Diversity and Antimicrobial Activity of Endophytic Fungi from Traditional Medicinal Plant *Curcuma wenyujin*

Jvfen Yan, Ningbo Qi, Suping Wang and Shulin Yang¹

School of Environmental and Biological Engineering, Nanjing University of Science and Technology, Nanjing - 210094, China.

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Endophytic fungi are ubiquitous in plants and reside their hosts for a part of their life cycle. Their various diversity associated with the potential bioactivity has played an important role in human health, especially that from Chinese traditional medicinal herbs. Here, we isolated endophytic fungi from plant *Curcuma wenyujin* and evaluated their antimicrobial activity. Sixty-six endophytic fungi were recovered from the fresh and healthy root, leaf and stem tissues of *C. wenyujin*. Of these isolates, the common species *Penicillium*, *Fusarium* and *Aspergillus* were the dominant species. 65% of endophytic fungi had the potential antibacterial activity against one or more tested microorganism of *E. coli*, *P. vulgaris*, *B. subtilis*, *S. aureus*, *Rhizopus*, *Microzyme*, *A. niger* and *Mucor*, and 4.5% strains could inhibit eight tested microbes, exhibiting a broad antimicrobial spectrum. These findings provide a strong platform for the isolation and purification of natural antimicrobial agents from endophytic fungi of *C. wenyujin*.

Key words: bioactivity, isolation, distribution, Chinese traditional herb.

The endophytic fungi are ubiquitous in the plant tissues by transmission via air or watersplash from surrounding saprobic colonies ^{1,2}. Its common definition is living at least part of their lifecycle inside plant tissues without causing any visible manifestation of disease to the host plant³. Although endophytic fungi are common in the herb plants, the composition of endophytic communities varies from the ecological and physiological factors in plants 4-7, such as geographical location, age and characteristics of host plant, seasonal changes (temperature, humidity, water content in the soil, etc.). These endophytic fungal interactions with their hosts range from mutualistic to antagonistic⁸⁻¹¹, providing resistance to hosts against different biotic and abiotic stresses and playing an important role in plant community health ^{12,13}. In turn, the host plants

 * To whom all correspondence should be addressed. Tel: +86-25-84315945;

E-mail: bioshuliny@yahoo.com.cn

make the fungal communities increase the absorption of soil nutrients and also change in nutrient cycle occurs¹⁴.

Furthermore, endophytic fungi are the origins of active secondary metabolites ¹⁵ and have been recognized as a repository of novel metabolites of pharmaceutical importance because of their unusual environments ^{16,17}, which may directly or indirectly be used as drugs against many diseases ¹⁸⁻²². Occasionally, fungi associated with medicinal plant are able to produce the same bioactive compounds as the host plant itself or valued metabolites ²³⁻²⁶. Therefore, exploiting endophytic fungi for the production of bioactive secondary metabolites is a challenging and rewarding work.

On the basis of these successful experiments and the theory of "co-evolution" and "gene horizontal transfer" ²⁷, the traditional Chinese medicinal plant *Curcuma wenyujin* was taken as target for the study of its endophytic fungi. *C. wenyujin* is famous for the volatile components including curcumol, β -elemene,

curzerene, curzerenone, curcumenol, etc. that is anticancer activity and used for therapeutical drugs²⁸.

The foregoing contents suggest fungi associated with plant especially the medicinal herb plant are important both from an ecological perspective and from a biochemical and molecular standpoint. So, the aim of this paper is to isolate the endophytic fungi from the herb *C. wenyujin*, study the characteristics of fungal communities and evaluate their potential antimicrobial ability.

MATERIALS AND METHODS

Endophytic fungi isolation and identification

The fresh healthy plant C. wenyujin (C. wenyujin Y.H.Chen & C.Ling) collected from Zhejiang Province, China was carefully washed under running water to discard the surface debris. Then following the procedure of surfacesterilization method ²⁹, the samples were steeped successively in 75% ethanol (v/v) for 1 min, 0.1% mercuric chloride (v/v) for 1 min to roots (5 mm³), 30 s for stems (5 mm) and leaves (5×3 mm), finally washed with sterile distilled water for 1 min several times. These sterilized segments were placed in Petri dishes containing potato dextrose agar (PDA) medium with 50 ig/ml streptomycin, incubated at 28 ! with darkness. Meanwhile, both the last washing water and the print of sterilized tissue segments were tested on the respective PDA as negative control to check the effective of sterilization. After several days, the colonies appeared around the segments were then subcultured in fresh PDA medium on the premise that no microbe grew on the control medium. The pure endophytic fungal strains were then preserved in the laboratory of Institute of Biological Engineering, Nanjing University of Science and Technology and identified according to their morphology based on the manual ³⁰. **Antimicrobial activity assay**

These isolated endophytic fungi were evaluated for their antibacterial activity against eight tested strains including bacterial of *Escherichia coli*, *Proteus vulgaris*, *Bacillus subtilis*, *Staphylococcus aureus* and fungi of *Rhizopus*, *Microzyme*, *Aspergillus niger* and *Mucor* using the agar diffusion method ³¹. The endophytes were inoculated into a 250 ml conical flask containing 100 ml of potato dextrose broth (PDB) and cultured at 28 !, 180 rpm for 10 days. Acquired fermentation liquid was centrifuged to remove mycelium. The supernatant liquid was then condensed for antimicrobial assay. The inhibition

RESULTS AND DISCUSSION

zones were observed, measured and recorded.

During the process of isolation, there was no microbe occurred on the control medium plates, excluding the possibility of surface fungi and suggesting the isolated fungi was endophytic. On this premise, sixty-six isolates endophytic fungi

Family	Species	Number	Root	Stem	Leaf
Moniliaceae	Penicillium sp.	14	10	3	1
	Aspergillus sp.	8	3	3	2
	<i>Ceohalosporium</i> sp.	2	1	1	
	Staphlosporonites sp.	1	1		
	Mycogone sp.	3	2	1	
Dematiaceae	Cladosporium sp.	4	2	1	1
	Alternaria sp.	2	2		
	Monotospora sp.	3	2	1	
Tuberculariaceae	Fusarium sp.	13	9	3	1
Saccharomycetaceae	Saccharomyces sp.	1	1		
Mucoraceae	Mucor sp.	4	2	1	1
Sphaeropsidaceae	Hendersonula sp.	2			2
Agonomycetaceae	Rhizoctonia sp.	5	3		2
	Mycelia sterilia	4	2		2
		66	40	14	12

Table 1. The endophytic fungi isolated from C. wenyujin

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were recovered from the healthy, symptomless leaves, stems and roots of C. wenyujin followed by the surface sterilization method. Among, fortythree strains isolated from the roots belonging to thirteen genera, fourteen strains from stems under eight ranks and twelve from leaves (Table 1). Obviously, Penicillium (21.2%), Fusarium (19.7%) and Aspergillus (12.1%) were the dominant species. This result was similar to that of Xuan and Zhang ³² and Wang *et al.* ³³ who also isolate endophytic fungi from Cucurma. They are common a genus of ascomycetous fungi of major importance in the natural environment, but can also occur as endophytes. The common species Penicillium sp., Aspergillus sp., Cladosporium sp., Fusarium sp. and Mucor sp. were found in all the tissues (Table 1), but Staphlosporonites, Alternaria and Saccharomyces were occurred in the roots and Hendersonula only from leaves, which illustrated that on the one hand, a flora of diversity endophytic fungi resided in the medicinal plant C. wenyujin and on the other hand there were some differences in the quantity and composition of endophytic fungi associated with different tissues.



Fig 1. The summary of active endophytic fungi from *C*. *wenyujin*. X axis stands for the number of tested microbes which can be inhibited

a strong platform for the isolation and purification of natural antimicrobial agents from endophytic fungi of *Curcuma*.

REFERENCES

1. Bills, G.F.: Isolation and analysis of endophytic fungal communities from woody plants. In: *Endophytic fungi in grasses and woody plants*

Endophytic fungi have been described as mutualists that protect host plants against insect herbivory through producing biologically active secondary metabolites ³⁴. Huang et al. ³⁵ isolated 172 endophytic fungi from three medicinal plants and tested their fermentation broths for cytotoxicity, showing that the percentage of the broths at a dilution at 1:50 displayed a cytotoxic activity of 50% growth inhibition. Moreover, Phongpaichit et al., ³⁶ obtained 65 crude extracts from 51 endophytic fungi isolated from Garcinia plants and assessed these extracts for various bioactivities. In this research, 66 endophytic fungi isolated from C. wenyujin were tested for antibacterial activity by Oxford cup diffusion assay against eight isolates. Of these endophytes, 65% of endophytic fungi showed antibacterial activity against one or more than one microorganism, and 4.5% strains could inhibit eight tested microbes, exhibiting a broad antimicrobial spectrum (Fig 1). These active isolates mainly recovered from the root of C. wenyujin and distributed in the species of Penicillium (18.2%), Fusarium (12.1%) and Aspergillus (9.1%) (Fig 2). These findings provide

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Fig. 2. The distribution of active entophytic fungi in *C. wenyujin*

(Redlin SC, Carris LM, eds). St Paul, Minnesota: APS Press, 1996; pp 31-65.

- Wilson, D. Ecology of woody plant endophytes. In: *Microbial Endophytes* (Bacon CW, White JF, eds). New York & Basel: Marcel Dekker, 2000; pp 389-420.
- Bacon, C.W., White, J.F. (eds): Microbial Endophytes. New York: Marcel Deker Inc., 2000.
- 4. Khan, R., Shahzad, S., Choudhary, M.I., et al.

J PURE APPL MICROBIO, 8(2), APRIL 2014.

Communities of endophytic fungi in medicinal plant *Withania somnifera*. *Pak. J. Bot.*, 2010; **42**: 1281-1287.

- Mocali, S., Bertelli, E., Cello, F.D., *et al.* Fluctuation of bacteria isolated from elm tissues during different seasons and from different plant organs. *Res. Microbiol.*, 2003; **154**: 105-114.
- 6. Araujo, W.L., Marcon, J., Maccheroni, Jr.W., *et al.* Diversity of endophytic bacterial populations and their interaction with *Xylella fastidiosain* Citrus plants. *Appl. Environ. Microb.*, 2002; **68**: 4906-4914.
- Clay, K., Holah, J. Fungal endophyte symbiosis and plant diversity in successional fields. *Science*, 1999; 285: 1742-1744.
- Saikkonen, K., Faeth, S.H., Helander, M., *et al.* Fungal endophytes: a continuum of interactions with host plants. *Annu. Rev. Ecol. Syst.*, 1998; 29: 319-343.
- Ganley, R.J., Brunsfeld, S.J., Newcombe, G. A community of unknown, endophytic fungi in western white pine. *Proc. Natl. Acad. Sci. USA*, 2004; **101**: 10107-10112.
- 10. Schulz, B., Boyle, C. The endophytic continuum. *Mycol. Res.*, 2005; 109: 661-686.
- 11. Hatcher, M.J., Dick, J.T.A., Dunn, A.M. How parasites affect interactions between competitors and predators. *Ecol. Lett.*, 2006; **9**: 1253-1271.
- Kharwar, R.N., Verma, V.C., Strobel, G., et al. The endophytic fungal complex of *Catharanthus* roseus (L.) G. Don. Curr. Sci., 2008; 95: 228-233.
- Gond, S.K., Verma, V.C., Mishra, A., et al.: Role of fungal endophytes in plant protection. In: *Management of Fungal plant pathogens* (Arya A, Perello'AE, ed). Wallingford: CAB International, 2010; pp 183-197.
- Krishnamurthy, Y.L., Naik, S.B., Jayaram, S. Fungal communities in herbaceous medicinal plants from the Malnad Region, Southern India. *Microbes Environ.*, 2008; 23: 24-28.
- Kusari, S., Hertweck, C., Spiteller, M. Chemical ecology of endophytic fungi: origins of secondary metabolites. *Chem. Biol.*, 2012; 19: 792-798.
- Verma, V.C., Kharwar, R.N., Strobel, G.A. Chemical and functional diversity of natural products from plant associated endophytic fungi. *Nat. Prod. Commun.*, 2009; 4: 1511-1532.
- Tan, R.X., Zou, W.X. Endophytes: a rich source of functional metabolites. *Nat. Prod. Rep.*, 2001; 18: 448-459.
- Strobel, G.A., Daisy, B., Castillo, U., et al. Natural products from endophytic microorganisms. J. Nat. Prod., 2004; 67: 257-

J PURE APPL MICROBIO, 8(2), APRIL 2014.

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- Staniek, A., Woerdenbag, H.J., Kayser, O. Endophytes: exploiting biodiversity for the improvement of natural product-based drug discovery. J. Plant. Interact., 2008; 3: 75-93.
- 20. Aly, A.H., Debbab, A., Kjer, J., *et al.* Fungal endophytes from higher plants: a prolific source of phytochemicals and other bioactive natural products. *Fungal Divers.*, 2010; **41**: 1-16.
- Kharwar, R.N., Mishra, A., Gond, S.K., *et al.* Anticancer compounds derived from fungal endophytes: their importance and future challenges. *Nat. Prod. Rep.*, 2011; 28: 1208-1228.
- Kusari, S., Spiteller, M.: Metabolomics of endophytic fungi producing associated plant secondary metabolites: progress, challenges and opportunities. In: *Metabolomics* (Roessner U, ed). InTech, 2012; pp 241-266.
- Stierle, A., Strobel, G.A., Stierle, D. Taxol and taxane production by *Taxomyces andreanae*, an endophytic fungus of Pacific yew. *Science*, 1993; 260: 214-216.
- Kusari, S., Lamsho"ft, M., Spiteller, M. *Aspergillus fumigatus Fresenius*, an endophytic fungus from *Juniperus communis* L. Horstmann as a novel source of the anticancer pro-drug deoxypodophyllotoxin. *J. Appl. Microbiol.*, 2009a; **107**: 1019-1030.
- 25. Kusari, S., Zu"hlke, S., Spiteller, M. An endophytic fungus from *Camptotheca acuminatathat* produces camptothecin and analogues. J. Nat. Prod., 2009b; **72**: 2-7.
- Kusari, S., Verma, V.C., Lamsho"ft, M., et al. An endophytic fungus from Azadirachta indica A. Juss. that produces azadirachtin. World J. Microbiol. Biotechnol., 2012; 28: 1287-1294.
- 27. Nisbet, L.J., Fox, F.M. The importance of microbiodiversity to biotechnology. In: *The Biodiversity of microorganisms and invertebrates; its role in sustainable agriculture* (Hawksworth DL, ed). Wallingford: CAB International, 1991; pp 229-244.
- 28. Mau, J.L., Lai, E.Y.C., Wang, N.P., *et al.* Composition and antioxidant activity of the essential oil from *Curcuma zedoaria*. *Food Chem.*, 2003; **82**: 583-591.
- 29. Cankar, K., Kraigher, H., Ravnikar, M., *et al.* Bacterial endophytes from seeds of *Norway spruce* (Picea abies L. Karst). *FEMS Microbiol. Lett.*, 2005; **244**: 341-345.
- Domsch, K.H., Gams, W., Anderson, T.H. (eds): Compendium of soil fungi, 2nd edn. New York: Academic Press, 2007.
- 31. Bauer, A.W., Kirby, W.M., Sherries, J.C., *et al.* Antibiotics susceptibility testing by the standardized single disc method. *Am. J. Clin.*

Pathol., 1966; 45: 493-496.

- 32 Xuan, Q., Zhang, L.Q. The isolation and identification of endophytic fungi from *Curcuma. Journal of Chinese National Folk Medicine*, 2007; **84**: 45-46.
- Wang, Y.H., Xu, L., Yang, X.D., *et al.* Isolation and antibacterial activities of endophytic fungi from *Curcuma aromatica* Salisb. *Journal of Chinese Pharmacy*, 2009; 44: 972-975.
- Stone, J.K., Bacon, C.W., White, J.F. An overview of endophytic microbes: endophytism defined. In: *Microbial endophytes* (Bacon CW,

White JF, eds). New York: Marcel Dekker, 2000; pp 3-29.

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- Huang, Y.J., Wang, J.F., Li, G.L., *et al.* Antitumor and antifungal activities in endophytic fungi isolated from pharmaceutical plants. *FEMS Immunol. Med. Mic.*, 2001; **31**: 163-167.
- Phongpaichit, S., Nikom, J., Rungjindamai, N., et al. Biological activities of extracts from endophytic fungi isolated from *Garcinia* plants. *FEMS Immunol. Med. Microbiol.*, 2007; **51**: 517-525.