Occurrence of Keratinophillic Fungi and Other Dermatophytes from Soils of Various Habitats of Hyderabad, A.P (India)

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The main goal of the present study was to isolate and identify keratinophilic fungi and related dermatophytes in different soil of Hyderabad. Soil of school playground, public parks, agricultural area and hospitals, which are frequently visited by children, adults and occupational workers, were chosen for the study. A total of 112 soil samples of various habitats were screened for the revalence of these fungiby hair-baiting technique, of which 99 samples showed positive growth on hair baits. Public parks showed maximum number of positive samples (97%) followed by school playgrounds (88%), agricultural soils (84%) and hospitals (78%) respectively. Our findings revealed the presence of 242 fungal isolates from soils of various habitats. These isolates represented 30 species of 16 genera, predominated by six Chrvsopsorium species followed by four species of Trichophyton and three species of Microsporum and Aspergillus each, besides a single species representing the genera Alternaria, Aphanoascus, Bipolaris, Fusarium, Cladosporium, Ctenomyces, Geotrichum, Malabranchea, Mucor, Rhizopus, Paecilomyces, Penicillium and Scopulariopsis. The high prevalence of keratinophilic fungi along with dermatophytes are important bioindicators of soil contamination with keratin remnants.Most of these fungi are potential pathogens causing mycoses and are a threat to all those individuals visiting or working in such environments. To our knowledge, this appears to be the first report concerning the isolation of keratinophilic fungi from Hyderabad soils.

Key words: Keratinophilic fungi, Dermatophytes, Public parks, School playgrounds, Agricultural soil, Hospital soil.

The ubiquity of fungi in soil and various other environments is well recognized. However, certain groups of fungi are very specific about their substrate and nutritional requirement, among them keratinophilic fungi occupy an important place. Majority of these represent a host of filamentous fungi comprising hyalohyphomycetes and many other taxonomic groups¹. Hyphomycetes include the dermatophytes, and many other nondermatophytic fungi mainly occurring as saprophytes in soil, and some are known plant pathogens². They have the ability to colonize and degrade various keratinous substrates like skin, nail, hair, hooves, wool and other cornified appendages in various soil environment completely. Hence, they are important ecologically, as they cycle one of the most abundant and highly stable animal proteins on earth i.e. keratins³. Keratinolytic fungi occur in many natural and manmade habitats and exist in communities together with keratinophilic fungi, that have weaker affinity to keratin and utilize chiefly the products of its decomposition⁴. While dermatophytes are known to cause dermatophytosis which are superficial cutaneous infections of keratinized tissues i.e skin, hair and nails of humans, animals and birds. They create inflammation, pruritus and desquamation by invading the keratin of the stratum corneum. These infections are contagious

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and are have been reported from different parts of the world.

The modern Hyderabad is spread over an area of 650 km² (250 sq mi), making it one of the largest metros in India. The city is nestled on Deccan Plateau and rises to an average height of 540 m above the sea level. The city lies at 17.366° N latitude and 78.476° E longitude. Summer day temperatures rise to a maximum of 44°C often in May and night temperatures in winters may fall to a minimum of 12 °C. Thus the metropolitian is quite dry and hot from February to May, much humid from end of June to September and mildly cool from October to January. Soils of locations with such tropical climatic conditions are known to serve as good habitat for the growth of keratinophilic fungi and dermatophytes. Several researchers have explored soils of India from different states . These fungi were isolated from Indian soils from different habitats viz. public places^{5,3}. Agricultural soils and parks⁶, water sediments⁷, sewage waters⁸, Hilly areas9 and birds and their environment10. However, to the best of our knowledge, soils of Hyderabad city has not been explored yet for the prevelance of keratinophilic fungi. Our attempt in the present work was to explore soil of different habitats, for the occurrence of keratinophilic fungi and related dermatophytes. This study will help children, adults, farmers and gardeners who frequent these places and are at a risk of exposure to these potential pathogens through direct soil contact. To our knowledge this is the first report on the distribution of keratinophilc fungi and related dermatophytes from Hyderabad soil.

MATERIALSAND METHODS

Collection of soil samples

One hundered and twelve soil samples were collected from various locations. The superficial layer of soil at a depth not exceeding 2-5 cm was scooped and placed in sterile polyethylene bags, brought to the laboratory, and stored at 4°C for a maximum of two weeks. I thank Mr. Ibrahim and Mr. Ashok for their assistance during soil samples collection.

Isolation of Keratinophilic fungi

Keratinophilic fungi were isolated using the hair baiting technique¹¹. Sterile Petri dishes were half filled with thoroughly homogenised soil samples and the soil was moistened with sterile distilled water; the amount of water added varied from sample to sample, depending on the moisture content of the sample. It was then baited with short strands of sterilized human hair. Strands of human hair were spread uniformly as baits on the surface of moistened soil samples. The plates were then incubated at 28±2°C and then examined daily from day 5 for any fungal growth over a period of 4 -8 weeks. Five replicates were set for each sample. Samples of hair showing fungal growth were carefully observed, and then aseptically picked up and cultured on Sabourauds dextrose agar (SDA, Scharlau Chemie, Spain) supplemented with chloramphenicol (0.05mg/ml) alone and another set with chloramphenicol (0.05 mg/ml) and cycloheximide (0.5 mg/ml) respectively. They were sub-cultured to get pure cultures.

Identification

Identification of the purified fungal isolates was based on the monographs of (12-15) using macro- and micro-morphological features.

RESULTS

The results of isolations on hair baits are presented in Table 1. Out of 112 soil samples collected only 99 samples showed positive growth. Public parks showed maximum number of positive samples (97%) followed by school playgrounds (88.88%), agricultural soils (78.26%) and hospitals (84.21%) respectively. Our findings revealed the presence of 242 fungal isolates from soils of various habitats. These fungi represented 30 species of 16 genera. Public parks showed presence of 80 fungal isolates followed by 61 isolates from school playground, 58 isolates from agricultural area and 43 isolates from hospital soils. Among the important genera dominating Hyderabad soils, Chrysosporium was the most common with 97 isolates; the others being 31 isolates of Trichophyton ,Microsporum,Aspergillus 25 isolates each and 18 isolates of Fusarium. Table 2, Fungal isolates were predominated by six Chrysopsorium species followed by four species of Trichophyton and three species of Microsporum and Aspergillus each. graph-1,2 The percentage distribution of the keratinophillic fungi and dermatopytes are shown in Graphs(1,2). Soils of Hyderabad harboured a variety of keratinophillic fungi in comparison to dermatophytes .*C.tropicum* (14.88%) was found to be the most dominating species representing keratinophillic fungi while it was *M.gypseum* (7.02%) amongst the dermatophytes.Percentage distribution of some important fungi among others are as follows; *C*.*indicum* (9.92%),*C.keratinophillum*(7.85%), *Aspergillus niger* (6.20%), *Trichophyto*

mentagrophytes (5.79%), Penicillium spp (4.13%), Aspergillus flavus (3.31%), Aphanoascus fulvescens and Trichophyton rubrum (2.89%) each. Almost all the fungi isolated were present in Public parks. C.tropicum, C.indicum and C.keratinophillium was present in large numbers in all soils screened.

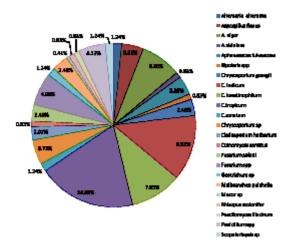
Table 1. Percent prevalence of keratinophilic fungi and related dermatophytes in soil samples of various habitats

Soil collection site	Public parks	School playground	Hospital	Agricultural area	Total
No .of samples examined	34	36	23	19	112
No .of samples positive	33	32	18	16	99
Distribution (%)	97	88.88	76.26	84.21	88.39

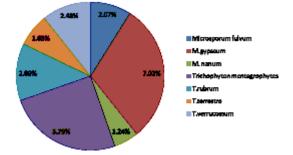
Fungi isolated	Public parks	School playground	Hospital	Agricultural area	Total			
Alternaria alternata	1	0	0	2	3			
Aspergillus flavus	2	3	1	2	8			
A. niger	5	3	5	2	15			
A.nidulans	1	0	0	1	2			
Aphanoascus fulvescens	2	1	4	0	7			
Bipolaris spp	1	1	0	0	2			
Chrysosporium georgii	2	1	1	2	6			
C. indicum	7	5	4	8	24			
C. keratinophilum	6	5	3	5	19			
C.tropicum	10	12	6	8	36			
C.zonatum	0	2	0	1	3			
Chrysosporium sp	3	2	0	4	9			
Cladosporium herbarium	2	3	0	0	5			
Ctenomyces serratus	2	0	0	0	2			
Fusarium solani	2	1	2	1	6			
Fusarium spp	5	2	1	4	12			
Geotrichum sp	2	0	0	1	3			
Microsporum fulvum	2	2	0	1	5			
M.gypseum	6	5	2	4	17			
M. nanum	1	2	0	0	3			
Malbranchea pulchella	2	0	0	4	6			
Mucor sp	0	1	0	0	1			
Paecilomyces lilacinum	1	1	0	0	2			
Penicillium spp	4	2	1	3	10			
Rhizopus stolonifer	0	0	2	0	2			
Scopulariopsis sp	2	1	0	0	3			
Trichophyton mentagrophytes	4	3	5	2	14			
T.rubrum	2	2	3	0	7			
<i>T.terrestre</i>	1	1	0	2	4			
T.verrucossum	2	0	3	1	6			
Total	80	61	43	58	242			
Percent frequency	33.1%	25.2%	17.8%	24.0%	100.0%			

 Table 2. Frequency distribution of keratinophillic fungi and related dermatophytes in various soils of Hyderabad

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Graph 1. Percentage distribution of keratinophillic from various soils of Hyderabad



Graph 2. Percentage distribution of dermatophytes in soils of Hyderabad

DISCUSSION

Throughout the world great interest is shown by research workers in soil mycoflora that can degrade keratinized residues. Every keratinophilic fungi can be considered a potential pathogen and its epidemiological study gains importance firstly because of the extreme resistance of keratin to biological attack, and secondly it's pathogenic potential¹⁶.

Our findings reveal that soils of Hyderabad to be a rich source of both the keratinophilic fungi and dermatophytes. The high positivity rates of fungal isolation (97%) from Public parks and School playground (88%) could be attributed to abundant keratinic material mainly in the form of hairs and feathers. Plant litter, soil pH, moisture content, organic matter and manure besides the keratinic debris in the soil of Public parks and gardens provide the most congenial

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environment for these fungi to proliferate.

The high percentage prevalence of the genus Chrysosporium in our study is indicative of the fact that it can withstand wide range of temperatures and can survive in soils of varying keratin load. It has been reported that the wide spread distribution of the genus Chrysosporium could be attributed to its mesophilic, thermotolerant and hydrophilic nature¹⁷. Occurrence of species of Chrysosporium was reported from varied habitats of Indian soil, Srivasata¹⁸, reported 14 species of the genus Chrysosporium from paddy fields, besides this there are reports of its occurrence from public parks, indoor and outdoor hospital soils and other habitats therefore it shows that this fungi dominates Indian soil mycobiota as it is well adapted to warmer conditions^{19,20-21}. Incidence of infection with different species of Chysoporium have increased in the past few years. Chrysosporium zonatum was reported to cause disseminated infection in a patient with chronic granulomatous disease²². Non sporulating Chrysopsorium sp was found to cause invasive infection in a Neutropenic patient²³. Lyskova²⁴ isolated Chrysosporium queenslandicum, C. sulfureum, C. tropicum, Malbranchea pulchella, Myriodontium keratinophilum from infections of the skin and nails of patients in the Moravian-Silesian Region (Czech Republic). Chrysosporium keratinophilum a fungus present in cultivated soil was isolated from a case of fungal infection of skin and nail plates²⁵. Saidi et al., reported comb lesion in two different breeds of chicken in India by*Chrysosporium tropicum*²⁶.

We isolated three species of Aspergillus, Fusarium solani, Cladosporium herbarium, Alternaria alternata, Chaetomium sp, Mucor sp and other keratinophilic fungi from soil public parks, Agricultural soils and school playground .However many of these fungal species were absent in soil of hospitals, which could be due to competitive utilization of keratinic material by dermatophytes and other competent keratinophillic fungi present in these soils.Indian soil habitats like parks, school playground, indoor and outdoor soils of hospitals have shown the presence of keratinophilic fungi like Aspergillus, Mucor, Rhizopus Penicillum, Trichoderma, Malranchea fulva, Penicillium expansum and and many others in varying numbers^{20,27}.

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Worldwide fungal infections that were not previously recognised as pathogenic have become increasingly common^{28,29}. Fusarium cause a broad spectrum of infections in humans including superficial infections, such as keratitis and onychomycosis, as well as locally invasive and disseminated infections^{30,31,32}. Aspergillus spp are commonly isolated from the soil, plant debris, and the indoor environment, including hospitals. They may cause lifethreatening infections especially in immunocompromised hosts³³.Some species of Aspergillus cause cutaneous aspergillosis³⁴, while in most cases, Aspergillus is introduced to the lower respiratory tract by inhalation of the infectious spores and Aspergillus infection may also disseminate aematogenously to other organs, including the brain³⁵. Some species of Cladosporium are known to be the cause of cerebral and cutaneous phaehyphomycoses³⁶. Isolation of these fungi in our study from various soils is a sign of alarm to those individuals working in these environments.

Present study however revealed fungal isolates belonging to nondermatophyte to be more common than dermatophytes. Among the dermatophytes isolated, the geophilic dermatophyte M .gypseum was the most predominant followed by Trichophyton mentagrophytes, T. rubrum, T. terestre and T.verrucossum, although M.fulvum and M.nanum were isolated but in low numbers. Dermatophytic infections are common disorders worldwide and dermatophytes represent those type of fungi that cause infection of the hair, skin and nails³⁷⁻³⁹. Variety of clinical manifestations are represented by Tinea infections besides Onychomycosis. The high prevalence of superficial mycotic infections makes these one of the most frequent forms of infection.

Pathogens responsible for skin mycoses are primarily anthropophilic and zoophilic dermatophytes and few geophilic from the genera *Trichophyton, Microsporum* and *Epidermophyton* . *Trichophyton rubrum, T. mentagrophytes, M.gypseum M. canis, M. audouinii, T. tonsurans* and *T. verrucosum* are the most common, but the attack rates and incidence of specific mycoses can vary widely. *M.gypseum* causes ringworm of the scalp and glabrous skin in human and animals^{40,41,42}.

There appears to be considerable interand intra-continental variability in the global incidence of these fungal infections. Several workers have screened children's and adults of all ages for dermatophytosis all over the world. Ndako et al., screened school children in the age group of 5-13 years and reported infection from 91% children. Both dermatophytes and nondermatophytes were isolated. The etiological agents of dermatophycoses were *M. canis*, *T. verrucosum*, T. rubrum, T. mentagrophyte and many others. A. flavus, A. niger, Penicillium sp, Mucor sp, Trichoderma sp. and A. fumigatus constituted the non-dermatophytes associated with cutaneous infections. They reported a higher frequency of dermatophytosis occurred more in children with greater propensity for play, interaction with domestic animals⁴³. Similarly several dermatophytes were isolated from 162 children of nomadic herdsmen who were in a continuous exposure to different animals⁴⁴. Many zoophilic and geophilic fungi from genera Acremonium, Fusarium and Aspergillus were cultured occasionally from infected nail plate of farmers⁴⁵.

Hence amongst the wide spectrum of fungal species that was isolated from soils of Hyderabad,most are pathogens causing skin lesions and mycotic infections in birds,animals and humans. Therefore individuals like Farmers, Gardeners and children who are in constant exposure to soil habitats are exposed to these pathogens on regular basis. Additionally ocupations which cause prolong exposure to specific working conditions like humid environments with varying keratin loads in gardens and agricultural fields further expose them to a greater risk of invasion by these fungi.

CONCLUSION

This study revealed that all the soils screened were contaminated with dermatophytes, Keratinophillic and keratinolytic fungi. The high percentage prevalence in public parks could be due to invasion of these places by animals like dogs, cats, pigs, cows, bullocks, various birds and of course humans activities. Thereby enriching these soil environments with keratinic materials. Our study highlights the distribution pattern of keratinophillic fungi as they are important bio

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indicators of environmental pollution with keratin remnants. Both dermatophytes and nondermatophytes being potential pathogens, pose a threat to all those who are regularly in contact with such soil habitats. Special hygiene awareness programme and implementation of precautions will ensure safety of these individuals.

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REFERENCES

- 1. Kunert J.Physiology of keratinophilic fungi. In: Biology of dermatophytes and other keratinophilic fungi.Revista Iberoamericana de *Micologia: Bilbao*, Spain. 2000; 77-85.
- 2. Gugnan CH .Nondermatophytic filamentous keratinophilic fungi and their role in human infection. In: Biology of dermatophytes and other keratinophilic fungi Revista *Iberoamericana de Micología:Bilbao*, Spain. 2000; 109-114.
- Sharma M., Sharma M.Incidence of dermatophytes and other keratinophilic fungi in the schools and college playground soils of Jaipur,India. *Afr J Microbiol Research.*, 2010; 4: 2647-2654.
- Dominik T., Majchrowicz I. A trial for isolating keratinolytic and keratinophilic fungi from the soils of the cemeteries and forests of Szczecin. *Ekologia Polska-Seria.*, 1964; **12**: 79-105.
- Ramesh V. M., Hilda A. Incidence of keratinophilic fungi in the soil of primary schools and public parks of Madras City, India.*Mycopathologia.*,1998-1999; 143: 139– 145.
- Singh I., Kushwaha R.K.S. Dermatophytes, related Keratinophilic in soils of Parks and Agricultural fields of U.P, India. *Ind J Dermatol.*, 2010; 55: 306-308.
- Katiyar S., Kushwaha R.K.S. Human hair colonizing fungi in water sediments of India. *Mycopathologia.*,2001; 152: 81-84.
- Avasn M.Y., Kaizar Hossain D., Hari Priya ., Tejaswi B . Prevalence of Keratinophilic fungi from Sewage Sludge at Some Wastewater out lets along the coast of Visakhapatnam: A case study. *Adv in Appl Sci Resear.*, 2012; 3: 605-610.

J PURE APPL MICROBIO, 8(2), APRIL 2014.

- 9. Deshmukh S.K., Verekar S.A. The occurrence of dermatophytes and other keratinophilic fungi from the soils of Himachal Pradesh (India). Czech *Mycol.*, 2006; **58**: 117–124.
- Deshmukh S.K., Verekar S.A. Incidence of keratinophilic fungi from the soils of Vedanthangal Water Bird Sanctuary (India). *Mykosen.*, 2011; 54: 487-490.
- Vanbreuseghem R. Technique biologique pour 1' isolement des dermatophytes du sol. Ann Soc Belge Med Trop., 1952; 32:173-8.
- Sigler L., Flis A.L., Carmichael J.W. The genus Uncinocarpus (Onygenaceae) and its synonym Brunneospora: new concepts, combinations and connections to anamorphs in *Chrysosporium*, and further evidence of relationship with Coccidioides immitis. *Canad J Bot.*, 1998; **76**:1624-1636.
- Van Oorschot C.A.N.S. A revision of Chrysosporium and allied genera. *Stud Mycol.*, 1980; 20: 1-89.
- Currah R.S. Taxonomy of Onygenales:Arthrodermaceae, Gymnoascaceae, Myxotrichaceae and Onygenaceae. Mycotaxon., 1985; 24:1-216.
- 15. Cano J., Gurrao J. The genus Aphanoascus. Mycological Research.,1990; **94**:355-77.
- Rippon J.W.(ed): Medical Mycology. The pathogenic fungi and the pathogenic Actinomycetes. Philadelphia WB Saunders Company., 1982; Pp 38.
- Garg A.P., Gandotra S., Mukerji K.G., Pugh G.J.E .Ecology of keratinophilic fungi. Proc. Indian Acad. Sci., 1985; 94:149-163.
- Shrivastava J.N., Satsangi G.P., Ajay Kumar. Incidence of keratinophilic fungi in waterlogged condition of paddy soil. J Environ Biol., 2008; 29:125-126.
- Deshmukh S.K., Verekar S.A. Prevalence of keratinophilic fungi in public park soils of Mumbai, India. Microbiol Research., 2012; 3:e6; 24-27
- Singh I.A. ,Mishra., Kushwaha R.K.S. Dermatophytes ,Related Keratinophilic and opportunistic fungi in indoor dust of house and hospitals. *Indian J Med Microbiol.*, 2009; 27: 242-246.
- 21. Saxena P., Kumar A., Shrivastava J.N. Diversity of keratinophilic mycoflora in the soil of Agra (India). Folia Microbiol.,2004; 49:430-4.
- 22. Roilides E., Sigler L., Bibashi E ., Katsifa H., Flaris N., Panteliadis C. Disseminated infection due to *Chrysosporium zonatum* in a patient with Chronic granulomatous disease and review of non-*Aspergillus* fungal infection in patients with this disease. J Clin

Microbiol., 1999; 37:18-25.

- Gan G.G., Kamarulzaman A., Goh K. Y., Ng K.P., Na S.L., Soo-Hoo T.S.Non-Sporulating *Chrysosporium*: An Opportunistic Fungal Infection in a Neutropenic Patient. Med J Malaysia.,2002; 57:118-22
- 24. Lyskova P. Saprotrophic microscopic fungi and dermatophytes accompanying infections of the skin and nails of patients in the Moravian-Silesian Region (Czech Republic). 2007; Czech Mycol.,59:125-37.
- 25. Spiewak R. Zoophilic and geophilic fungi as a cause of skin disease in farmers. Ann Agri Environ Med., 1998; 5: 97-102.
- Saidi S.A., Bhatt S., Richard J.L., Sikdar A.,Ghosh G.R. Chrysosporium tropicum as a probable cause of mycoses of poultry in India.Mycopathologia.,1994; 125:143-147.
- Ganaie M.A., Sood S., Rizvi G., Khan T.A. Isolation and Identification of Keratinophilic fungi from soil samples in Jhansi City (India).Plant Pathol Journ.,2010; 9:194-197.
- Richardson M., Lass- Florl C: Changing epidemiology of systemic fungal infections *Clin Microbiol Infect.*, 2008; 14:5-24.
- 29. Munoz P., Guinea J., Bouza E: Treatment options in emerging mold infections. *Curr Infect Dis Rep.*, 2008; **10**:473-479
- Nucci M., Anaissie E. Fusarium infections in immunocompromised patients. Clin Microbiol Rev., 2007; 20:695-704.
- Doczi I., Gyetvai T., Kredics L., Nagy E Involvements of Fusarium spp.in fungal keratitis.Clin Microbiol Infec.,2004;10:773-6
- Gupta A.K., Baran R., Summerbell R.C. Fusarium Infections of the skin. Curr Opin Infect Dis., 2000; 13:121-128
- Zmeili O.S., Soubani A.O. Pulmonary aspergillosis: a clinical update. Q J Med .,2007; 100:317–334
- Allo M.D., Miller J., Townsend T., Tan C. Primary cutaneous aspergillosis associated with Hickman intravenous catheters. N Engl J Med.,

1987; **317**: 1105–1108

- Denning D.W. Invasive aspergillosis. Clin Infect Dis., 1998; 26:781-803.
- Suzana Tasic., Natasa M.T. Cladosporium Spp. -cause of opportunistic mycoses. *Acta Fac Med Naiss.*, 2007; 24(1):15-19
- Havlickova B., Czaika V. A., Friedrich. Epidemiological trends in skin mycoses worldwide. *M Mycoses*. 2008; 51: 2-15
- Seebacher C., Bouchara J.P., Mignon B. Updates on the epidemiology of dermatophyte infections. *Mycopathologia.*, 2008; 166: 335–352.
- 39. Ameen. Epidemiology of superficial fungal infections. *Clin Dermatol.*, 2010; **28**, 197–201.
- Ali-Shtayeh M.S., Jamous R.M.F. Keratinophilic fungi and related dermatophytes in polluted soil and water habitats. In: Biology of dermatophytes and other keratinophilic fungi Eds, Kushawaha RKS, Guarro J, *Revista Iberoamericana de Micologia*. 2000; Spain, pp. 51-59.
- 41. Nardoni S., Mugnaini L., Papini R., Fiaschi M., Mancianti F. Canine and feline dermatophytosis due to Microsporum gypseum: a retrospective study of clinical data and therapy outcome with griseofulvin. J Mycol Med., 2013; 23:164-7.
- 42. Gungnani H.C., Sharma S., Gupta B.Keratinophilic fungi recovered from feathers of different species of birds in St Kitts and Nevis. *West Indian Med J.*, 2012; **61**: 912-915.
- Ndako J.A., Osemwegie O.O., Spencer T. H., Olopade B.K., Yunusa G.A., Banda. Prevalence of Dermatophytes and other associated Fungi among school children. *Global Adv Research J Med and Medical Sci.*, 2012; 1: 049-056.
- Nweze., Emeka Innocent. Dermatophytosis among children of Fulani/Hausa herdsmen living in southeastern Nigeria. *Rev Iberoam Micol.*, 2010; 27: 191-4.
- 45. Spiewak R., Szostak W. Zoophilic and geophilic dermatophytes among farmers and non-farmers in eastern Poland. *Ann Agr Environ Med.*, 2000; 7:125-129.