

Evaluation of the IGRs Alsystin and Pyriproxyfen as Well as the Plant Extract Jojoba Oil against the Mosquito *Aedes aegypti*

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The biological activity of two insect growth regulators alsystin and pyriproxyfen as well as the plant extract jojoba oil on larval stages and pupae until adult emergence of *Aedes aegypti* was evaluated. According to IC_{50} values obtained (concentration which to inhibit the emergence of 50% of mosquito adults), alsystin (0.00048ppm) proved to be highly effective against *A. aegypti* than pyriproxyfen (0.005 ppm) and jojoba oil (85 ppm). On the other hand, larval treatments with the present compounds led to a decrease in egg production and hatchability of eggs produced by mosquito adults which survived from larval treatments.

Key words: *Aedes aegypti*, insect growth regulators, plant extract, Mosquito larvae, reproductive potential

The success of insecticide-based control programmes in reducing the prevalence of insect vector-borne diseases has been accompanied by growing interest regarding the harmful effects of wide scale and prolonged used of insecticides on human health and the environment (WHO, 1995). Furthermore, many mosquito vectors have developed resistance to a number of conventional chemical insecticides (Xu *et al.*, 2006; Pridgeon *et al.*, 2008), creating an urgent need to seek and identify new effective insecticides to control these important disease vectors. Therefore, more attention has been recently paid to the use of non-conventional insecticides such as insect growth regulators (IGRs) and plant extracts for mosquito control in different parts of the world (Mohsen and Mehdi, 1989; Koumu *et al.*, 2007; Siddigui *et al.*, 2009).

The present study was planned in part to evaluate the biological activity of two IGRs alsystin and pyriproxyfen as well as the plant extract mosquito oil against mosquito larvae of *Aedes aegypti*, the primary vector of dengue fever in Jeddah governorate, Saudi Arabia. Additional trials were also conducted to study the possible delayed effects of larval treatments with the tested compounds on the reproductive potential of mosquito adult survivors.

MATERIALS AND METHODS

Mosquito strain

A field strain of *A. aegypti* was used in the present study. The parental strain was raised from wild larvae collected from Jeddah governorate, Saudi Arabia. This stock colony was maintained under laboratory conditions of $27 \pm 1^\circ\text{C}$ and $70 \pm 5\%$ R.H. with 14:10 (L:D) photoperiod throughout this study. The larvae were reared until pupation and adult emergence took place for maintaining the stock culture.

The tested compounds

The following compounds were used:

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1. The IGR pyriproxyfen (Sumilarv 0.5% G), 2-[1-methyl-2-(4-phenoxyphenoxy) ethoxy] pyridine, kindly provided by Sumitomo Chem. Co., Japan.
2. The IGR alsystin (tiflumuron 84%EC), 2-Chloro-N-(4-trifluoromethoxy phenyl) amino carbonyl benzamide, kindly supplied by Bayer Ltd., Germany.
3. The plant extract jojoba oil (*Simmondsia chinensis*, Fam. Simmondsiaceae), kindly provided prof. Dr. Moustafa S. Saleh, Dept., of Applied Entomology, Fac. Of Agric., Alexandria Univ. Egypt. The stock solution of the plant extract was prepared by adding 1 ml of it to 99 ml of distilled water containing 0.5% triton X-100 as an emulsifier to ensure complete solubility of the extract in water. Series of concentrations were prepared in distilled water.

Test Experiments

The larval susceptibility test was conducted according to the method of WHO (2005). Treatments were carried out by exposing early 4th instar larvae to various concentrations of the test compounds in groups of glass beakers containing 100 ml tap water. Five replicates of 20 larvae each per concentration, and so for control trails were set up. The larvae were given the usual larval food during these tests. Data on larval mortality, pupation and adult emergence were recorded daily, and compared with control trails. The biological effects of the tested compounds was expressed as the percentage of larvae that do not develop into successfully emerging adults or the inhibition of adult emergence (% IE) according to the formula (WHO, 2005):

$$\%IE = \frac{C-T}{C} \times 100$$

Where C represents the percentage emergence in the control and T represents the percentage emergence in the treatments. The inhibition of adult emergence - concentration-probability line (IC-p line) was drawn for each compound using the method of Litchfield and Wilcoxon (1949).

Values of IC₅₀ (concentrations which to give approximately 50% inhibition of adult emergence) were obtained from the IC-p lines of alsystin, pyriproxyfen and jojoba oil. The

concentrations corresponding to these values were prepared and used for treating the early fourth instar larvae of *A. aegypti*. Fifteen replicates of 20 larvae each were conducted for each concentration. Mosquito adults which survived from the above larval treatments were isolated in clean cages. Seventy two hours later, emerged mosquito females were fed on a pigeon for a blood meal. Each engorged female was kept with a male in a small glass-cup, half-filled with tap water and covered with muslin cloth. They were fed on a 10% sugar solution soaked on cotton pads and put on top of the covered cups. Number of eggs laid per female and hatchability of eggs were recorded for the 1st gonotrophic cycle.

RESULTS AND DISCUSSION

Table 1 shows susceptibility levels of *A. aegypti* larvae following treatments with different concentrations of the IGRs alsystin and pyriproxyfen as well as the plant extract jojoba oil. The effective concentrations of the above compounds against 4th larval instar ranged from 0.0002–0.004 ppm, 0.002–0.02 ppm and 30–250 ppm, respectively. The corresponding larval mortalities for these compounds were 3–25%, 8–17% and 12–51%. The records showed that the test compounds did not appear to give high percentage of mortality against larval stages. Moreover, most pupae were often died before the adult emergence or as albino pupae (Bridges *et al.*, 1977). Also, some mosquito adults were emerged incompletely or left their antennae, mouth parts and legs attached in the pupal exuvia Fig(1). Therefore, in the present work cumulative mortalities during larval development to pupae and adults have been taken as a criterion for evaluating the biological effects of the tested compounds as they have more juvenilizing effects than toxic mode of action.

Generally, larval treatments with the effective concentrations of the IGRs alsystin and pyriproxyfen as well as the plant extract jojoba oil caused 20.4–89.2%, 16.1–91.4% and 18.3–90.3% inhibition of adult emergence, respectively. Taking IC₅₀ values (concentration which to inhibit the emergence of 50% of adults) into consideration, the records indicated that alsystin (0.00048 ppm) proved to be highly effective against *A. aegypti* than pyriproxyfen (0.005 ppm) and the jojoba oil

extract (85 ppm). However, variations in the susceptibility status of the present *A. aegypti* larvae may be attributed to the differential mode of action of the compounds tested and its effective concentrations. The fluctuations in the percentage inhibition of adult emergence obtained for the different concentrations of the test compounds against the present mosquito strain support this conclusion. Laboratory and field trials in this respect were carried out by several authors to determine the susceptibility status of different mosquito species to IGRs and plant extracts (Saleh and Wright, 1990; Sharma *et al.*, 2005; Bai *et al.*, 2007; Pridgeon *et al.*, 2008). However, it can be

concluded that determination of susceptibility levels of mosquito larvae in any area will provide baseline data for planning control programmes and making decisions about insecticides usage in these areas (Paeporn *et al.*, 2005).

The possible delayed effects of larval treatments with IC₅₀ values of alsystin (0.00048 ppm), pyriproxyfen (0.005 ppm) and jojoba oil (85 ppm) on the reproductive potential of mosquito adult survivors are shown in Table 2. The results indicated that larval treatments with the test compounds led to a marked decrease in egg production and hatching levels of eggs laid by mosquito female survivors. The mean number of

Table 1. The biological effects of the IGRs alsystin and pyriproxyfen as well as the plant extract jojoba oil on the developmental stages of *A. aegypti*

Compounds	Effective concentrations (ppm)	Larval mortality ^a (%)	Pupae produced (%)	Adult emergency (%)	(%IE) ^b	IC ₅₀ ^c (ppm)
alsystin	0.0002	3	97	74	20.4	0.00048
	0.0004	15	85	57	38.7	
	0.0007	18	82	32	65.6	
	0.001	11	89	20	78.5	
	0.004	25	75	10	89.2	
Pyriproxyfen	0.002	8	92	78	16.1	0.005
	0.004	11	89	58	37.6	
	0.007	19	81	29	68.8	
	0.001	20	80	18	80.6	
	0.02	17	83	8	91.4	
jojoba oil	30	12	88	76	18.3	85
	70	16	84	53	43	
	120	25	75	36	61.3	
	200	36	64	18	80.6	
	250	51	49	9	90.3	
Control		4	96	93		

^a Five replicates, 20 larvae each.

^b WHO(2005)

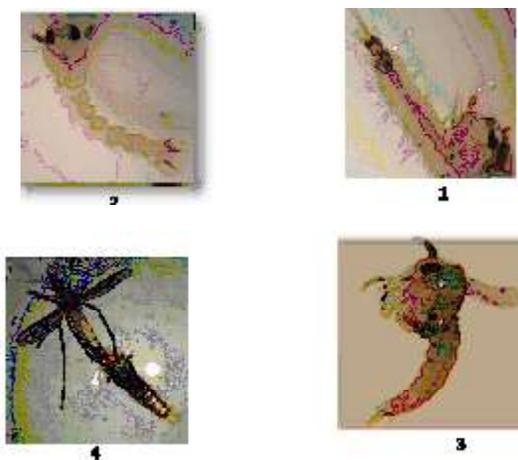
^c Litchfield and Wilcoxon (1949)

Table 2. The delayed effects of larval treatment with IC₅₀ values of alsystin, pyriproxyfen and jojoba oil on the reproductive potential of mosquito adult survivors

Compounds	IC ₅₀ (ppm)	Total number of eggs*	Mean number of eggs/female	Total number of larvae hatched	Hatchability (%)
alsystin	0.00048	1271	50.8	796	62.6
Pyriproxyfen	0.005	1448	57.9	851	58.8
Jojoba oil	66	1206	48.2	665	55.1
Control		1693	67.7	1546	91.3

* Mean of 25 engorged mosquito females.

eggs per female during the 1st gonotrophic cycle was in respect 50.8, 57.9 and 48.2 eggs when compared with the control which was 67.7 eggs (Table 2). This means that larval treatments caused a reduction in egg-laying capacity of female survivors by about 24.9, 14.5 and 28.8%, respectively. On the other hand, the percentage of hatchability of eggs produced by mosquito females that emerged from larval treatments with IC₅₀ levels of alsystin, pyriproxyfen and jojoba oil was in respect 62.6, 58.8 and 55.1% when compared with the control (91.3%). The results thus indicate that egg hatchability was decreased by about 31.4, 35.6 and 39.6% after larval treatment with the above compounds. A possible explanation is that larval treatments with sub-lethal doses of the test IGRs and the plant extract may be affect the larval gonads and accordingly the reproductive capacity of mosquito adult survivors (Saleh *et al.*, 2003). Generally, studies in this respect were carried out by several investigators using different IGRs and plant extracts against many species of mosquito vectors (Vasuki, 1999; Chowdhury *et al.*, 2007; Howard *et al.*, 2009; Saleh *et al.*, 2013). However,



1. Intermediate stage between larvae and pupa
 a- Terminal segments of larvae siphon.
 b- cephalothorax of pupa respiratory trumpets.
2. Pupa lacked the hardening and darkening of the cuticle (albino pupa).
3. Intermediate stage between pupae and adult.
4. Adult completely emerged but they are unable to fly, legs attached in the pupal exuvia.

Fig. 1. Abnormalities in the developmental stages of against *A. aegypti* after larval treatments with the compounds test

much additional trails are needed to evaluate the possible delayed effects of such IGRs and plant extracts on some biological and behavioural aspects of mosquito adults which survived from larval treatments.

REFERENCES

1. Bai, L.; Wang, L.; Zhao, M.; Toki, A.; Hasegawa, T.; Ogura, H.; Kataok, T.; Higosem, K.; Sakai, J., Bai, J. and Ando, M., Bioactive pregnanes from *Nerium oleander*. *J. Nat. Prod.* 2007; **70**: 14–8.
2. Bridges, A. C.; Coke, J.; Olsen, J.K. and Mayer, R.T. Effects of new fluorescent insect growth regulators on larval instars of *Aedes aegypti*. *Mosq. News*, 1977; **37**: 227–233.
3. Chowdhury, N.; Bhattachajee, I.; Laskar, S. and Chandra, G., Efficacy of *Solanum villosum* Mill (Solanceae: Solanales) as a biocontrol agent against fourth instar larvae of *Culex quinquefasciatus* Say. *Turk. J. Zool.* 2007; **31**: 365-370.
4. Ghosh, A.; Chowdhury, N. and Chandra, G., Plant extracts as potential mosquito larvicides. *Indian J. Med. Res.* 2012; **135**: 581-598.
5. Mohsen, Z. H. and Mehdi, N. S., Effects of insect growth inhibitor Alsystin on *Culex quinquefasciatus*. *Int. J. Trop. Ins.* 1989; **10**(1):29-33.
6. Howard, A.F.; Adongo, E.A.; Hassanali, A.; Omlin, F.X.; Wanjoya, A.; Zhou, G. and Vulule, J., Laboratory evaluation of the aqueous extract of *Azadirachta indica* (neem) wood chippings on *Anopheles gambiae* (Diptera: culicidae) mosquitoes. *J. Med. Entomol.* 2009; **46**(1): 107-104.
7. Litchfield, J. T. and Wilcoxon, E., A simplified method of evaluating dose- effect exper. *J. Phar. Exp. Ther.* 1949; **96**: 99-113.
8. Okumu, F.O.; Knols, B.G.J. and Fillinger, U., Larvicidal effect of a neem oil (*Azadirachta indica*) formulation on the malaria vector *Anopheles gambiae*. *Malar. J.* 2007; **6**: 63.
9. Paeporn, P.; Kasin, S.; Sathantriphop, S. and Sangkitporn, S., Insecticides Susceptibility of *Aedes aegypti* in Tsunami-affected Areas in Thailand, *Dengue Bull.* 2005; **29**: 210-213.
10. Pridgeon, J.W.; Pereira, R.M.; Becnel, J.J.; Allan, S.A.; Clark, G.G. and Linthicum, K.J., Susceptibility of *Aedes aegypti*; *Culex quinquefasciatus* and *Anopheles quadrimaculatus* to 19 pesticides with different mode of action. *J. Med. Ent.* 2008; **45**(1): 82–87.

11. Saleh, M.S. and Wright, R.E., Evaluation of the IGR cyromazine as a feed-through treatment against *Culex pipiens* and *Aedes epacticus*. *J. App. Ent.* 1990; **109**: 247-250.
12. Saleh, M.S.; El-Meniawi, F.A.; Kelada, N.L. and Zahran, H.M., Resistance development in mosquito larvae *Culex pipiens* to the bacterial agent *Bacillus thuringiensis* var. *israelensis*. *J. App. Ent.* 2003; **127**: 29-32.
13. M. S. Saleh, Osama Abdullah Abuzinadah, Khalid Saeed AlGhamdi, Ahmad Ibrahim Assagaf and Jazem A. Mahyoub, Effectiveness of slow-release tablet formulations of the IGR Dudim and the bioinsecticide Natular against mosquito larvae of *Aedes aegypti* (L.). *African Entomology*. 2013; **21**(2).
14. Sharma, R.S.; Kaul, S.M. and Sokhay, J., Fluctuations of dengue vector, *Aedes aegypti* (Diptera: Culicidae) in Delhi, India. *Southeast Asian. J. Trap. Med. Public. Health* 2005; **36**(1): 186-90.
15. Siddigui, B.S.; Ali, S.K.; Naqvi, S.N. and Tariq, R.M., Variation of major limonoids in *Azadirachta indica* fruits at different ripening stages and stages and toxicity against *Aedes aegypti*. *Nat. Prod. Commun.* 2009; **4**(4): 473-6.
16. Vasuki, V., Influenced of IGR treatment on oviposition of three of vector mosquitoes at sublethal concentrations. *South Asian J. Tropical Medicine and Public Health* 1999; **30**: 200-203.
17. World Health Organization., Instructions for determining the susceptibility or resistance of mosquito larvae to insecticides. WHO/VBC. 1981; **807**: 1-6.
18. World Health Organization., Vector control for malaria and other mosquito borne diseases. Rept of WHO study group. Technical Rep. Ser. No. 857. WHO, Geneva, 1995.
19. World Health Organization., Guidelines for laloratory and field testing of mosquito larvicides. WHO/CDS/WHOPES/CDPP/13, 2005.
20. Xu, Q.; Wang, H.; Zhang, L. and Liu, N., Kdr allelic variation in pyrethroid resistant mosquito, *Culex quinquefasciatus*. *Biochem. Biophys. Res. Commun.* 2006; **345**: 774-780.