

Review on the Bioremediation by *Aspergillus niger*

S.M. Yakout^{1,2*}

¹Department of Biochemistry, College of Science, King Saud University,
PO Box, 2455, Riyadh, 11451, Kingdom of Saudi Arabia.

²Atomic Energy Authority, Hot Laboratories Centre, 13759, Egypt.

(Received: 20 October 2013; accepted: 11 December 2013)

Aspergillus niger (*A. niger*) is one of the most important microorganisms used in biotechnology. It has been in use already for many decades to produce extracellular (food) enzymes and citric acid. In addition, *A. niger* is used for biotransformations and waste treatment in the last two decades. This review summarizes available information on *A. Niger* derived biomass for bioremediation of different types of pollutants include metals, radionuclides, pesticides, phenols and dyes. It is clear that *A. niger* can be used to prepare biomass, that can be applied in various aqueous treatment processes, namely to remove organic pollutants, dyes, organic compounds, radionuclides and heavy metals.

Key words: *Aspergillus niger*, Bioremediation, biomass.

Bioremediation refers to the use of microorganisms to eliminate or reduce the concentrations of hazardous wastes at a contaminated site¹. Bioremediation is often considered a cost effective and environmental friendly method and is gradually making roads for environmental clean-up applications. Bioremediation relies on improved detoxification and degradation of toxic pollutants either through intracellular accumulation or via enzymatic transformation to less or non-toxic compounds². Many microorganisms naturally possess the ability to degrade, transform, or chelate various toxic chemicals. Fungi possess the biochemical and ecological capacity to degrade environmental organic chemicals and to decrease the risk associated with metals, metalloids and

radionuclides, either by chemical modification or by influencing chemical bioavailability.

A.niger is a dark colored filamentous fungus belonging to phylum Ascomycota (Fig. 1). *A. niger* has been the subject of research and industrial use for several decades. It first acquired practical importance in 1919, when its ability to produce citric acid was industrially exploited. Since the 1960s, *A. niger* has become one of the most important industrial microorganisms and produces a variety of enzymes such as cellulase and xylanase³, phytases⁴, amylases^{5,6}, lipase⁷ and peptidases⁸. Enzymes from *A. niger* have been used in food production for several decades, and even though their peptidases have been studied⁹

A. niger plays a significant role in the global carbon cycle, As a common member of the microbial communities found in soils. Moreover, *A. niger* has a remarkably versatile metabolism, enabling growth on a wide range of substrates under various environmental conditions. Its ability to degrade a range of xenobiotics through various oxidation, hydroxylation, and demethylation reactions provides immense potential for use in bioremediation¹⁰⁻¹².

* To whom all correspondence should be addressed.
Tel.: +966558448693; Fax: +96614675931;
E-mail: sobhy.yakout@gmail.com

Fungi are recognized for their superior ability to produce a large variety of extra cellular proteins, organic acids, enzymes and other metabolites, and their waste biomass may be used as effective biosorbent for removal, reduction and detoxification of industrial effluents ingredients¹³. The analysis of cell-wall composition of *A.niger* biomass has been studied by Johnston, IR, 1965¹⁴. Sugars such as, glucose, galactose, mannose, arabinose, glucosamine and galactosamine, all in the d-configuration and the small amount of l-galactose have been found in the cell wall analysis. According to Johnston report¹⁴, the main structure of cell wall contains carbohydrate and hexosamine and small amounts of lipid and protein.

Large amounts of biomass are produced during the citric acid fermentation process and finding the best way for its consumption is well worth considering. Fungal biomass seems to be a very efficient and inexpensive sorbent that can be produced at low cost by using simple fermentation techniques and cheap growth media. It is also available as a by-product or waste material from various industrial processes¹⁵. The aim of this study is the investigation of some applications of *A. niger* biomass.

Heavy Metals and radionuclides

Wastewater treatment containing heavy metal ions is one of the major concerns in environmental cleaning. There are various treatment methods, e.g. chemical precipitation, evaporation, coagulation–flocculation, flotation, ion exchange, membrane filtration, and adsorption that can be employed to remove heavy metals from contaminated wastewater¹⁶. All these conventional methods have some advantages and disadvantages

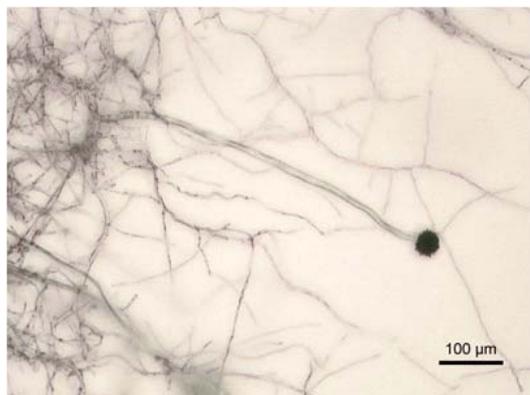


Fig. 1. Micrograph of *A. niger*

such as high consumption of reagent and energy, low selectivity, high operational cost and generation of secondary pollutants¹⁷. Therefore, environmental and public health engineers have been searching for an inexpensive and efficient treatment for metal-containing wastewaters. Nowadays use of renewable resources like microbes and plants (bioremediation and phytoremediation) to tackle heavy metal problems is a feasible and useful treatment method leading to detoxification of industrial effluents and wastewaters¹⁸. Recently, the use of some fungal biomasses for metal elimination has been investigated by some studies¹⁹⁻²¹. The use of dead biomass appears to be more attractive as compared with live biomass, since heavy metals or other toxic pollutants have few influences on the biosorption process. Besides, there is no requirement for the supplement with any nutrients for maintaining growth of dead biomass²². The cell walls of these types of biosorbents containing polysaccharides

Table 1. Bioremediation of Heavy Metals and radionuclides using *A. niger* biomass

Heavy Metals and radionuclides	References
Lead, zinc, cadmium, chromium, copper, nickel	[30, 35]
Cobalt, lead, cadmium, chromium, copper, nickel	[36]
Copper, manganese, zinc, nickel, iron, lead, cadmium	[37]
Copper and zinc	[38]
Copper, lead	[39]
Lead, copper	[40]
Zinc	[41, 42]
Nickel	[43]
Copper	[44]
Chromium	[31, 45, 46]
Uranium	[32]
Thorium	[33]
Radioactive cobalt	[34]

Table 2. Bioremediation of organics, pesticides and PAH using *A. niger* biomass

Phenol	[50]
Pentachlorophenol	[49]
Endosulfan	[48, 54]
Phenanthrene	[47, 59]
Chrysene and benzo(a)pyrene	[55]

and proteins with different functional groups such as amine, carboxyl, hydroxyl, sulfates, and phosphates responsible for interaction with metal ions²².

A. niger has been used to remove metals from the environment by either adsorption of the metals to fungal cell wall components, or complexation of the metals with organic acids produced by the fungus as shown in Table 1. The fungus has been used commercially in the production of organic acids, such as citric acid²³, oxalic acid²⁴ and gluconic acid²⁵. These acids are well-known lixiviants for the leaching of heavy metals from solid wastes²⁶, and may reduce the cost of commercial heavy metal decontamination and decrease any environmental impacts resulting from metal contamination²⁷. Metal leaching by heterotrophic micro-organisms generally involves an indirect process with microbial production of amino acids, other organic acids, and other metabolites. These metabolites dissolve metals from minerals by displacement of metal ion from the ore or soil matrix by hydrogen ions, or by the formation of soluble metal complexes and chelates²⁸. *A. niger* has been found to overproduce organic acids that can serve as leaching agents for the solubilisation of Al, Fe, Mn, Ni, Pb, Cd, Cu, and Zn from fly ash²⁹. *A. niger* was used for the leaching of heavy metals from contaminated soils²⁸. *A. niger* are used for chromium biosorption through ion exchange mechanism³⁰. *A. niger* strain, isolated from soil of leather tanning effluent, has been found to remove chromium very effectively from liquid medium as well as from soil microcosm with different concentrations of chromate³¹. The ability of *A. niger* biomass to bind and remove heavy metals, i.e. Ni, Pb, Cr from real wastewater was investigated. Also *A. niger* was used for the removal of radionuclides such as uranium, and thorium^{32, 33}. Bioremediation of ⁶⁰Co from simulated spent decontamination solutions by *A. niger* was reported³⁴.

Organics

In modern society, an increasing number of hazardous organic compounds are being discharged into the environment. Most are degraded or detoxified by physical, chemical and biological treatments before released into the environment. Although the biological treatments are a removal process for some organic

compounds, their products of biodegradation may also be hazardous. Moreover, some nondegradable compounds discharged into the environment along with the treated compounds can cause problems because they usually come back to human beings through the several channels such as bioaccumulation. As a result, organic molecules that are not biodegradable, can still be removed from the wastewater by the microbial biomass via the process of biosorption. Biosorption is also becoming a promising alternative to replace or supplement the present removal processes of organic pollutants from wastewaters. Among these pollutants, phenolics and pesticides have recently been of great concern because of the extreme toxicity and/or persistency in the environment. Biosorption of these type of hazardous organics by selected live and dead microorganisms has been investigated by various workers⁴⁷⁻⁵⁰.

Phenol is an important toxic compound listed as a priority pollutant by the EPA and other environmental protection agencies because of its high toxicity and possible accumulation in the environment. Mathialagan and Viraraghavan

Table 3. Bioremediation of Dyes using *A. niger* Biomass

Dyes	
Acid Black 1	[70]
Acid Red, Orange II,	[71]
Acid Blue 2	[68]
Acid Blue 29	[68]
basic Blue 9	[72]
Basic Blue Acid Blue Congo Red	
Disperse Red 1	[72, 73]
Ci direct blue 199	[66]
Congo Red	[49, 69]
Direct Violet,	
Direct Brown and Direct Green	[74]
Direct Blue 199	[66]
Gryfalan black	[75]
Orange II	[76]
Polar Red	[77]
Reactive black 8	[63]
Reactive blue 140	[65]
Reactive Brilliant Red	[52]
Scarlet Red; Greenish Blue; Brilliant Violet	[78]
Sulfur black, acid red 151, dimarene	
blue K2RL, Orange II Synazo	[53]
yellow HF2GR	[64]

studied the biosorption of pentachlorophenol (PCP) on treated *A.niger* biomass. They pretreated live fungal pellicles in five different ways: autoclaving, treatment with 0.1M HNO₃, treatment with 0.1M H₂SO₄, treatment with 0.1M NaOH, treatment with a laboratory detergent (5 g l⁻¹). They found that the biosorption was pH dependent. The removal of PCP decreased with the increase of pH for all types of biomass. They also evaluated the results with different kinetic and isotherm models⁵¹. They observed that sulphuric acid-treated biomass powder was the most effective for the removal of phenol. Sodium hydroxide pretreated and laboratory detergent pretreated biomasses gave very close removals. Alkali pretreatment has also been shown to be a probable cause for destroying autolytic enzymes that cause putrefaction of the biomass and for removing lipids and proteins that mask reactive sites⁵⁰. Gallagher *et al.*,⁵² have also suggested that NaOH pretreatment may also increase the percentage of chitin/chitosan in the whole cell wall fraction by dissolving certain biopolymers from the cell wall. Chitin/chitosan units may then be responsible for the uptake of phenol in the case of NaOH pretreated biomass⁵³.

Pesticides

The chlorinated organic pesticide endosulfan is the major agrochemical responsible for environmental pollution. It enters air, water, and soil environments during its use and manufacturing. Due to its hydrophobic nature, it is also associated with soil. Among different groups of microorganisms, fungi have their own advantages in cultivation, maintenance, and tolerance to pesticides over bacteria, especially for chlorinated pesticides. All of these applications attract scientists to use fungi in the biodegradation process. As per the earlier report of *A.niger* showing good results at broth condition⁵⁴ in endosulfan degradation, endosulfan was degraded in just 120 h. Bhalerao (2012) applied strategy of *in situ* bioremediation by bioaugmentation for soil contaminated with organochlorine pesticide endosulfan in an open pot⁴⁸. It showed that previously isolated fungal culture *A. niger* can be effectively used for endosulfan degradation.

Poly Aromatic Hydrocarbons (PAH)

Previous work⁵⁵ has reported that *A. niger* was efficient degraders of PAHs in high concentrations compared to other non-ligninolytic

fungal strains which produce more toxic intermediaries than the original compound and tolerate lower PAH concentrations, and also compared to ligninolytic fungi not grown in soil and which are displaced by native soil microflora. *Niger* able to transform a variety of PAHs, including chrysene and benzo(a)pyrene, to polar metabolites^{55,56}. This microorganisms carry out a monooxygenation of the PAH molecules by the intracellular cytochrome P-450 dependent oxidase system⁵⁷. These fungi do not produce extracellular peroxidases, however, they do produce cytochrome P450 monooxygenase which can oxidize PAHs to epoxides and dihydrodiols: highly potent carcinogens that accumulate in soil⁵⁸. Other study in which *A niger*, grown on sugarcane bagasse and added to a soil spiked with 400 ppm phenanthrene, achieved 54% removal of the pollutant from the soil after 7 days,⁵⁹.

Dyes

The textile industry is a substantial consumer of water and produces enormous volumes of contaminated water; the most important contaminants are dyes. The complex aromatic structure of the dyes is resistant to light, biological activity, ozone and other degradative environmental conditions. Thus conventional waste water treatment remains ineffective. Also, anionic and nonionic azo dyes release toxic amines due to the reactive cleavage of azo groups⁶⁰. There are various methods for the treatment of textile wastewater for the removal of dye. These broadly fall into three categories: Physical, Chemical and Biological. The major disadvantage of physico-chemical methods has been largely due to the high cost, low efficiency, limited versatility, interference by other wastewater constituents and the handling of the waste generated. Microbial decolorization have the advantage of being cost-effective and environmentally friendly and producing less sludge. The mechanism of microbial decoloration occurs from adsorption, enzymatic degradation or a combination of both.

A. niger is studied more than other biomasses for decolorization of waste water samples (Table 3). The biomass of *A.niger* have been developed for decolorization of waste waters containing, acid blue⁶¹ basic blue 9⁴⁹Congo red⁶² reactive black 8 ⁶³ Sulfur black⁵³, acid red 151, dimarene blue K2RL, orange II ⁵³ yellow HF2GR⁶⁴

from textile effluent, reactive blue 140⁶⁵, Ci direct blue199⁶⁶. Reported results show the efficiency of the *A. niger* biomass for dye removal from wastewaters. Both active and inactive forms of *A. niger* have been applied for afore mentioned researches and investigations. The mechanism of dye adsorption on the surface of fungal biomass is related to electrostatic attraction between negatively charged dyes and positively charged cell walls and this is because of chitin, polysaccharides amino acids and lipids which make the cell wall structure⁶⁷.

Gallagher *et al.*⁵² investigated three types of fungi, including *Laminaria digitata*, *Rhizopus oryzae*, and *A.niger*, to remove Reactive Brilliant Red, a reactive dye suggesting that adsorption was occurred by combined mechanisms onto a heterogeneous surface⁵². Fu and Virarahavan^{68, 69} investigated the removal of Basic Blue 9 (cationic), Acid Blue 29 (anionic), CongoRed (anionic), and Disperse Red 1 (nonionic) dyes from aqueous solutions by biosorption on dead and pretreated *A.niger* fungus. They found that *A. niger* is capable of removing dyes from an aqueous solution. They explored that three major functional groups: carboxyl, amino and phosphate, and the lipid fraction in the biomass of *A.niger* played an important role in the biosorption of these dyes^{68, 69}.

CONCLUSION

This review paper highlights the capacity *A. niger* biomass in different pollutants treatment. We overview about using *A. niger* in the removal of different types of pollutants including metals radionuclides, phenols, PAH, pesticides and dyes.

ACKNOWLEDGMENTS

This project was supported by King Saud University, Deanship of Scientific Research, College of science Research Centre.

REFERENCES

1. De Lorenzo V: Systems biology approaches to bioremediation. *Curr Opin Biotechnol* 2008; **19**: 579–589.
2. Brar S, Verma M, Surampalli R, Misra K, Tyagi R, Meunier N, Blais J: Bioremediation of hazardous wastes: a review. *Pract Periodical Hazard. Toxic Radioactive Waste Manag* 2006; **10**: 59-72.
3. Farinas CS, Loyo MM, Junior AB, Tardioli PW, Neto VB, Couri S, . : Finding stable cellulase and xylanase: evaluation of the synergistic effect of pH and temperature. *New Biotechnology* 2010, **27**, :810-815.
4. Bhavsar K, Kumar VR, Khire JM: High level phytase production by *Aspergillus niger* NCIM 563 in solid state culture: response surface optimization, upscaling, and its partial characterization. *Journal of Industrial Microbiology & Biotechnology* 2011; **38**: 1407-1417.
5. Mitidieri S, Martinelli AHS, Schrank A, Vainstein MH: Enzymatic detergent formulation containing amylase from *Aspergillus niger*: a comparative study with commercial detergent formulations. *Bioresource Technology* 2006; **97**: 1217-1224.
6. Mahalakshmi C, Miller Samson S, Alagendran S, Anusha R, Neelamathi E: Effect of UV and EMS Mutation on *Aspergillus niger* in the Production of a-amylase. *J Pure Appl Microbio.* 2009; **3**(2): 705-708.
7. Anvari M: Optimization of Lipase Production from Rice Straw using Response Surface Methodology by *Aspergillus niger*. *J Pure Appl Microbio.* 2012; **6** .
8. Morya VK, Yadav S, Kim E, Yadav D: In silico characterization of alkaline proteases from different species of *Aspergillus*. *Applied Biochemistry and Biotechnology* 2012; **166**: 243-257.
9. Basten DEJW, Dekker PJT, Schaap PJ: Aminopeptidase C of *Aspergillus niger* is a novel phenylalanine aminopeptidase. *Applied and Environmental Microbiology* 2003; **69**: 1246-1250.
10. Raj HG, Saxena M, Allameh A: Metabolism of foreign compounds by fungi. In: Arora, D.K., Elander, R.P., Mukerji, K.G. (Eds.), *Handbook of Applied Mycology, Fungal Biotechnology*. Marcel Dekker Inc., New York; 1992.
11. Mangalam H, Mahalingam PU, Raja D: A Study on Mycoremediation as a Novel Treatment for Textile Mill Effluent. *J Pure Appl Microbio.* 2010; **4**(1): 183-187.
12. Afroz S, Srivastava AK: Evaluation of some Species of *Aspergillus* for their Biodegradation capability to Degrade Silk Industry Effluent of Reservoir Khajua Lake at Mubarakpur, Azamgarh, (U.P.). *J Pure Appl Microbio* 2007; **1**(1): 83-90.

13. Christian V, Shrivastava R, Shukla D, Modi HA, Vyas BRM: Degradation of xenobiotic compounds by ligninedegrading white rot fungi: enzymology and mechanism involved. *Indian Journal of Experimental Biology* 2005; **43**: 301-312.
14. Johnston I: The composition of the cell wall of *Aspergillus niger*. *Biochem J* 1965; **96**: 651-658.
15. Maurya NS, Mittal AK, Cornel P, Rother E: Biosorption of dyes using dead macro fungi: effect of dye structure, ionic strength and pH. *Bioresour Technol* 2006; **97**: 512-521.
16. Paul S, Bera D, Chattopadhyay P, Ray L: Biosorption of Pb(II) by *Bacillus cereus* M1 16 immobilized in calcium alginate gel. *J Hazard Sub Res* 2006; **5**: 2-13.
17. Chakravarty S, Mohanty A, Sudha TN, Upadhyay AK, Konar J, Sircar JK: Removal of Pb (II) ions from aqueous solution by adsorption using bael leaves (*Aegle marmelos*). *J Hazard Mater* 2009; **170**: 969-977.
18. Ozdemir G, Ceyhan N, Ozturk T, Akirmak F, Cosar T: Biosorption of chromium(VI), cadmium(II) and Cu(II) by *Pantoea* sp. TEM18. *Chem Eng J* 2004; **102**: 249-253
19. Gorgievskia M, Bozica D, Stankovicb V: Kinetics, equilibrium and mechanism of Cu²⁺, Ni²⁺ and Zn²⁺ ions biosorption using wheat straw. *Ecol Eng* 2013; **58**: 113-122
20. Al-Homaidan AA, Al-Houri HJ, Al-Hazzani AA, Elgaaly G, Moubayed NMS: Biosorption of copper ions from aqueous solutions by *Spirulina platensis* biomass. *Arabian J Chem* 2013, <http://dx.doi.org/10.1016/j.arabjc.2013.05.022>.
21. Ghosh A, Das P: Optimization of copper bioremediation by *Stenotrophomonas maltophilia* PD2. *J Environ Chem Eng*, 2013, <http://dx.doi.org/10.1016/j.jece.2013.04.012>.
22. Razmovski R, Sciban M: Biosorption of Cr(VI) and Cu(II) by waste tea fungal biomass. *Ecol Eng* 2008; **34**: 179-186.
23. Legisa M, Gradisnik-Grapulic M: Sudden substrate dilution induces a higher rate of citric acid production by *Aspergillus niger*. *Appl. Environ Microbiol* 1995; **61**: 2732-2737.
24. Strasser H, Burgstaller W, Schinner F: High yield production of oxalic acid for metal leaching processes by *Aspergillus niger*. *FEMS Microbiol Lett* 1994; **119**: 365-370.
25. Dronawat SN, Svihla CK, Hanley TR: The effects of agitation and aeration on the production of gluconic acid by *Aspergillus niger*. *Appl. Biochem. Biotechnol* 1995; 51-52.
26. King AB, Dudeney AWL: Bioleaching of nephelin. *Hydrometallurgy* 1987; **19**: 69-81.
27. Mulligan CN, KamLi M, Gibbs BF: Bioleaching of heavy metals from a low grade mining ore using *Aspergillus niger*. *J Hazard Mater* 2004; **110**: 77-84.
28. Ren WX, Li PJ, Geng Y, Li XJ: Biological leaching of heavy metals from a contaminated soil by *Aspergillus niger*. *J Hazard Mater* 2009; **167**: 164-169.
29. Bosshard PB, Bachofen R, Brandl H: Metal leaching of fly ash from municipal waste incineration by *Aspergillus niger*. *Environ Sci Technol* 1996; **30**: 3066-3070.
30. Ahluwalia S, Goyal D: Microbial and plant derived biomass for removal of heavy metals from wastewater. *Bioresour Technol* 2006; **98**: 2243-2257.
31. Srivastava S, Thakur IS: Evaluation of bioremediation and detoxification potentiality of *Aspergillus niger* for removal of hexavalent chromium in soil microcosm. *Soil Biol Biochem* 2006; **38**: 1904-1911.
32. Abbasi WA, Streat M: Adsorption of Uranium from Aqueous Solutions Using Activated Carbon. *Separation Science and Technology*, 1994, 29 1217-1230.
33. Tsezos M, Volesky B: Biosorption of uranium and thorium. *Biotechnology and Bioengineering* 1981; **23**: 583-604.
34. Rashmi K, Sowjanya TN, Mohan PM, Balaji V, Venkateswaran G: Bioremediation of 60Co from simulated spent decontamination solutions. *Sci Total Environ* 2004; **328**: 1-14.
35. Mukherjee I, Gopal M: Degradation of chlorpyrifos by two soil fungi *Aspergillus niger* and *Trichoderma viride*. *Environ Toxicol Chem* **37**: 1996.
36. Dwivedi S, Mishra A, Saini D: Removal of Heavy Metals in Liquid Media through Fungi Isolated from Waste Water *International Journal of Science and Research* 2012; 1.
37. sekova K, Todorova D, Ganeva S: Removal of heavy metals from industrial wastewater by free and immobilized cells of *Aspergillus niger*. *International Biodeterioration and Biodegradation* 2010; **64**: 447-451.
38. Price MS, Classen JJ, Payne GA: *Aspergillus niger* absorbs copper and zinc from swine wastewater. *Bioresour Technol* 2001; **77**: 41-49.
39. Iskandar N, Zainudin N, Tan S: Tolerance and biosorption of copper (Cu) and lead (Pb) by filamentous fungi isolated from a freshwater ecosystem. *Journal of Environmental Sciences* 2011; **23**: 824-830.
40. Kapoor A, Viraraghavan T, Cullimore DR: Removal of heavy metals using the fungus

- Aspergillus niger*. *Bioresour Technol* 1999; **70**: 95-104.
41. Saraswat S, Rai JPN: Heavy metal adsorption from aqueous solution using *Eichhornia crassipes* dead biomass. *International Journal of Mineral Processing* 2010; **94**: 203-206.
 42. Sutjaritvorakul T, Whalley AJS, Roengsumran S, Sihanonth P: Solubilization and Accumulation of Insoluble Zinc and Lead Compounds by Fungi Isolated from Zinc Mine. 2013; **2**: 1043-1046.
 43. Magyarosy A, Laidlaw R, Kilaas R, Echer C, Clark D, Keasling J: Nickel accumulation and nickel oxalate precipitation by *Aspergillus niger*. *Appl Microbiol Biotechnol* 2002; **59**: 382-382.
 44. Dursun AY, Uslu G, Tepe O, Cuci Y, Ekiz HI: A comparative investigation on the bioaccumulation of heavy metal ions by growing *Rhizopus arrhizus* and *Aspergillus niger*. *Biochem Eng J* 2003; **15**: 87-92.
 45. Shugaba A., Buba F., Kolo BG., Nok AJ., Ameh DA., JA. L: Uptake and Reduction of Hexavalent Chromium by *Aspergillus niger* and *Aspergillus parasiticus*. *J Pet Environ Biotechnol* 2012; **3**: 119.
 46. Shaili S, Indu ST: Evaluation of bioremediation and detoxification potentiality of *Aspergillus niger* for removal of hexavalent chromium in soil microcosm. *Soil Biology & Biochemistry* 2006; **38**.
 47. Cortés-Espinosa DV, Absalón ÁE: Phenanthrene Removal from Soil by a Strain of *Aspergillus niger* Producing Manganese Peroxidase of *Phanerochaete chrysosporium*. In *Hydrocarbon*. CC BY 3.0 license InTech, DOI: 10.5772/51944; 2013
 48. Bhalerao TS: Bioremediation of endosulfan-contaminated soil by using bioaugmentation treatment of fungal inoculant *Aspergillus niger*. *Turk J Biol* 2012; **36**: 561-567.
 49. Fu Y, Viraraghavan T: Column studies for biosorption of dyes from aqueous solutions on immobilized *Aspergillus niger* fungal biomass. *Water South Africa* 2003; **29**: 465-472.
 50. Rao J, Viraraghavan T: Biosorption of phenol from an aqueous solution by *Aspergillus niger* biomass. *Bioresour Technol* 2002; **85**: 165-171.
 51. Mathialagan T, Viraraghavan T: Biosorption of chlorophenols: a review. *Int J Environ Pollut* 2008; **34**: 164-194.
 52. Gallagher KA, Healy MG, Allen SJ: *Biosorption of synthetic dye and metal ions from aqueous effluents using fungal biomass.*: In: Wise, D.L. (Ed.), *Global Environ. Biotechnol*. Elsevier, UK, ; 1997.
 53. Khalaf M: Biosorption of reactive dye from textile wastewater by non-viable biomass of *Aspergillus niger* and *Spirogyra* sp. *Bioresour Technol* 2008; **99**: 6631-6634.
 54. Bhalerao T, Puranik P: Biodegradation of organochlorine pesticide, endosulfan, by a fungal soil isolate *Aspergillus niger*. *Int Biodeter Biodegr* 2007; **59**: 315-332.
 55. Cortés-Espinosa DV, Fernández-Perrino FJ, Arana-Cuenca A, Esparza-García JF, Loera O, Rodríguez-Vázquez R: Selection and identification of fungi isolated from sugarcane bagasse and their application for phenanthrene removal from soil. *J Environ Sci Health, Part A* 2006; **41**: 475-486.
 56. Cerniglia CE, White GL, Helflich RH: Fungal metabolism and detoxification of polycyclic aromatic hydrocarbons. *Arch Microbiol* 1985; **143**: 105-110.
 57. Gramss G, Voigt K, Kirsche B: Degradation of polycyclic aromatic hydrocarbons with three to seven aromatic rings by higher fungi in sterile and unsterile soils. *Biodegradation* 1999; **10**: 51-62.
 58. Shimada T: Xenobiotic-metabolizing enzymes involved in activation and detoxification of carcinogenic polycyclic aromatic hydrocarbons. *Drug Met Pharmacokinet* 2006; **21**: 257-276.
 59. Chávez-Gómez B, Quintero R, Esparza-García F, Mesta-Howard A, de la Serna FJZD, Hernández-Rodríguez C, Gillen T, Poggi-Varaldo H, Barrera-Cortés J, Rodríguez-Vázquez R: Removal of phenanthrene from soil by co-cultures of bacteria and fungi pregrown on sugarcane bagasse pith. *Bioresour Technol* 2003; **89**: 177-183.
 60. Joshi M, Bansal R, Purwar R: Colour removal from textile effluents. *Ind J Fibre Textile Res* 2004; **29**: 239-259.
 61. Fu Y, Viraraghavan T: Removal of Acid Blue 29 from an aqueous solution by fungus *Aspergillus niger*. In *31st MidAtlantic Ind and Hazardous Waste Conf* (Nikolaidis N ECaSB ed. pp. 510-519. Lancaster, Pennsylvania:: Technomic Publishing Co., 1999: 510-519.
 62. Fu Y, Tiraraghavan Y: Removal of Congo red from an aqueous solution by fungus *Aspergillus niger*. *Adv Environ Res* 2004; **7**: 239-247.
 63. Ali N, Hameed A, Ahmed S, Khan A: Decolorization of structurally different textile dyes by *Aspergillus niger* SA1. *World J Microbiol Biotechnol* 2008; **24**: 1067-1072.
 64. Paymann M, Mehnaz M: Decolorization of textile effluent by *Aspergillus niger* (marine and terrestrial). *Fresen Environ Bull* 1998; **7**: 1-7.
 65. Nanthakumar K, Karthikeyan K, Lakshmanaperumalsamy P: Investigation on Biosorption of Reactive Blue 140 by Dead

- Biomass of *Aspergillus niger* HM11: Kinetics and Isotherm Studies. *Global Journal of Biotechnology & Biochemistry* 2009; **4**:169-178.
66. Xiong X-J, Meng X-J, Zheng T-L: Biosorption of C.I. Direct Blue 199 from aqueous solution by nonviable *Aspergillus niger*. *J Hazard Mater* 2010, **175**:241-246.
 67. Aksu Z, Tezer S: Equilibrium and kinetic modelling of biosorption of Remazol Black B by *Rhizopus arrhizus* in a batch system: effect of temperature. *Process Biochem* 2000; **36**: 431–439.
 68. Fu Y, Viraraghavan T: Removal of Acid Blue 29 from an aqueous solution by *Aspergillus niger*. *Am Assoc Text Chem Color Rev* 2001; **1**: 36-40.
 69. Fu Y, Viraraghavan T: Dye biosorption sites in *Aspergillus niger*. *Biores Technol* 2002; **82**: 139-145.
 70. Kanagaraj J, Mandal A: Combined biodegradation and ozonation for removal of tannins and dyes for the reduction of pollution loads. *Environ Sci Pollut Res Int* 2012; **19**: 42-52.
 71. Ali N, Hameed A, Siddiqui M, Ghumro P, Ahmed S: Application of *Aspergillus niger* SA1 for the enhanced bioremoval of azo dyes in simulated textile effluent. *Afr J Biotechnol* 2009; **8**: 3839-2845.
 72. Fu YZ, Viraraghavan T: Removal of a dye from an aqueous solution by the fungus *Aspergillus niger*. *Water Qual Res J Can* 2000; **35**: 95-111.
 73. Fu Y, Viraraghavan T: Removal of C.I. Acid Blue 29 from an aqueous solution by *Aspergillus niger*. 2001. *AATCC Mag* 2001; **1**: 36-40.
 74. Wafaa M, El-Rahim A, Moawad H: Testing the performance of small scale bioremediation unit designed for bioremoval/enzymatic biodegradation of textile azo dyes residues. *New York Sci J* 2010; **3**: 77-92.
 75. Aksu Z, Karabayir G: Comparison of biosorption properties of different kinds of fungi for the removal of Gryfalan Black RL metal-complex dye. *Bioresour Technol* 2008; **99**: 7730-7741.
 76. Ali N, Hameed A, Siddiqui M, Ghumro P, Ahmed S: Application of *Aspergillus niger* SA1 for the enhanced bioremoval of azo dyes in simulated textile effluent. *Afr J Biotechnol* 2009; **8**: 3839-3845.
 77. Abd El-Rahim WM, Moawad H: Enhancing bioremoval of textile dyes by eight fungal strains from media supplemented with gelatine wastes and sucrose. *J Basic Microbiol* 2003; **43**: 367-375.
 78. Laxminarayana E, Chary M, Kumar M, Charya M: Decolourisation and biodegradation of sulphonated azo dyes by fungi to clean dye contaminated soil environments. *J Nat Environ Sci* 2010; **1**: 35-42.