. Viruses outside a host are inert particles possessing no intrinsic metabolism and do not require any nutrients to persist⁸². They are, however, resilient enough to survive in the environment for long periods of time and still retain their infectivity during the various conditions that they may encounter between one host and another^{83; 84; 85}. This is illustrated by the number of outbreaks of enteric viral diseases attributable to waterborne transmission⁸⁶. Rzezutka and Cook⁸² reviewed works previously done by other researchers on enteroviruses (polio-, echo- and coxsackieviruses). Summarising the observations from these studies and grouping them into freshwater sources gave mean viral inactivation rates of: 0.576 log₁₀ d⁻¹ (tap water); 0.325 log₁₀ d⁻¹ (polluted river water); 0.25 log₁₀ d⁻¹ (unpolluted river water); 0.374 log₁₀ d⁻¹ (impounded water); and 0.174 log₁₀ d⁻¹ (ground water). These rates were all less than 1 log₁₀ per day, and indicated that viruses could survive in freshwater sources for prolonged periods of time.

Viruses have been found to be inactivated by prolonged holding in reservoirs exposed to sunlight, elevated temperature and extremes of pH⁸⁷. A study carried out by Phanuwat *et al.*⁴⁸ in Jakarta, Indonesia, showed a statistically significant correlation between the physicochemical parameters (including conductivity, turbidity, temperature and total dissolved solids [TDS]) with all viruses tested (enterovirus, hepatitis A virus, Norovirus GI & GII and adenovirus). Turbidity showed positive correlation with all the microbes tested while conductivity, temperature, TDS showed negative correlation. The same study also showed a high prevalence of enteric viruses in floodwater compared to river water. While the authors did not explain this phenomenon, possible explanation could be that floods may cause the overflow of sewage treatment plants carrying with them large amounts of untreated and partially treated faecal matter. The erosive power of a flood causes the flood waters to be very muddy (turbid) thereby shielding the viruses (especially the RNA viruses) from the damaging effects of UV-rays of the sun.

In a study done by de Cardona *et al.*⁸⁸ in a tropical lagoon, they found that virus inactivation

rates were significantly higher in that tropical lagoon than in temperate areas, probably due to higher temperature and salinity. Hurst *et al.*⁸⁹ examined the long-term survival of coxsackievirus B3, echovirus 7 and poliovirus 1 in samples of surface freshwater collected from five sites of physically different characteristics (artificial lake, small groundwater outlet pond, large- and medium-sized river and a small suburban creek). Survival was studied at temperatures of -20°C , 1°C and 22°C . The average viral inactivation was 6.5–7.0 \log_{10} units over 8 weeks at 22°C , 4–5 \log_{10} units over 12 weeks at 1°C and 0.4–0.8 \log_{10} units over 12 weeks at -20°C . Several physical and chemical parameters (hardness and conductivity) appeared detrimental to virus survival. The turbidity of the water and suspended solids represented a beneficial influence for virus survival. These findings concurred to a large extent with the work of Phanuwat *et al.*⁴⁸ which was done in Jakarta, Indonesia where turbidity was found to enhance viral survival rates while sunlight and high temperatures were detrimental to virus survival.

Two particularly notable factors that have been associated with the level of human enteric viruses in freshwaters are seasonal changes in water temperature⁹⁰ and a “rainy season” effect observed by Keswick *et al.*⁹¹. Water temperature exerts an extremely strong influence on viral stability, lower temperatures increasing the survival time⁸⁷. The “rainy season” effect arises because of the very high turbidity that characterises freshwater bodies at that time of the year, which correlates in a statistically significant manner with the presence of indigenous viruses in water and with virus stability in water under laboratory conditions⁸⁷. In another study by Espinosa *et al.*⁸⁵ in Mexico City (tropical highland), they reported that the presence of enteroviruses (EVs) and rotaviruses (RVs) was significantly more frequent during the cold-dry season (0.75 and 0.35, respectively) with an average low temperature of 4°C and an average precipitation of less than 10 mm than in the warm-rainy season (0.10 and 0.05, respectively) whereas astrovirus showed no significant relationship with the environmental variables recorded (pH, temperature, conductivity and dissolved oxygen concentration). This study also indicated that enteric viruses could be damaged by rising temperatures, as reported

previously^{87,92}, when EVs and RVs were studied in freshwater at 22°C and 20°C .

Determination of virus infectivity

Virus infectivity is generally believed to provide more relevant estimates of virus decay than disappearance of viral particles^{93; 94; 95}. Tropical phages may be presumed to be genetically adapted to protect DNA and capsids against UV damage because of their capsid structure⁹⁶, or the dimerisation in DNA that may reduce the susceptibility of destructive enzymes⁹⁷. Experiments on the persistence and infectivity of phage isolates have demonstrated that temperature is a strong determinant of both⁹⁸. Suttle and Chen⁶⁷ and Noble and Fuhrman⁹³ also showed that temperature notably affects the decay of virus infectivity.

While molecular assays such as direct reverse transcription-polymerase chain reaction (RT-PCR) are sensitive, they provide inaccurate estimates of infectious viruses in the environment because they detect both inactivated and infectious virus particles⁹⁹. On the other hand, cell-culture based techniques reveals infectivity of viruses and consequently risk of illness to water consumers^{100; 101}. In a study done by Greening and co-workers⁹⁹, whose aim was to establish C-PCR methods for detection of culturable enteroviruses and adenoviruses in a broad range of environmental samples, and calibrate these methods against direct RT-PCR, PCR and plaque assay methods for sensitivity, as well as speed and ability to provide virus infectivity data, observed that direct RT-PCR detected 0.05–0.2 pfu/RT-PCR and was 10–100 times more sensitive than other methods but did not provide information on infectivity. Results for adenovirus also indicated that the direct PCR was 10 times more sensitive than C-PCR and detected 0.16 pfuD PCR, but did not give information on infectivity. They concluded that while direct RT-PCR or PCR methods are the most sensitive methods, their main disadvantage is the inability to provide information on infectivity. This limits their use in environmental virology applications where it is important to ascertain virus infectivity. On the contrary, they found that the C-PCR assay provided sensitive detection and confirmation of infectious enteroviruses and adenoviruses within 2–5 days of sampling. However, infectivity can be inferred for certain uncultivable RNA viruses (norovirus,

enteroviruses, Hepatitis A and E) from molecular detection data where the viruses have been subjected to chemical but not UV disinfection⁹⁹. Murrin *et al.*¹⁰⁰ also suggested that while the presence of viral DNA does not necessarily indicate the presence of infectious viruses, virus viability is inferred whenever virus nucleic acid is detected because the nucleic acids, single stranded RNAs in particular, are extremely susceptible to degradation in the environment.

Health concerns

Enteric viruses are important waterborne pathogens which are frequently isolated from faecally contaminated water and have been linked to numerous waterborne outbreaks^{102; 50; 103}. Discharge of effluents from wastewater treatment plants into rivers that are used as source water in drinking water treatment plants (DWTPs) could present a risk of infection in the population if efficient drinking-water treatment is not applied and properly controlled before tap water distribution and consumption¹⁰⁴. The presence of viruses and other pathogens in the environment is an indicator of faecal pollution that poses a potential risk to the exposed population, since such pathogens do not constitute normal gastrointestinal microbiota, and are only excreted by sick individuals¹⁰⁵. Human adenoviruses are present at a higher frequency in sewage compared to other enteric viruses⁸¹ and are excreted in high concentrations of up to 10¹¹ viral particles per gram of faeces from infected patients¹⁰⁶. Adenoviruses are second only to rotaviruses as major etiologic agents of infantile gastroenteritis¹⁰⁷⁻¹¹⁰ causing a variety of clinical manifestations associated with the gastrointestinal, respiratory and urinary tracts, as well as the eyes¹¹¹. Adenoviruses are ubiquitous in water environments and these viruses are exceptionally resistant to purification and disinfection processes¹¹². Enteric human adenoviruses (HAd) have a double-strand DNA genome which is more resistant to UV-light than the single strand RNA of other enteric viruses such as polio and hepatitis A viruses¹¹³. The occurrence of Ad in treated drinking water and tap water has been reported in South Korea and South Africa¹¹⁴. Health outcomes attributed to Ad infection include enteric related illnesses, respiratory system, eye infections and fatal outcome for immunocompromised patients and organ and bone

marrow transplant recipients¹¹⁵. The consumption of clams harvested from a sewage-polluted area¹¹⁶ also exposes people to risk of virus-related food poisoning, especially debilitating infectious hepatitis which may also lead to death. Numerous outbreaks of HAV infection have been reported worldwide¹¹⁷⁻¹²³ with the most severe occurring in Shanghai, China in 1988¹²⁴. Viral contamination of wastewater, recreational water, drinking water, irrigation water, ground or subsurface water have been reported frequently as a primary source of gastro-enteritis or hepatitis outbreaks¹²⁵⁻¹³¹.

Water quality as an economic growth determinant

Improved water supply and sanitation and water resources management boosts countries' economic growth and contributes greatly to poverty eradication¹³². Economic growth itself can also drive increasing investments in improved water management and services, initiating a virtuous cycle that improves the lives of the people across socio-economic boundaries¹³². The South African Department of Water Affairs and Forestry²⁵ defined the term *water quality* as the physical, chemical, biological and aesthetic properties of water that determine its fitness for a variety of uses and for the protection of aquatic ecosystems. Freshwater resources have important social and economic benefits as a result of tourism and recreation, and are culturally and aesthetically important for people throughout the world¹⁶. According to CSIR¹³³, a healthy society and productive workforce play an important role in long-term economic growth and sustainable development. Water pollution therefore causes not only the deterioration of water quality, but also threatens human health, the balance of aquatic ecosystems, economic development and social prosperity¹³⁴.

Water quality and agriculture: Irrigation water

Typical sources of agricultural water include surface water, groundwater, and municipal supplies¹³⁵. Sewage spills, runoff from concentrated animal production facilities, storm-related contamination of surface waters, illicit discharge of waste, and other sources of pathogens threaten the quality of both surface water and groundwater used for fruit and vegetable production and therefore the safety of the consumed product¹³⁶.

DWAF²⁵ defines irrigation water as water which is used to supply the water requirements of

crops and plants which are not provided for by rain, and refers to all uses water may be put to including water for the production of commercial crops; irrigation water application and distribution systems; home gardening; the production of commercial floricultural crops and potted plants.

Whenever water comes into contact with produce, its source and quality are directly linked to the potential for contamination ¹³⁵. These

potential contaminants are classified into microbiological (bacteria, virus, and protozoa); chemical, and physical agents ¹³⁵. Chemical and physical properties of irrigation water are of paramount importance because they affect crop yield and soil physical conditions; fertility needs; irrigation system performance and longevity ¹³⁷. Some of the chemical agents of concern in irrigation water are listed in Table 1.

Table 1. Guidelines for nutrient concentrations in irrigation water (mg/L)

Macronutrient	Low	Normal	High	Very High
Nitrate	<5	5-50	50-100	>100
Ammonium	<2	2-75	75-100	>100
Phosphorous	<0.01	0.1-0.4	0.4-0.8	>0.8
Potassium	<5	5-20	20-30	>30
Calcium	<20	20-60	60-80	>80
Magnesium	<10	10-25	25-35	>35
Micronutrient	Acceptable range Suggested maximum concentration			
Iron	2.4-4.0	5.0		
Manganese	<0.2	0.2		
Copper	<0.2	0.2		
Zinc	<0.3	2.0		
Boron	<2.0	2.0		

Adapted from Landschoot, ¹³⁸.

However, current data from the Center for Disease Control and Prevention (CDC) shows that 90% of foodborne illnesses come from microbiological agents ¹³⁵. In the 1990s, the CDC

estimated that up to 12% of reported foodborne illness outbreaks were linked to fresh produce ¹³⁵. Table 2 shows some of the pathogens that have been associated with fresh produce since the 1990s.

Table 2. Selected confirmed multiple outbreaks of foodborne pathogens associated with fresh produce since the 1990s

Produce	Pathogens/chemical
Cantaloupe	<i>Salmonella</i> spp./ <i>E. coli</i> O157:H7
Raspberries	<i>Cyclospora cayatenensis</i>
Tomatoes	<i>Salmonella</i> spp.
Basil	<i>Cyclospora cayatenensis</i>
Parsley	<i>Shigella</i> spp.
Green onions/scallions	Hepatitis A virus, <i>Shigella</i> spp.
Various berries	<i>Cyclospora cayatenensis</i>
Lettuce	<i>E. coli</i> O157:H7
Cabbage	<i>L. monocytogenes</i>
Watermelon	<i>Salmonella</i> spp./ Aldicarb

Adapted from Simonne, ¹³⁵.

Most victims of foodborne illnesses contract the diseases either through the ingestion of contaminated water or by eating minimally

processed or raw vegetables that were irrigated with contaminated water ¹³⁹. Apart from the concern for the safety of consumers, there is also concern

over the safety of pickers, handlers, packers and farmers that participate in the production of vegetables during pre-harvest and post-harvest, especially young children from families of farming communities who tend to be most vulnerable to

salmonella infection as a result of contaminated irrigation water ^{139; 140}. Table 3 shows the trigger values for faecal coliforms in irrigation waters used for food and non-food crops.

Table 3. Trigger values for faecal coliforms in irrigation waters used for food and non-food crops.

Intended use	Level of faecal coliforms
Raw human food crops in direct contact with irrigation water (e.g. via sprays, irrigation of salad vegetables)	<10 CFU/100 mL
Raw human food crops not in direct contact with irrigation water (edible product separated from contact with water, e.g. by peel, use of trickle irrigation); or crops sold to consumers cooked or processed	<1000 CFU/100 mL
Pasture and fodder for dairy animals (without withholding period)	<100 CFU/100 mL
Pasture and fodder for dairy animals (with withholding period of 5days)	<1000 CFU/100 mL
Pasture and fodder (for grazing animals except pigs and dairy animals, i.e. cattle, sheep and goats)	<1000 CFU/100 mL
Silviculture, turf, cotton, etc. (restricted public access)	<10 000 CFU/100 mL

For the protection of public health, WHO ¹⁴² set a bacterial guideline of ≤ 1000 faecal coliforms (FC) per 100 ml for unrestricted irrigation. This figure was reached at after data on pathogen removal by efficient wastewater treatment plants showed that at an effluent concentration of 1000 FC/100 ml, which reflects >99.99% removal, bacterial pathogens would have been eliminated and viruses would be at very low levels ¹⁴³.

Water quality and public health

Most river stretches are used for various activities such as bathing, drinking, municipal water supply, navigation, irrigation, fishing and recreation ¹⁴⁴. Simultaneously, they are also used as recipients for discharge of industrial effluent, municipal sewage and dumping of solid wastes, which can potentially contaminate surface- and ground water resources ¹⁴⁴. An adequate supply of safe drinking water is one of the major prerequisites for a healthy life ¹⁴⁵, but waterborne disease is still a major cause of death in many parts of the world, particularly in children, and it is also a significant economic constraint in many subsistence economies ¹⁴⁶. There is a clear link between the state of the environment and human health and well-being ¹³³. For example, the run-off of nutrients to surface waters, often combined with sewage discharges, leads to significant growths of cyanobacteria which can produce a wide range of

toxins ¹⁴⁷ and, where drinking water treatment is limited or non-existent, there is a potential for undesirable concentrations to be present in drinking water ¹⁴⁶. The abundance of organic compounds, radionuclides, toxic chemicals, nitrites and nitrates in water may cause unfavourable effects on the human health especially cancer, other human body malfunctions and chronic illnesses ¹⁶. The World Health Organisation ¹⁴⁸ estimates that 23% of all deaths in Africa are the result of avoidable environmental hazards such as contaminated water, poor hygiene, inadequate sanitation and poor water resource management among others. According to DWAF ⁷, WHO recognises diarrhoeal diseases as the leading cause of death in developing countries where many communities are still relying on untreated water from surface resources for their daily supply, with limited or no access to adequate sanitation facilities. The lack of access to safe water, basic sanitation and good hygiene is the third most significant risk factor for poor health in developing countries with high mortality rates ¹³². In South Africa alone, it had been estimated that as many as 43 000 people might die annually as a result of diarrhoeal diseases ⁷. The microbiological quality of recreational water bodies is of utmost public health concern since some population groups such as the very young, the elderly, the

immunocompromised and tourists might be more susceptible to local endemic pathogens and, thus, may be at higher risk to swimming-associated disease ¹⁴⁹. Children are clearly at higher risk because of their swimming behaviour and immature immune systems, while visiting populations may be at higher risk because they have not been previously exposed to local pathogens ¹⁴⁹. For full body contact recreational waters, USEPA ¹⁵⁰ suggested that the geometric mean of bacterial densities should not exceed 126 CFU/100 ml for *E. coli* or 33 CFU/100 ml for Enterococci.

Pollution burden of surface water resources: South Africa as a case in view

South Africa is the 30th driest country in the world ¹⁵¹ and its available freshwater resources are already almost fully-utilised and under stress ¹⁵². At the projected population growth and economic development rates, it is unlikely that the projected demand on water resources in South Africa will be sustainable ¹⁵³. Water is increasingly becoming the limiting resource in South Africa ¹⁵⁴, and supply will become a major restriction to the future socio-economic development of the country, in terms of both the quantity and quality of available water ¹⁵⁵. Predictions are that South Africa as a whole is likely to have a water deficit of approximately 1.7% by 2025 ^{156; 157; 158}. A 2010 report issued after experts sat down to roundtable talks with Business Leadership South Africa (BLSA) and the Centre for Development and Enterprise (CDE) (an independent policy research and advocacy organisation that focuses on critical national development issues and their relationship to economic growth and democratic consolidation) issued the following warning...

“On current trends, South Africans may one day have to make do with significantly less water per capita. For a country already using almost all its available water resources, this would be a dramatic change, with far-reaching implications for households, businesses, communities and government,” ¹⁵⁹.

South Africa is located in a predominantly semi-arid part of the world whose climate varies from desert and semi-desert in the west to sub-humid along the eastern coastal area, with an average rainfall for the country of about 450 mm per year, well below the world average of about 860 mm per year ¹⁶⁰. Only 8.5% of this low average

annual rainfall finds its way to rivers as runoff. The total annual surface runoff of South Africa is $1.5 \times 10^6 \text{ m}^3/\text{a}$ ¹⁶¹ which is less than half that of the Zambezi River ¹⁵³. South Africa's local geology of hard rocks also means there are few exploitable aquifers ¹⁵³ while water which is naturally of poor quality also occurs in some areas, which limits its utilisation ¹⁶⁰. Because of the spatial variability of water resources and the scarcity of water throughout the country, the need for water far exceeds supply in many catchments ¹⁵⁵. While the discrepancies in the water situation of different catchments have been managed through inter-basin water transfers, South Africa cannot afford to build more dams and water transfer schemes as they cost large amounts of money ¹⁵⁸. This situation is likely to worsen as the discrepancies between water requirements and availability in other water-scarce catchments increase. As of 2003, the available surface water resources in South Africa had been fully utilised, with an estimated water deficit of about 600 million m^3 per year ¹⁶². Groundwater resources were said to account for about 350 million m^3 per year, reducing the overall water deficit to about 250 million m^3 per year ¹⁶³.

South Africa's scarce freshwater resources are also decreasing in quality because of an increase in pollution and the destruction of river catchments ¹⁵⁸. Rivers play a major role in assimilating or carrying of industrial and municipal wastewater, manure discharges and runoff from agricultural fields, roadways and streets, which are responsible for river pollution ¹⁶⁴. Typical pollutants of South Africa's freshwater environment include industrial effluents, domestic and commercial sewage, acid mine drainage, agricultural runoff, and litter. At one point, freshwater pollution (in the form of Chemical Oxygen Demand) was estimated to be $4.74 \text{ ton}/\text{km}^3$ while the average phosphorous concentrations (as orthophosphate) were estimated at $0.73 \text{ mg}/\text{L}$; values which indicated that South Africa's freshwater resources could be excessively enriched and may be considered to be moderately to highly eutrophic ¹⁶⁵. Access to water was one of the key needs identified by poor communities in 1994, as well as jobs, housing, health care and education ¹⁶⁶. Between 1996 and 2009, the share of South Africa's households with access to clean water rose from 62 percent to 92 percent, inclusive of

shared neighborhood taps¹⁵⁹. However, population growth and economic growth are regarded as the primary determinants with respect to future water requirements, both scenarios for which deficits are generally projected to increase and surpluses to diminish¹⁶⁰. This necessitates better understanding, management and mitigation of pollution issues to help the situation.

Global perspectives on water quality: legislative approach

Water pollution control has been a matter of public concern for more than a century¹⁶⁷. The World Health Organization (WHO) has been proactive in this aspect¹⁶⁸; developing important guidelines of universal application and has, in recent years, promoted a more preventive approach dubbed the Water Safety Plans¹⁶⁹, which takes

into account all factors that endanger the quality of drinking water from the source to the consumer¹⁶⁸. It has been proven that reactive measures to clean up polluted sites and water bodies are generally much more expensive than pro-active measures to prevent pollution from occurring¹⁷⁰. It is for this reason that national governments the world-over have been formulating landmark pieces of legislation to safe-guard both the quality and quantity of water available to their citizens. Examples are shown in Box 1 to Box 4.

As more information is disseminated and public awareness of water quality issues increases, national governments continue to develop and enforce better programs aimed at the preservation of water resources. In the face of competing water uses like growing populations, energy production,

Box 2.1: Water quality governance in Zimbabwe

In Zimbabwe, the Environmental Management Act Chapter 20:27 and Statutory Instrument 6 of 2007 are used by the Environmental Management Agency to advocate for the application of the "Polluter Pays" principle in its entirety. Local authorities operating a sewerage system or owner or operator of any trade or industrial undertaking is required to obtain a licence from the Environmental Management Agency to discharge any effluents or other pollutants into the environment. The effluent is divided into four categories denoting risk as safe, low hazard, medium hazard and high hazard (Government of Zimbabwe, 2007). The scale of the charges for the licence are related to the quality and quantity of the effluent, the poorer the management or quality of the effluent, the higher the charge. In addition, the polluter pays for the policing of the regulations and for the monitoring of the effluent. Penalties for polluting that were moderately punitive under the Water Act, 1998 (imprisonment for a period not exceeding one year or/and a fine) are even more punitive under the Environmental Management Act (CAP 20:27) of 2002 (imprisonment not exceeding five years or/and a fine). The polluter also remedies damage caused either to the environment or to a third party. In practice, although this command and control approach is in force, the Water Quality Section is using a co-operative rather than confrontational approach, with fines being used as a last resort¹⁷¹.

Box 2.2: Water quality governance in Nepal

In Nepal, The Water Resource Act 1992 (2049 BS) contain provisions for the prevention and control of pollution of water resources. Section 19 provides that: No one shall pollute water resources by placing litter, industrial waste, poisons, chemicals or other toxicants to the effect that it exceeds the pollution tolerance limit. The "pollution tolerance limit" for water resources shall be prescribed by His Majesty's Government (HMG), by way of a public notice published in the Nepal Gazette. The prescribed officer (prescribed in the Nepal Gazette) may examine, or cause to examine, a water resource in order to determine whether or not the water resource has been polluted and if pollution tolerance limit has exceeded. Section 22 of the Water Resource Act 1992 (2049 BS) provides that any person or corporation who pollutes water resources will incur a fine of up to NRs.5000 and must pay compensation to any person sustaining a loss as a result of the pollution

¹⁷¹.

Box 2.3: Water quality governance in the USA

In the United States of America, The Clean Water Act (CWA) of 1972 establishes the basic structure for regulating discharges of pollutants into the waters of the United States and regulating quality standards for surface waters. Under the CWA, Environmental Protection Agency (EPA) has implemented pollution control programs such as setting wastewater standards for industry and water quality standards for all contaminants in surface waters. The CWA made it unlawful to discharge any pollutant from a point source into navigable waters, unless a permit was obtained. EPA's National Pollutant Discharge Elimination System (NPDES) permit program controls discharges. Individual homes that are connected to a municipal system, use a septic system, or do not have a surface discharge do not need an NPDES permit; however, industrial, municipal, and other facilities must obtain permits if their discharges go directly to surface waters

172

Box 2.4: Water quality governance in South Africa

In South Africa, the National Water Act (NWA) (Republic of South Africa National Water Act (NWA) No. 36 of 1998) provides for protection of the quality of water resources and for the integrated management of water resources. While also dealing with a number of diverse issues, the NWA oversees pollution prevention in South Africa's water resources by putting a responsibility on the person who owns, controls, occupies or uses the land from where the water is polluted to take measures to prevent pollution. If he or she does not take these measures the Catchment Management Agency (CMA) may do what is necessary to remedy the situation and recover the cost from the person that is responsible¹⁷³.

and agriculture among others, all of which claim a share from increasingly limited water supplies, conservation of both the quality and quantity of water is now a predominant issue on a global scale. Hence, development of better water conservation practices and policies are critical to the sustenance of our water quantity and quality to ensure protection of public health.

CONCLUSION

Pollution of water resources remains a global concern, moreso with the on-going climatic changes that are negatively impacting on the rainfall patterns in every part of the world. Water quality deterioration will most likely see an escalation in the treatment costs for potable water production with a concomitant negative impact on the world's economies and general lifestyles of the world's citizens. Pollution can also have drastic ecological consequences and if not attended to as a matter of urgency, the world's aquatic ecosystems might face certain demise. Pollution of the world's water resources, coupled with the scarcity thereof, is likely to see powerful tribes and/or nations besieging the weaker ones for their freshwater

resources in the near future, as is already the situation of some parts of the world. The only way forward now is pollution reduction by all nations of the world, rich or poor; we all need to take responsibility over the state of our water resources.

REFERENCES

1. Carpenter, S., Chair, Caraco, N.F., Correll, D.L., Howarth, R.W., Sharpley, A.N., Smith, V.H. Issues in ecology: Nonpoint Pollution of Surface Waters with Phosphorus and Nitrogen. *Ecol. Society Am.*, 1998; **3**: 1-14.
2. Kumar, N. A View on Freshwater Environment. *Ecol. Env. Cons.*, 1997; **3**: 3-4.
3. Ewa, E.E., Iwara, A.I., Adeyemi, J.A., Eja, E.I., Ajake, A.O., Otu, C.A. Impact of industrial activities on water quality of Omoku Creek. *Sacha J. Environ. Stud.*, 2011; **1**(2): 8-16.
4. Sangodoyin, A.Y. Groundwater and Surface Water Pollution by Open Refuse Dump in Ibadan, Nigeria. *J. Disc. Innov.*, 1991; **3**(1) 24-31.
5. Adekunle, A.S., Eniola, I.T.K. Impact of Industrial Effluents on Quality of Segment Of Asa River within an Industrial Estate in Ilorin,

- Nigeria. *New York Sci. J.*, 2008; **1**(1): 17-21.
6. Gray, N.F. *Biology of Water Treatment*. New York: Oxford University Press. In: Ewa, E.E., Iwara, A.I., Adeyemi, J.A., Eja, E.I., Ajake, A.O., Otu, C.A. Impact of industrial activities on water quality of Omoku Creek. *Sacha J. Environ. Stud.*, 2011; **1**(2): 8-16.
7. DWAF. Guideline for the Management of Waterborne Epidemics, with the emphasis on Cholera – Co-ordination, Communication, Action and Monitoring. Edition 1. 2001.
8. Mahananda, M.R., Mohanty, B.P., Behera, N.R. Physico-chemical analysis of surface and ground water of Bargarh District, Orissa, India. *IJRRAS.*, 2010; **2**(3): 1-12.
9. The Herald (Friday 13 July 2012). Let's contain water pollution. http://www.herald.co.zw/index.php?option=com_content&view=article&id=46597:lets-contain-water-pollution&catid=44:environment-a-tourism&Itemid=136. Accessed 13 July 2012.
10. Oke, A.D. *Proceedings of 2003 Georgia Water Resources Conference*, held April 23-24, 2003, at The University of Georgia. Kathryn J. Hatcher, editor, Institute of Ecology, The University of Georgia, Athens, Georgia. 2003.
11. Metcalfe, C.D., Miao, X.-S., Koenig, B.G., Struger, J. Distribution of acidic and neutral drugs in surface waters near sewage treatment plants in the lower Great Lakes, Canada. *Environ. Toxicol. Chem.*, 2003; **22**:2881-2889.
12. Levy, S.B. Antibiotic resistance: an ecological imbalance. In: Chadwick, D.J., Goode, J. (Eds.), *Ciba Foundation Symposium 207*. Wiley, West Sussex, 1997; pp. 1-14.
13. Batt, A.L., Bruce, I.B., Aga, D.S. Evaluating the vulnerability of surface waters to antibiotic contamination from varying wastewater treatment plant discharges. *Environ. Poll.*, 2006; **142**: 295-302.
14. Kundzewicz, Z.W., Mata, L.J., Arnell, N.W., Döll, P., Kabat, P., Jiménez, B., Miller, K.A., Oki, T., Sen, Z., Shiklomanov, I.A. Freshwater resources and their management. *Climate Change 2007: Impacts, Adaptation and Vulnerability. Contribution of Working Group II to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*, M.L. Parry, O.F. Canziani, J.P. Palutikof, P.J. van der Linden and C.E. Hanson, Eds., Cambridge University Press, Cambridge, UK, 173-210. 2007.
15. Kolawole, O.M., Ajayi, K.T., Olayemi, A.B., Okoh, A.I. Assessment of Water Quality in Asa River (Nigeria) and Its Indigenous *Clarias gariepinus* Fish. *Int J Environ Res Public Health.*, 2011; **8**(11): 4332-4352.
16. Arain, M.B., Kazi, T.G., Jamali, M.K., Afridi, H.I., Baig, J.A., Jalbani, N., Shah, A.Q. Evaluation of Physico-chemical Parameters of Manchar Lake Water and Their Comparison with Other Global Published Values. *Pak. J. Anal. Environ. Chem.*, 2008; **9**(2): 101-109.
17. Mustapha, M.K. Assessment of the Water Quality of Oyun Reservoir, Offa, Nigeria, Using Selected Physico-Chemical Parameters. *Turk. J. Fisheries Aqua. Sci.*, 2008; **8**: 309-319.
18. Tahir, A., Kanwal, F., Mateen, B. Surveillance of microbial indicators and physicochemical parameters to investigate pollution status of Lahore Canal. *Pak. J. Bot.*, 2011; **43**(6): 2821-2824.
19. Crawshaw, L.I. Responses to rapid temperature change in vertebrate ectotherms. *Am. Zool.*, 1979; **19**: 225-237.
20. Kumar, A., Gupta, H.P., Singh, D.K. Impact of sewage pollution on chemistry and primary productivity of two fresh water bodies in Santal Paragana (Bihar). *Indian J. Ecol.*, 1996; **23**(2): 82-86.
21. Kulkarni, S.V., Tapase, B.S. Physico-chemical parameters and water quality index of Gandhisagar Lake of Umrer in Nagpur District. *Indian Streams Res. J.*, 2012; **2**(5): 2230-7850.
22. WHO. International Standard for drinking water. WHO. In: Dhakal, S. (2006). Study on Physiochemical Parameters and Benthic Macroinvertebrates of Balkhu Khola in Kathmandu Valley, Central Nepal. Paper presented on "Management of Water, Wastewater and Environment: Challenges for the Developing Countries" held in 13-15 Sept 2006, Kathmandu. 1971.
23. Yousafzai, A.M., Khan, A.R., Shakoori, A.R. Pollution of Large, Subtropical Rivers-River Kabul, Khyber-Pakhtun Khwa Province, Pakistan: Physico-Chemical Indicators. *Pakistan J. Zool.*, 2010; **42**(6): 795-808.
24. Zafar, A.R. On the ecology of algae in certain fish ponds on Hyderabad, India: Physico-chemical complexes. *Hydrobiol.*, 1984; **23**: 179-195.
25. DWAF. South African Water Quality Guidelines (second edition). Volume 4: Agricultural Use: Irrigation. 1996.
26. Manson, C.F. *Biology of fresh water pollution. Longman Sci. Tech. Publ.*, 1989. Essex, U.K.
27. Davies, T.C. Chemistry and pollution of natural waters in Western Kenya. *J. Afr. Earth Sci.*, 1996; **23**: 547-563.
28. Dhakal, S. Study on Physiochemical Parameters and Benthic Macroinvertebrates of Balkhu Khola in Kathmandu Valley, Central Nepal. Paper

- presented on "Management of Water, Wastewater and Environment: Challenges for the Developing Countries" held in 13-15 Sept 2006, Kathmandu.
29. Ahmed, W., Goonetilleke, A., Gardner, T. Human and bovine adenoviruses for the detection of source-specific faecal pollution in coastal waters in Australia. *Water Res.*, 2010; doi:10.1016/j.watres.2010.05.017.
 30. Ahmed, W., Neller, R., Katouli, M. Population similarity of enterococci and *Escherichia coli* in surface waters: A predictive tool to trace the sources of faecal contamination. *J. Water Health.*, 2006; doi: 10.2166/wh.2006.042. 347-356
 31. WHO. Monitoring Bathing Waters - A Practical Guide to the Design and Implementation of Assessments and Monitoring Programmes. Ed by Jamie Bartram and Gareth Rees. 2000; ISBN 0-419-24390-1.
 32. Kay, D., Fleischer, J.M., Salmon, R.L., Jones, F., Wyer, M.D., Godfree, A.F., Zelenau-Jacquotte, Z., Shore, R. Predicting likelihood of gastroenteritis from sea bathing: results from randomized exposure. *Lancet.*, 1994; **344**: 905-909.
 33. WHO. *Guidelines for Safe Recreational Water Environments*. Draft for consultation. World Health Organization, Geneva. 1998.
 34. Richardson, K.J., Stewart, M.H., Wolfe, R.L. Application of gene probe technology to the water industry. *J. Am. Water Works Ass.*, 1991; **83**: 71-81.
 35. Dufour, A.P. *Escherichia coli*: the faecal coliform. In: A.W. Hoadley and B.J. Dutka [Eds] *Bacterial Indicators/Health Hazards Associated with Water*. American Society for Testing and Materials, Philadelphia, 1977; 48-58.
 36. Tyagi, V.K., Chopra, A.K., Kazmi, A.A., Kumar, A. (2006). Alternative microbial indicators of faecal pollution: current perspective. *Iran. J. Environ. Health. Sci. Eng.*, 2006; **3**(3): 205-216.
 37. Ashbolt, N.J., Dorsch, M.R., Cox, P.T., Banens, B. Blooming *E. coli*, what do they mean? In: D. Kay and C. Fricker [Eds] *Coliforms and E. coli, Problem or Solution?* The Royal Society of Chemistry, Cambridge, 1997; 78-85.
 38. Lechevallier, M.W. Coliform regrowth in drinking water: a review. *J. Am. Water Works Ass.*, 1990; **82**: 74-86.
 39. Polo, F., Figueras, M.J., Inza, I., Sala, J., Fleisher, J.M., Guarro, J. Prevalence of *Salmonella* serotypes in environmental waters and their relationships with indicator organisms. *Anton Leeuw. Int. J. G.*, 1999; **75**: 285-292.
 40. Leeming, R., Nichols, P.D., Ashbolt, N.J. *Distinguishing Sources of Faecal Pollution in Australian Inland and Coastal Waters using Sterol Biomarkers and Microbial Faecal Indicators*. 1998; Research Report No. 204, Water Services Association of Australia, Melbourne, 46 pp.
 41. Ferguson, C.M., Coote, B.G., Ashbolt, N.J., Stevenson, I.M. Relationships between indicators, pathogens and water quality in an estuarine system. *Water Res.*, 1996; **30**(9): 2045-2054.
 42. Lucena, F., Araujo, R., Jofre, J. Usefulness of bacteriophages infecting *Bacteriodes fragilis* as index microorganisms of remote faecal pollution. *Water Res.*, 1996; **30**(11): 2812-2816.
 43. Balzer, M., Witt, N., Flemming, H.-C., Wingender, J. Faecal indicator bacteria in river biofilms. *Water Sci. Technol.*, 2010; doi: 10.2166/wst.2010.022.
 44. Shakalisava, Y., Doherty, C., Hahnel, W., Diamond, D. A survey of the microbiological water quality of coastal and fresh waters in the Dublin area. 2010; 1-51.
 45. Pipes, W.O. Indicators and water quality. In: W.O. Pipes [Ed.] *Bacterial Indicators of Pollution*. CRC Press Inc., Boca Raton, 1982; 83-95.
 46. USEPA. National Water Quality Inventory: 2000 Report. EPA-841-R-02-001. U.S. Environmental Protection Agency, Washington, DC. 2002.
 47. Wyn-Jones, A.P., Sellwood, J. Enteric viruses in the aquatic environment. *J. Appl. Microbiol.*, 2001; **91**: 945-962.
 48. Phanuwat, C., Takizawa, S., Oguma, K., Katayama, H., Yunika A., Ohgaki S. Monitoring of human enteric viruses and coliform bacteria in waters after urban flood in Jakarta, Indonesia. *Water Sci. Technol.*, 2006; **54**(3): 203-210.
 49. Hot, D., Legeay, O., Jacques, J., Gantzer, C., Caudrelier, Y., Guyard, K., Lange, M., Andreoletti, L. Detection of somatic phages, infectious enteroviruses and enterovirus genomes as indicators of human enteric viral pollution in surface water. *Water Res.*, 2003; **37**: 4703-4710.
 50. Lee, S.H., Kim, S.J. Detection of infectious enteroviruses and adenoviruses in tap water in urban areas in Korea. *Water Res.*, 2003; **36**: 248-256.
 51. Pusch, D., Oh, D.-Y., Wolf, S., Dumke, R., Schröter-Bobsin, U., H'ohne, M., R'oske, I., Schreier, E. Detection of enteric viruses and bacterial indicators in German environmental waters. *Arch. Virol.*, 2005; **150**: 929-947.
 52. Griffin, D.W., Donaldson, K.A., Paul, J.H., Rose, J.B. Pathogenic human viruses in coastal waters. *Clin. Microbiol. Rev.*, 2003; **16**: 129-

- 143.
53. Bosch, A., Lucena, F., Diez, J.M., Gajardo, R., Blasi, M., Jofre, J. Waterborne viruses associated with hepatitis outbreak. *J Am. Water Works Assoc.*, 1991; **83**: 80–83.
54. Morris, R. Reduction of microbial levels in sewage effluents using chlorine and peracetic acid disinfectant. *Water Sci. Technol.*, 1993; **27**: 387–393.
55. Tree, J.A., Adams, M.R., Lees, D.N. Virus inactivation during disinfection of wastewater by chlorination and UV irradiation and the efficacy of F⁺ bacteriophage as a viral indicator. *Water Sci. Technol.*, 1997; **35**: 227–232.
56. Tree, J.A., Adams, M.R., Lees, D.N. Chlorination of indicator bacteria and viruses in primary sewage effluent. *Appl. Environ. Microbiol.*, 2003; **69**: 2038–2043.
57. Wommack, K.E., Colwell, R.R. Virioplankton: viruses in aquatic ecosystems. *Microbiol. Mol. Biol. Rev.*, 2000; **64**: 69–114.
58. Peduzzi, P., Schiemer, F. Bacteria and viruses in the water column of tropical freshwater reservoirs. *Environ. Microbiol.*, 2004; **6**: 707–715.
59. Filippini, M., Buesing, N., Bettarel, Y., Sime- Ngando, T., Gessner, M.O. Infection paradox: high abundance but low impact of freshwater benthic viruses. *Appl. Environ. Microbiol.*, 2006; **72**: 4893–4898.
60. Hennes, K.P., Simon, M. Significance of bacteriophages for controlling bacterioplankton growth in a mesotrophic lake. *Appl. Environ. Microbiol.* 1995; **61**: 333–340.
61. Farnell-Jackson, E.A., Ward, A.K. Seasonal patterns of viruses, bacteria and dissolved organic carbon in a riverine wetland. *Freshwater Biol.*, 2003; **48**: 841–845.
62. Virological Compliance. Draft Guidelines for Drinking-water Quality Management for New Zealand. 2005; **7**: 1–12.
63. Yates, M.V., Gerba, C.P., Kelley, L.M. Virus persistence in groundwater. *Appl. Environ. Microbiol.*, 1985; **49**: 778–781.
64. Gantzer, C., Dubois, E., Crance, J.-M., Billaudel, S., Kopecka, H., Schwartzbrod, L., Pommepuy, M., Le Guyader, F. Influence of environmental factors on the survival of enteric viruses in seawater. *Ocean. Acta.*, 1998; **21**: 983–992.
65. Garza, D.R., Suttle, C.A. The effect of cyanophages on the mortality of *Synechococcus* spp. and selection for UV resistant viral communities. *Microb. Ecol.*, 1998; **36**: 281–292.
66. Bitton, G., Mitchell, R. Effect of colloids on the survival of bacteriophages. *Water Res.*, 1974; **8**: 227–229.
67. Suttle, C.A., Chen, F. Mechanisms and rates of decay of marine viruses in seawater. *Appl. Environ. Microbiol.*, 1992; **58**: 3721–3729.
68. Stallknecht, D. E., Kearney, M. T., Shane, S. M., Zwank, P.J. Effects of pH, temperature and salinity on persistence of Avian Influenza viruses in water. *Avian Dis.*, 1990; **34**: 412–418.
69. Sinton1, L.W., Hall, C.H., Lynch, P.A., Davies-Colley, R.J. Sunlight inactivation of fecal indicator bacteria and bacteriophages from waste stabilization pond effluent in fresh and saline waters. *Appl. Environ. Microbiol.*, 2002; **68**: 1122–1131.
70. Cissoko, M., Desnues, A., Bouvy, M., Sime- Ngando, T., Verling, E., Bettarel, Y. Effects of freshwater and seawater mixing on virio- and bacterioplankton in a tropical estuary. *Freshwater Biol.*, 2008; **53**: 1154–1162.
71. Guixa-Boixareu, N., Calderon-Paz, J. I., Haldal, M. Viral lysis and bacterivory as prokaryotic loss factors along a salinity gradient. *Aquat. Microb. Ecol.*, 1996; **11**: 215–227.
72. Bettarel, Y., Bouvy, M., Dumont, C., Sime- Ngando, T. Virus-bacterium interactions in water and sediment of West African inland aquatic systems. *Appl. Environ. Microbiol.*, 2006; **72**: 5274–5282.
73. Geslin, C., Le Romancer, M., Erauso, G., Gaillard, M., Perrot, G., Prieur, D. PAV1, the first virus-like particle isolated from a hyperthermophilic euryarcheote, “*Pyrococcus abyssi*”. *J. Bacteriol.*, 2003; **185**: 3888–3894.
74. Breitbart, M., Wegley, L., Leeds, S., Schoenfeld, T., Rohwer, T. Phage community dynamics in hot springs. *Appl. Environ. Microbiol.*, 2004; **70**: 1633–1640.
75. Lewis, G. D., Austin, F. J., Loutit, M. W., Sharples, K. Enterovirus removal from sewage - the effectiveness of four different treatment plants. *Water Res.*, 1986; **20**: 1291–1297.
76. Albinana-Gimenez, N., Clemente-Casares, P., Bofill-Mas, S., Hundesa, A., Ribas, F., Girones, R. Distribution of human polyomaviruses, adenoviruses and hepatitis E virus in the environment and in a drinking-water treatment plant. *Environ. Sci. Technol.*, 2006; **40**: 7416–7422.
77. Bofill-Mas, S., Pina, S., Girones, R. Documenting the epidemiologic patterns of polyomaviruses in human populations studying their presence in urban sewage. *Appl. Environ. Microbiol.*, 2000; **66**: 238–245.
78. Bofill-Mas, S., Albinana-Gimenez, N., Clemente-Casares, P., Hundesa, A., Rodriguez-Manzano, J., Allard, A., Calvo, M., Girones, R. Quantification and stability of human

- adenoviruses and polyomavirus JCPyV in wastewater matrices. *Appl. Environ. Microbiol.*, 2006; **72**(12): 7894–7896.
79. He, J., Jiang, S. Quantification of enterococci and human adenoviruses in environmental samples by real-time PCR. *Appl. Environ. Microbiol.*, 2005; **71**(5): 2250–2255.
 80. Katayama, H., Haramoto, E., Oguma, K., Yamashita, H., Tajima, A., Nakajima, H., Ohgaki, S. One-year monthly quantitative survey of noroviruses, enteroviruses, and adenoviruses in wastewater collected from six plants in Japan. *Water Res.*, 2008; **42**: 1441–1448.
 81. Pina, S., Puig, M., Lucena, F., Jofre, J., Girones, R. Viral pollution in the environment and shellfish: human adenovirus detection by PCR as an index of human viruses. *Appl. Environ. Microbiol.*, 1998; **64**: 3376–3382.
 82. Rzezutka, A., Cook, N. Survival of human enteric viruses in the environment and food. *FEMS Microbiol. Rev.*, 2004; **28**: 441–453.
 83. Lopman, B.A., Reacher, M.A., van Duynhoven, Y., Hanon, F.-X., Brown, D., Koopmans, M. Viral gastroenteritis outbreaks in Europe. *Emerg. Inf. Dis.*, 2003; **9**: 90–96.
 84. Skrabber, S., Gassiolloud, B., Schwartzbrod, L., Gantzer, C. Survival of infectious Poliovirus-1 in river water compared to the persistence of somatic coliphages, thermotolerant coliforms and Poliovirus-1 genome. *Water Res.*, 2004; **38**: 2927–2933.
 85. Espinosa, A.C., Espinosa, R., Maruri-Avidal, L., Méndez, E., Mazari-Hiriart, M., Arias, C.F. Infectivity and genome persistence of rotavirus and astrovirus in drinking and irrigation water. *Water Res.*, 2008; **42**: 2618–2628.
 86. Mead, P.S., Slutsker, L., Dietz, V. Food-related illness and death in the United States. *Emerg. Inf. Dis.*, 1999; **5**: 607–625.
 87. Sobsey, M. D. Inactivation of health-related microorganisms in water by disinfection processes. *Water Sci. Technol.*, 1989; **21**(3): 179–195.
 88. De Cardona, I.L., Bermudez, M., Billmire, E., Hazen, T.C. Enteric Viruses in a Mangrove Lagoon, Survival and Shellfish Incidence. *Caribbean J. Sci.*, 1988; **24** (3-4): 102–111.
 89. Hurst, C.J., Benton, W.H., McClellan, K.A. Thermal and water sources effects upon the stability of enterovirus in surface freshwaters. *Can J Microbiol.*, 1989; **35**: 474–480.
 90. Geldenhuys, J.C., Pretorius, P.D. The occurrence of enteric viruses in polluted water, correlation to indicator organisms and factors influencing their numbers. *Water Sci. Technol.*, 1989; **21**: 105–109.
 91. Keswick, B.H., Gerba, C.P., DuPont, H.L., Rose, J.B. Detection of enteric viruses in treated drinking water. *Appl. Environ. Microbiol.*, 1984; **47**: 1290–1294.
 92. Raphael, R.A., Sattar, S.A., Springthope, V.S. Long-term survival human rotavirus in raw and treated river water. *Can. J. Microbiol.*, 1985; **31**: 124–128.
 93. Noble, R.T., Fuhrman, J.A. Virus decay and its causes in coastal waters. *Aquat. Microb. Ecol.*, 1997; **63**: 77–83.
 94. Wilhelm, S.W., Weinbauer, M.G., Suttle, C.A., Jeffrey, W.H. The role of sunlight in the removal and repair of viruses in the sea. *Limnol. Oceanogr.*, 1998; **43**: 586–592.
 95. Weinbauer, M.G., Wilhelm, S.W., Suttle, C.A., Pledger, R.J., Mitchell, D.L. Sunlight-induced DNA damage and resistance in natural viral communities. *Aquat. Microb. Ecol.*, 1999; **17**: 111–120.
 96. Jacquet, S., Bratbak, G. Effects of ultraviolet radiation on marine virus-phytoplankton interactions. *FEMS Microb. Ecol.*, 2003; **44**: 279–289.
 97. Weinbauer, M.G. Ecology of prokaryotes viruses. *FEMS Microbiol. Rev.*, 2004; **28**: 127–181.
 98. Giladi, H., Goldenberg, D., Koby, S., Oppenheim, A.B. Enhanced activity of the bacteriophage-lambda p-l promoter at low-temperature. *PNAS.*, 1995; **92**: 2184–2188.
 99. Greening, G.E., Hewitt, J., Lewis, G.D. Evaluation of integrated cell culture-PCR (C-PCR) for virological analysis of environmental samples. *J. Appl. Microbiol.*, 2002; **93**: 745–750.
 100. Murrin, K., Slade, J. Rapid detection of viable enteroviruses in water by tissue culture and semi-nested polymerase chain reaction. *Water Sci. Technol.*, 1997; **35**: 429–432.
 101. Reynolds, K.A., Gerba, C.P., Pepper, I.L. Rapid PCR-based monitoring of infectious enteroviruses in drinking water. *Water Sci. Technol.*, 1997; **35**: 423–427.
 102. Tani, N., Dohi, Y., Jurumatani, N., Yonemasu, K. Seasonal distribution of adenoviruses, enteroviruses and reoviruses in urban river water. *Micro. Immunol.*, 1995; **39**: 577–580.
 103. Craun, G. F. Causes of waterborne outbreaks in the United States. *Wat. Sci. Technol.*, 1991; **24**: 17–20.
 104. Albinana-Gimenez, N., Miagostovich, M.P., Calgaa, B., Huguet, J.M., Matia, L., Girones, R. Analysis of adenoviruses and polyomaviruses quantified by qPCR as indicators of water quality in source and drinking-water treatment

- plants. *Water Res.*, 2009; **43**: 2011–2019.
105. Abad, F.X., Pintó, R.M., Villena, C., Bosch, A. Astrovirus survival in drinking water. *Appl. Environ. Microbiol.*, 1997; **63**: 3119–3122.
106. Fong, T.-T., Phanikumar, M.S., Xagorarakis, I., Rose, J.B. Quantitative detection of human adenoviruses in waste water and combined sewer overflows influencing a Michigan River. *Appl. Environ. Microbiol.*, 2009; 1–40.
107. Basu, G., Rossouw, J., Sebunya, T.K., Gashe, B.A., De Beer, M., Dewar, J.B., Steele, A.D. Prevalence of rotavirus, adenovirus and astrovirus infection in young children with gastroenteritis in Gaborone, Botswana. *East Afr. Med. J.*, 2003; **80**: 652–655.
108. Cruz, J. R., Cáceres, P., Cano, F., Flores, J., Bartlett, A., Torún, B. Adenovirus types 40 and 41 and rotaviruses associated with diarrhea in children from Guatemala. *J. Clin. Microbiol.*, 1990; **28**: 1780–1784.
109. Logan, C., O’Leary, J.J., O’Sullivan, N. Real-time reverse transcription-PCR for detection of rotavirus and adenovirus as causative agents of acute viral gastroenteritis in children. *J. Clin. Microbiol.*, 2006; **44**: 3189–3195.
110. Meqdam, M.M., Thwiny, I.R. Prevalence of group A rotavirus, enteric adenovirus, norovirus and astrovirus infections among children with acute gastroenteritis in Al-Qassim, Saudi Arabia. *Pak. J. Med. Sci.*, 2007; **23**: 551–555.
111. van Heerden, J., Ehlers, M.M., Grabow, W.O.K. Detection and risk assessment of adenoviruses in swimming pool water; *J. Appl. Microbiol.*, 2005; **99**: 1256–1264.
112. EPA. Drinking water contamination candidate list. Notice. *Fed. Regul.*, 1998; **63**: 10274–10287.
113. Maier, R.M., Pepper, I.L., Gerba, C.P. Viruses: In Environmental Microbiology. 2000; 473–475. London: Academic Press.
114. Jiang, S.C. Human Adenoviruses in Water: Occurrence and Health Implications: A Critical Review. *Environ. Sci. Technol.*, 2006; **40**: 7132–7140.
115. Kojaghlanian, T., Flomenberg, P., Horwitz, M.S. The impact of adenovirus infection on the immunocompromised host. *Rev. Med. Virol.*, 2003; **13**: 155–171.
116. Pinto, R.M., Costafreda, M.I., Bosch, A. Risk Assessment in Shellfish-Borne Outbreaks of Hepatitis A. *Appl. Environ. Microbiol.*, 2009; **75**(23): 7350–7355.
117. Conaty, S., Bird, P., Bell, G., Kraa, E., Grohmann, G., McNulty, J.M. Hepatitis A in New South Wales, Australia from consumption of oysters: the first reported outbreak. *Epidemiol. Infect.*, 2000; **124**: 121–130.
118. Dismukes, W.E., Bisno, A.L., Katz, S., Johnson, R.F. An outbreak of gastroenteritis and infectious hepatitis due to raw clams. *Am. J. Epidemiol.*, 1969; **89**: 555–561.
119. Leoni, E., Bevini, C., Esposti, S.D., Graziano, A. An outbreak of intrafamilial hepatitis A associated with clam consumption: epidemic transmission to a school community. *Eur. J. Epidemiol.*, 1998; **14**: 187–192.
120. Mackowiak, P.A., Caraway, C.T., Portnoy, B.L. Oyster-associated hepatitis: lessons from the Louisiana experience. *Am. J. Epidemiol.*, 1976; **103**: 181–191.
121. Mele, A., Rastelli, M.G., Gill, O.N., di Bisceglie, D., Rosmini, F., Pardelli, G., Valtriani, C., Patriarchi, P. Recurrent epidemic hepatitis A associated with consumption of raw shellfish, probably controlled through public health measures. *Am. J. Epidemiol.*, 1989; **130**: 540–546.
122. Sanchez, G., Pinto, R.M., Vanaclocha, H., Bosch, A. Molecular characterization of hepatitis A virus isolates from a transcontinental shellfish-borne outbreak. *J. Clin. Microbiol.*, 2002; **40**: 4148–4155.
123. Stroffolini, T., Biagini, W., Lorenzoni, L., Palazzesi, G.P., Divizia, M., Frongillo, R. An outbreak of hepatitis A in young adults in central Italy. *Eur. J. Epidemiol.*, 1990; **6**: 156–159.
124. Halliday, M.L., Kang, L.-Y., Zhou, T.-Z., Hu, M.-D., Pan, Q.-C., Fu, T.-Y., Huang, Y.S., Hu, S.-L. An epidemic of hepatitis A attributable to the ingestion of raw clams in Shanghai, China. *J. Infect. Dis.*, 1991; **164**: 852–859.
125. Beller, M., Ellis, A., Lee, S.H., Drebot, M.A., Jenkerson, S.A., Funk, E., Sobsey, M.D., Simmons III, O.D., Monroe, S.S., Ando, T., Noel, J., Petric, M., Middaugh, J.P., Spika, J.S. Outbreak of viral gastroenteritis due to a contaminated well. International consequences. *J. Am. Med. Assoc.*, 1997; **278**: 563–568.
126. Gray, J.J., Green, J., Cunliffe, C., Gallimore, C., Lee, J.V., Neal, K., Brown, D.W. Mixed genogroup SRSV infections among a party of canoeists exposed to contaminated recreational water. *J. Med. Virol.*, 1997; **52**: 425–429.
127. De Serres, G., Cromeans, T.L., Levesque, B., Brassard, N., Barthe, C., Dionne, M., Prud’homme, H., Paradis, D., Shapiro, C.N., Nainan, O.V., Margolis, H.S. Molecular confirmation of hepatitis A virus from well water: epidemiology and public health implications. *J. Infect. Dis.*, 1999; **179**: 37–43.
128. Kukkula, M., Maunula, L., Silvennoinen, E., von Bonsdorff, C.H. Outbreak of viral gastroenteritis due to drinking water contaminated by Norwalk-

- like viruses. *J. Infect. Dis.*, 1999; **180**: 1771–1776.
129. Haefliger, D., Hubner, P., Luthy, J. Outbreak of viral gastroenteritis due to sewage-contaminated drinking water. *Int. J. Food Microbiol.*, 2000; **54**: 123–126.
 130. Beuret, C., Kohler, D., Baumgartner, A., Luthi, T.M. Norwalklike virus sequences in mineral waters: one-year monitoring of three brands. *Appl. Environ. Microbiol.*, 2002; **68**: 1925–1931.
 131. Parshionikar, S.U., Willian-True, S., Fout, G.S., Robbins, D.E., Seys, S.A., Cassady, J.D., Harris, R. Waterborne outbreak of gastroenteritis associated with a norovirus. *Appl. Environ. Microbiol.*, 2003; **69**: 5263–5268.
 132. WHO. Water Sanitation Health. Making water a part of economic development: The economic benefits of improved water management and services. http://www.who.int/water_sanitation_health/waterandmacroeconomics/en/index.html. 2012; Accessed 19/09/2012.
 133. CSIR. The impact of an unhealthy environment on human health in South Africa. Briefing Note 2009/04. http://www.csir.co.za/nre/docs/Briefing%20Note%20No4%202010_environmental%20health_FINAL.pdf. Accessed 17/08/2012
 134. Baboviæ, N., Markoviæ, D., Dimitrijeviæ, V., Markoviæ, D. Some indicators of water quality of the Tamiš River. *Chem. Ind. Chem. Eng. Quart.*, 2011; **17**(1): 107–115.
 135. Simonne, A. Principles and Practices of Food Safety for Vegetable Production in Florida. 2010; <http://edis.ifas.ufl.edu/cv288>. Accessed 12/09/2012.
 136. Suslow, T.V., Oria, M.P., Beuchat, L.R., Garrett, E.H., Parish, M.E., Harris, L.J., Farber, J.N., Busta, F.F. Production Practices as Risk Factors in Microbial Food Safety of Fresh and Fresh-Cut Produce. *Compr. Rev. Food Sci. Food Safety*. 2003; **2**(1): 38–77.
 137. Bauder, T.A., Waskom, R.M., Sutherland, P.L., Davis, J.G. Irrigation Water Quality Criteria. Fact Sheet No. 0.506. Crop series/Irrigation. 2008; 1–4.
 138. Landschoot, P. Irrigation Water Quality Guidelines for Turfgrass Sites. 2012; <http://plantscience.psu.edu/research/centers/turf/extension/factsheets/water-quality>. Accessed 11/09/2012.
 139. Ait, A., Hassan, L. Salmonella infection in children from the wastewater spreading zone of Marrakesh City (Morocco). *J. Appl. Microbiol.* 1999; **87**: 536–539.
 140. USDA. The Census of Agriculture: Farm and Ranch Irrigation Survey. 2003; <http://www.agcensus.usda.gov/Publications/2002/FRIS/>. Accessed 21/09/2012.
 141. ANZECC & ARMCANZ. Australian guidelines for water quality monitoring and reporting. National Water Quality Management Strategy Paper No 7, Australian and New Zealand Environment and Conservation Council & Agriculture and Resource Management Council of Australia and New Zealand, Canberra. 2000.
 142. WHO. *Health Guidelines for the Use of Wastewater in Agriculture and Aquaculture. Report of a WHO Scientific Group*. Technical Report Series 1989; 778, WHO, Geneva.
 143. Bartone, C.R., Esparza, M.L., Mayo, C., Rojas, O., Vitko, T. *Monitoring and maintenance of treated water quality in the San Juan lagoons supporting aquaculture*, 1985; Final Report of Phases I and II, UNDP/World Bank/GTZ Integrated Resource Recovery Project GLO/80/004, CEPIS.
 144. Barua, A., Hubacek, K. Water Pollution And Economic Growth: An Environmental Kuznets Curve Analysis At The Watershed And State Level. 2012; http://umcp.academia.edu/klaushubacek/Papers/563447/Water_pollution_and_economic_growth_An_Environmental_Kuznets_Curve_analysis_at_the_watershed_and_state_level Accessed 03/09/2012.
 145. United Nations Committee on Economic Social and Cultural Rights. *General Comment No. 15 (2002). The Right to Water*. E/C.12/2002/11, United Nations Social and Economic Council, 2003; pp18.
 146. Fawell, J., Nieuwenhuijsen, M.J. Contaminants in drinking water. *British Medical Bulletin* 2003; **68**: 199–208. DOI: 10.1093/bmb/ldg027.
 147. Chorus, I., Bartram, J. Toxic cyanobacteria in water. *A Guide to their Public Health Consequences, Monitoring and Management*. 1999; Published on behalf of WHO by E & FN Spon, London and New York.
 148. WHO. Preventing disease through healthy environments: towards an estimate of the environmental burden of disease. In: CSIR (2010). The impact of an unhealthy environment on human health in South Africa. Briefing Note 2009/04. http://www.csir.co.za/nre/docs/Briefing%20Note%20No4%202010_environmental%20health_FINAL.pdf. Accessed 17/08/2012.
 149. WHO, (2009). Addendum to Guidelines for Safe Recreational Water Environments, Vol 1 World Health Organization – Geneva, Switzerland. WHO/HSE/WSH/10.04. 2009; <http://whqlibdoc.who.int/hq/2010/>

- WHO_HSE_WSH_10.04_eng. Accessed 19/09/2012.
150. USEPA, Bacterial water quality standards for recreational waters (freshwater and marine waters): Status Report. EPA-823-R-03-008. http://water.epa.gov/type/oceb/beaches/upload/2003_06_19_beaches_local_statrept.pdf. Accessed 11/09/2012, 2003.
 151. Nkwonta, O. I., Ochieng, G.M. Water Pollution in Soshanguve Environs of South Africa. World Academy of Science, Engineering and Technology. 2009; **56**: 499-503.
 152. Oberholster, P.J., Ashton, P.J. State of the Nation Report: An Overview of the Current Status of Water Quality and Eutrophication in South African Rivers and Reservoirs. 2008; <http://npconline.co.za/MediaLib/Downloads/Home/Tabs/Diagnostic/ons2/An%20ov>. Accessed 19/09/2012.
 153. Brulliard, N. Climate change and economic growth drain South Africa's low water supplies. *Global Post*. 2009.
 154. Binns, J.A., Illgner, P.M., Nel, E.L. Water shortage, deforestation and development: South Africa's working for water programme. *Land Degrad. Develop.*, 2001; **12**: 341-355.
 155. Walmsley, R.D., Walmsley, J.J., Silberbauer, M. National State of the Environment Report - South Africa: freshwater systems and resources. 1999; <http://www.ngo.grida.no/soesa/nsoer/issues/water/index.htm>. Accessed 05/09/2012.
 156. National Committee on Climate Changes. Discussion on Climate Changes. 1998; <http://www.environment.gov.za/nsoer/resource/climate/climate.htm>. Accessed 19/09/2012.
 157. Blignaut, J., van Heerden, J. The impact of water scarcity on economic development initiatives. *Water SA.*, 2009; **35**(4): 415-420.
 158. Rand Water. Water situation in South Africa. 2012; <http://www.waterwise.co.za/site/water/environment/situation.html>. Accessed 18/09/2012.
 159. Marshal, L. Water Crisis Looms in South Africa. http://newswatch.nationalgeographic.com/2010/11/05/water_crisis_looms_in_south_africa/. Accessed 18/09/2012.
 160. DWAF. Chapter 2: South Africa's water situation, and strategies to balance supply and demand. 2002; <http://www.dwaf.gov.za/docs/NWRS/4%20Chap%202%20South%20Africa's%20Water%20Situation.pdf>. Accessed 18/09/2012.
 161. DWAF. *Overview of the Water Resources availability and utilisation in South Africa*. 1997; ISBN 0 7970 3540 0.
 162. Government of South Africa – Department of Water Affairs and Forestry (GOSA–DWAF). Limpopo Water Management Area. *Overview of water resources availability and utilization* 2003. (available at www.dwaf.gov.za).
 163. CSIR Environmentek. Protection and strategic uses of groundwater resources in drought prone areas of the SADC region groundwater situation analysis of the Limpopo river basin, Report No. ENV-P-C 2003-026. Sanciahs <http://www.dbsa.org/Blog/Lets%20talk%20water%20with%20DBSA1/The%20water%20Crisis%20in%20South%20Africa.pdf>. Accessed 13 July 2012.
 164. Vega, M., Pardo, M.R., Barrado, E., Debaâ, N.L. Assessment of seasonal and polluting effects on the quality of river water by exploratory data analysis. *Wat. Res.*, 1998; **32**(12): 3581-3592.
 165. Nationmaster.com. South Africa: Environment. 2003; <http://www.nationmaster.com/country/sf/Environment>. Accessed 19/09/2012.
 166. Schreiner, B. The challenges of water resources management in South Africa. Keynote address: SANCIAHS Conference. 1999; 1-19.
 167. Löwgren, M., Hillmo, T., Lohm, U. Water pollution perspectives: Problem conceptualizations and abatement strategies in Sweden during the 20th century. *Geo J.*, 1989; **19**(2): 161-171, doi: 10.1007/BF00174645
 168. Figueras, M.J., Borrego, J.J. New Perspectives in Monitoring Drinking Water Microbial Quality. *Int. J. Environ. Res. Public Health.*, 2010; **7**: 4179-4202; doi:10.3390/ijerph7124179
 169. Bartram, J., Corrales, L., Davison, A., Deere, D., Drury, D., Gordon, B., Howard, G., Rinehold, A., Stevens, M. *Water Safety Plan Manual: Step-by-Step Risk Management for Drinking Water Suppliers*; World Health Organization: Geneva, Switzerland. 2009.
 170. WHO/UNEP. Water Pollution Control - A Guide to the Use of Water Quality Management Principles. 1997; http://www.who.int/water_sanitation_health/resourcesquality/watpolcontrol.pdf. Accessed 17/09/2012.
 171. WaterAid Nepal. Water Laws in Nepal. Laws Relating to Drinking Water, Sanitation, Irrigation, Hydropower and Water Pollution. 2005; pp 1-94
 172. USEPA. Water: Office of Wetlands, Oceans & Watersheds, 2012; <http://water.epa.gov/aboutow/owow/laws.cfm>. Accessed 13 July 2012.
 173. Republic of South Africa, Government Gazette. Act N 36 of 1998, National Water Act 1998. **398**(19182): 1-101. <http://www.info.gov.za/view/DownloadFileAction?id=70693>. Accessed 17/09/2012.