

Characterization of *Alternaria* Isolate from Organically-Grown Tomato (*Lycopersicon esculentum* L.) in Relation to the Fruit Antioxidant Compounds

Afrah E. Mohammed, Eman A. H. Alhomaidi and Dalal H. M. Alkhalifa

Department of Biology, Faculty of Science, Princess Nourah Bint Abdulrahman University,
Postal Code 11474, Riyadh, Saudi Arabia.

(Received: 03 September 2014; accepted: 12 October 2014)

Environmental pollution as a consequence of irrational usage of fungicides is an immense problem bothering the well-being of human kind globally. The real risk is the possibility of such pollutants to ultimately find their way to human food via food chain. Recently, a growing concern regarding food safety has increased in response to the high demand for organic products. Organic tomato cultivation gained great preference and acceptability for consumers. However, contamination of fresh fruits and vegetables with a variety of postharvest fungi is an essential problem which requires immediate measures to be taken seriously for the sake of maintaining human health. The main objective of this work is to study the susceptibility of organically-cultivated tomato fruit to post harvest pathogenic fungi and to find out whether there is relationship between total phenolic compounds, ascorbic acid content and antioxidant capacity of tomato fruit and the appearance of fungal growth. Furthermore, DNA-based method, Random amplified polymorphic DNA technique (RAPD) was used to fingerprint and assess the genetic information about the fungal isolate. The current results revealed that different tomato varieties showed different susceptibility levels to infection by postharvest pathogenic fungi besides different amounts of accumulated total phenolic compounds, ascorbic acid content and antioxidant capacity. Furthermore, higher concentrations of total phenolic compounds, ascorbic acid content and antioxidant capacity were detected in the small size tomato fruit, which might point to absence of fungal growth. Furthermore, morphological characteristics showed that the isolate detected from infected tomato fruit was *alternaria alternata*, finger print (RAPD) technique showed clear DNA bands for the isolate which indicated that the opa04 was an efficient primer that supported the amplification by producing unique patterns of banding for the *A. alternata* isolate. Further molecular work is needed to provide more authenticated information regarding the isolate.

Key words: *Lycopersicon esculentum*, antioxidants, *alternaria alternata*, RAPD

Tomato is one of the most widely grown vegetables in the world. Tomato fruit production has been mainly dependent on fungicides usage, but growing concerns about food safety and environmental pollution have increased the demand of organic products all over the world. Organic cultivation of tomato is one of the factors that are becoming major decision-making in tomato

* To whom all correspondence should be addressed.

preferences for many consumers¹. The total area of organically certified land in Saudi Arabia is expanding fast as more farmers are converting to organic production due to increase in consumer awareness. The most important organic crops in Saudi Arabia are vegetables with a cultivation area of about 8.373 hectare². Tomato is an excellent source of many secondary metabolites important for human health such as folate, ascorbic acid, flavonoids, chlorophyll, β -carotene and lycopene³. Phenolic compounds are synthesized by plants during normal development and also in response

to stress conditions. Generally, phenolics are characterized by a benzene ring and hydroxyl group which can be converted into lignin. Light enhances the biosynthesis of phenolic substances in chloroplast, but they tend to accumulate in vacuoles in relatively high amounts or deposit in secondary cell wall as lignin⁴. The interest in polyphenols as antioxidants is focused on flavonoids⁵. Flavonoids can interfere with several free radical-generating systems and their reactive hydroxyl groups via oxidation to more stable and less reactive radical compounds³. Recent studies reported that the regular consumption of tomato has been associated with decreased risk of chronic degenerative diseases due to the presence of antioxidant compounds such as carotenoids, particularly ascorbic acid and phenolics, particularly flavonoids as well as higher levels of potassium since it is an important factor in lowering blood pressure³⁻⁶. A comparative study of vegetables produced in organic farming showed predominately lower susceptible to pests and diseases and were characterized by higher contents of defense-related secondary metabolites⁷. Furthermore, Losses due to postharvest disease that leads to reductions in fruit quantity and quality may occur any time during postharvest handling, from harvest to consumption. *Alternaria alternata* is a common saprobe opportunistic pathogen affecting many cultivated plants in the field and during post-harvest storage of fruit and vegetables⁸⁻⁹. The pathogenicity factor of *A. alternata* with respect to tomato plant infection lies in the production of host-specific AAL-toxin that is capable of inducing cell death¹⁰. To date, little is known about the direct interaction of infection of tomato fruit by *A. alternata* and certain fruit compounds accumulation; nevertheless, plant might react with pathogenic attack by accumulation of compounds that increase plant resistance to infection. Soluble sugars, especially sucrose, glucose and fructose can be increased in tomato fruit due to infection pressure in order to restrict the accumulation of reactive oxygen species¹¹. On the other hand, the response of phenolic compounds to pathogenic attack is based mainly on defense response that is characterized by the early and rapid accumulation of phenolics at the infection site¹². Moreover, ascorbic acid contributes on the response to pathogenic attack

through the control of biosynthesis of hydroxyproline-containing protein, which is involved in the cross-linking of the cell wall in response to injury and extension genes¹³. Furthermore, several methods have been used to illustrate the variation among species of parasitic fungi such as morphology of spores and colonies, extracellular protein profiles, pathogenicity and growth requirements¹⁴. Nevertheless, variation among fungal strains and species using Random Amplified Polymorphic DNA (RAPD) which is described by Williams *et al.*¹⁵ is an easy tool to detect polymorphism in a large number of samples at relatively low cost. It is a modification of PCR (polymerase chain reaction) that allows revealing polymorphism within completely unknown samples without the need of probe hybridization or DNA sequencing¹⁶. The present study aimed to gain knowledge about antioxidant compounds accumulation in different tomato cultivars cultivated under organic farming conditions in relation to infection by post-harvest pathogen. Besides, a PCR-based DNA marker was used for pathogenic microorganism characterization.

MATERIALS AND METHODS

Plant Materials

Two different varieties of tomato fruit were obtained from an organic farm in Diyrab, Riyadh, KSA on the same harvest day in January 2013. One variety with small size (weight 5.5g), cherry and other one with a big size (weight 41.1 g). Some of the collected samples were stored at "50°C for further analysis. The rest of the samples were kept at room temperature (25° C) in the department of biology laboratory and observations were recorded every two days regarding appearance of any fungal growth. For the fruit quality measurements, frozen samples were freeze-dried (Epsilon2-40, Christ, Germany) for four days, then the samples were milled in a speedy mixer (KRUPS, Speedy Pro Plus F7247010F, France). Afterwards, samples were milled to a fine powder up to a particle size of 0.2 µm for further analysis.

Determination of ascorbic acid, total phenolic compounds and antioxidant capacity

Ascorbic acid was measured by titration from fresh fruits according to Albrecht¹⁷. The total phenolic compounds content was determined with

Folin-Ciocalteu's phenol reagent from the freeze-dried material by spectrophotometer according to Singleton and Rossi¹⁸ and the antioxidant capacity was determined using the Ferric Reducing Antioxidant Potential (FRAP) assay which works with iron for oxidation according to Binoy et al¹⁹. Three replicates for each sample were performed regarding the laboratory analysis.

Detection of fungal growth

Tomato samples with symptoms of fungal growth were collected and taken to the microbiology lab for further investigations. The isolation of the pathogen was carried out by cutting the pieces, at the edge of contaminated and healthy tissue²⁰. Thereafter, fragments were transferred on potato dextrose agar (PDA) and incubated at 25°C for three days. Growth characteristics of cultures were studied on PDA.

Fungal isolation and DNA extraction:

The selected isolate was grown on potato dextrose (PD) medium and allowed to grow for 7 days at 28°C. After the incubation, the mycelium was scraped off for DNA extraction. Total DNA was extracted by DNeasy Plant Mini Kit (QIAGEN)²¹.

Characterization of isolate by RAPD:

RAPD analysis was carried according to Gherbawy²² and Parak *et al.*²³, 25µl PCR reaction, containing 1µl of template DNA, 12.5µl KAPA Taq ready mix, 10.5µl water and 1µl of primer OPA04 (5'-AAT CCG GCT G-3').

The amplification was carried out as follows:

The thermal cycling program was as follows: 4 minutes initial denaturation at 95 C, followed by 45 cycles of 9 seconds denaturation at 95 C, 25 seconds 40 C, 90 seconds with final extension at 72 C for 4 minutes. A negative control using water instead of template DNA was included

in the amplification process. DNA fragments were separated by electro-phoresis in 1.5% (w/v) agarose gel containing ethidium bromide (0.5 g/ml) using 1×TBE buffer. Visualization was performed in UV transilluminator and the images were captured with DOC PRINT system (VilberLour-mat, USA). The lengths of the amplification products were estimated by comparison with a 100 bp DNA ladder.

RESULTS AND DISCUSSION

Tomato is an excellent source of many secondary metabolites³. The present study revealed significant variations in the content of fruit antioxidant compounds between the investigated varieties. The individual compounds and their relationship to fruit weight were presented in table 1. Total phenolic compounds, ascorbic acid content and antioxidant capacity were detected in both varieties which had the same trend compatible with^{19,24} who studied different tomato varieties in conventional and organic farming system. Variations between studied varieties were observed in the content of the total phenolic compounds, ascorbic acid and antioxidant capacity; many different studies showed that the secondary metabolites of tomato fruit differ according to different varieties²⁴. Our results indicated that, cherry tomato had the higher content of total phenolic compound (894.1 ± 8.3), ascorbic acid (325.5 ± 5.2)mg g⁻¹ fresh weight and antioxidant capacity (70.85 ± 1.5)mmol Fe²⁺ kg⁻¹ fresh weight compared with the big size tomato (Table 1). Similar results were also detected by^{19,24,25} who stated that cherry tomatoes with a low fruit weight represent a better source of antioxidants compounds than fruits of a larger size. The antioxidant capacity was

Table 1. Fruit total phenolic compounds, ascorbic acid and antioxidant capacity for two different tomato varieties grown organically at Diyrabfield. Riyadh, Saudi Arabia.

Antioxidant capacity (mmol Fe ²⁺ kg ⁻¹ fresh weight)	Ascorbic acid (mg g ⁻¹ fresh weight)	Total phenolic compounds (mg g ⁻¹ fresh weight)	Fruit size (g)	Genotype
70.85 ± 1.5 a	325.5 ± 5.2 a	894.1 ± 8.3 a	05.5 b	Cherry tomato
53.35 ± 3.3 b	172.4 ± 7.1 b	489.7 ± 9.2 b	41.1 a	Big size tomato

Data are expressed as mean ± SD. Different letters within a column indicate significant differences in the Tukey test; ±5%.

higher in the cherry tomato where the total phenolic compound was higher. This might support the information that the total phenolics are important contributors to the total antioxidant activity²⁶. Measurement of antioxidant capacity in an organic solvent extract, e.g. in acetone or dichloromethane, as in this study, is usually well correlated to carotenoid and lycopene concentrations, as carotenoids are part of the lipophilic extract²⁷. On the other hand, tomato with a big size 31.1 g showed fungal growth 10 days after harvest, but cherry tomato (5.5 g) showed no growth although both tomato varieties were kept in the same area at the same room temperature. Variations among different varieties were also observed by Mohammed *et al.*²⁷ who studied 28 organic tomato cultivars that differ in their susceptibility to fungal infection in the field by *Phytophthora infestans*. Interestingly, the total phenolics compounds and antioxidant capacity had higher concentrations on the cherry tomato which showed no fungal growth compared with a big size one, same result was also described by Jinsin²⁷ who stated that small tomato fruits had higher resistance to disease. Differences in plant susceptibility to disease could be attributed to possible variations in the growth type, physiological and genetic attributes besides differences in the accumulation of plant secondary metabolites²⁸. Early study²⁹ reported that cultivar

resistant to fungi infection expressed much higher levels of phenolics and oxidative enzymes. Further observation³⁰ found that ascorbic acid content was higher in tomato cultivars resistant to tomato yellow leaf curl virus than the non-resistant ones. These observations might explain the absence of the fungal growth in cherry tomato since resistant to fungi infection was related to much higher levels of phenolics and oxidative enzymes which could create an inhibiting environment in and around the infected tissues, comprising very reactive phenylpropanoid free radicals and lignifications process²⁹. Another study revealed that the elevation of the antioxidant capacity of plant is expected to increase their tolerance to pathogens or a biotic stress³¹.

Furthermore, one of the major causes of poor fruit quality during storage and transport are diseases caused by phytopathogenic fungi³². According to the morphological characteristics isolated fungus from tomato fruit in this study was identified, it considered as the hardest step in the detection process³³. *A. alternata* was determined following the growth characteristics on PDA. Initially; the mycelium was hyaline that turned to grey- brownish, multicelled, with cottony texture and white margin. Findings about mycelium colour was in agreement with results of different studies^{34,35,36} who noticed mycelium of *Alternaria spp.*, as grayish to green, brown and deep olive grey in colour in infected watermelon, chilli, gerbera and tomato respectively. Furthermore, *A. alternata* isolated from contaminated tomato fruit was confirmed by comparing the amplified DNA fragments. In the present study, Amplification of genomic DNA of the isolates of endophytic *Alternaria alternata* which was performed Using opa04 primer that produced 8 DNA bands. All the bands were clear and reproducible, and the size ranged from 750-3000 base pairs. No visible fragment was produced below 750 bp or above 3000 bp. Our results indicated that the opa04 was an efficient primer that supported the amplification by producing unique patterns of banding for *A. alternata* isolate.

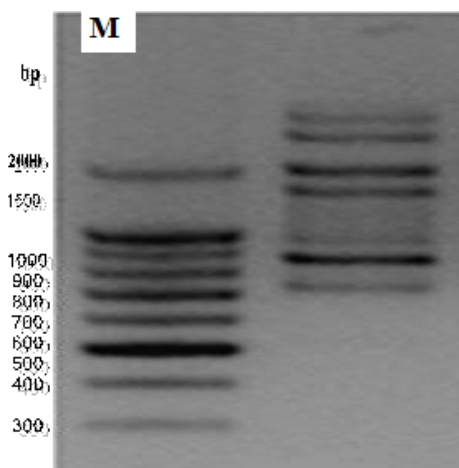


Fig. 1. Random amplified polymorphic DNA patterns obtained by using the primer OPA04 (5'- AAT CGG GCT G-3') from *Alternaria alternata* isolate. The results showed that the tested primer recorded amplification with tested isolate; it amplified 8 DNA bands. 100 bp DNA ladder

CONCLUSION

In light of the present findings it was obvious that the size of tomato fruit is an important

parameter to be considered in fruit selection with regard to fruit quality attributes. More interestingly the size of the fruit was found to be strongly correlated with the degree of susceptibility to fungal growth. Seemingly, it would be quite helpful to investigate the mechanisms underlying the secret between fruit size and susceptibility level. Answers to such enquiries would no doubt enable the most appropriate of tomato varieties for the purpose of breeding selection.

REFERENCES

- Jones, J.B. Tomato plant culture: in the field, greenhouse, and home garden 2nd ed. CRC Press is an imprint of Taylor and Francis Group, USA 2008.
- Hartmann, M., Khalil, S., Bernet, T., Ruhland, F., Al Ghamdi, A. Organic Agriculture in Saudi Arabia 2012. <http://www.giz.de/en/downloads/giz2012-organic-agriculture-saudi-arabia-en.pdf>
- Willcox, J.K., George, L., Catignani, G.L., Lazarus, S. Tomatoes and Cardiovascular Health Critical Reviews. *Journal of Food Science and Nutrition* 2003; **43**: (1) 1–18.
- Kefeli, V.I., Kalevitch, M.V., Borsari, B. Phenolic cycle in plant and environment. *J. cell and mol.bio.* 2003; **2**: 13 – 18.
- Gomez-Romero, M., Arraez-Roman, D., Segura-Carretero, A., Fernandez-Gutierrez, A. Review, Analytical determination of antioxidants in tomato: Typical components of the Mediterranean diet. *J. Separ. Sci.* 2007; **30**: 452 – 46.
- Frusciante, L., Carli, P., Maria, R., Ercolano, M. R., Pernice, R., Matteo, A., Fogliano, V., Pellegrini, N. Antioxidant nutritional quality of tomato. *Mole. Nutr. Food Res.* 2007; **51**: 609 – 617.
- Brandt, K., Molgaard, J.P. Organic agriculture: Does it enhance or reduce the nutritional value of plant foods? *J. the Sci. of Food and Agri.* 2001; **81**: 924 - 931.
- Nasim, G., Khan, S., Khokhar, I. Molecular polymorphism and phylogenetic relationship of some *Alternaria alternata* isolate. *Pak. J. Bot.* 2012; **44**(4): 1267-1270.
- Tokumasu, S., Aoiki, T. A new approach to studying microfungus succession on decaying pine needles in an oceanic subtropical region in Japan. *Fungal Diver.* 2002; **10**: 167-183.
- Yamagishi, D., Akamatsu, H., Otani, H., Kodama, M. Pathological evaluation of host-specific AAL-toxins and fumonisin mycotoxins produced by *Alternaria* and *Fusarium* species. *J Gen Plant Pathol* 2006; **72**: 323-327.
- Rosales, M.A., Rubio-Wilhelmi, M.M., Castellano, R., Castilla, N., Ruiz, J.M., Romero, L. Sucrolytic activities in cherry tomato fruits in relation to temperature and solar radiation. *Science Horticulture* 2007; **113**: 244 – 249.
- Chérif, M., Sadafi, N., Benhamou, N., Hajlaoui, M.R., Boubaker, A., Tirilly, Y. Ultrastructure and cytochemistry of in-vitro interactions of the antagonistic bacteria *Bacillus cereus* X16 and *B. thuringiensis* 55T with *Fusarium roseum* var. *sambucinum*. *J Plant Path* 2002; **84**: 83 – 93 (2002).
- Carpita, N.C., Gibeaut, D.M. Structural models of primary cell walls in flowering plants: Consistency of molecular structure with the physical properties of the cell wall during growth. *Plant Journal* 1993; **3**: 1 – 30.
- Samson, R.A. Identification – Entomopathogenic Deuteromycetes. In: BURGESH.D. (ed.): Microbial Control of Pests and Plants Diseases 1970–1980. Acad. Press, London 1981:93–106
- Williams, J.G.K., Kubelik, A.R., Livak, K.J., Rafalski, J.A., Tingey, S.V. DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. *Nucleic Acid. Res.* 1990; **18** (22), 6531–6535.
- Bielikova, L., Landa, Z., Osborne, L. S., urn V. Characterization and Identification of Entomopathogenic and Mycoparasitic Fungi using RAPD-PCR Technique. *Plant Prot. Science* 2002; **38** (1): 1–12
- Albrecht, J.A. Ascorbic acid and retention in lettuce. *J Food Qual* 1993; **16**: 311 – 316.
- Singleton, V.L., Rossi, J.A. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *Amer J Eno Vitic* 1965; **16**:144 – 158.
- Binoy, G., Charanjit, K., Khurdiyaa, D.S., Kapoorb, H.C. Antioxidants in tomato (*Lycopersium esculentum*) as a function of genotype. *J Food Chem* 2004; **84**: 45 – 51.
- Mathur, S.B., Kongsdal, O. Common laboratory seed health testing methods for detecting fungi. ISTA, Switzerland, 2003.
- Hsin-Hung, C., Wen-Shi, W. Phylogenetic analysis of internal transcribed spacer regions of the genus *Alternaria*, and the significance of filament-broken conidia. *Mycol. Res.*, 2002; **106** (2): 164-169.
- Gherbawy, y. Genetic variation among isolates of *Alternaria spp.* from select egyptian crops. *Archives of Phytopath and plant protect.* 2005; **38**(2):77-89.

23. Parak, M., Romanoski, C., Pryor, B. A re-examination of the phylogentic relationship between the causal agents of carrot black rot, *Alternaria radicina* and *A. caroliincultae*. *Mycologia* 2008; **100** (3):511-527.
24. Mohammed, A. E., Smit, I., Pawelzik, E., Keutgen, A. J., Horneburg, B. Organically grown tomato (*Lycopersicon esculentum* Mill.): bioactive compounds in the fruit and infection with *Phytophthora infestans*. *J. Sci. Food Agric.* (2012). (Wiley online library.com) DOI10.1002/jsfa.4720
25. Lenucci, M.S., Leucci, M.R., Piro, G., Dalessandro, G. Variability in the content of soluble sugars and cell wall polysaccharides in red-ripe cherry and high-pigment tomato cultivars. *J. of Sci Food and Agric.* 2008; **88**: 1837 – 1844.
26. Leonardi, C., Ambrosino, P., Esposito, F., Fogliano, V., Antioxidant activity and caotenoid and tomatine contant in different typologies of fresh consumption tomatoes. *J Agric Food Chem* 2000; **48**: 4723 – 4727.
26. Toor, R.K., Geoffrey, P., Anuschka, H.S. Influence of different types of fertilizers on the major antioxidant components of tomatoes. *J Food Compo Anal* 2006; **19**: 20 – 27.
27. Jinsin, Y., Evaluation of cherry tomato inbred lines for multiple disease resistance. Asian regional center (AVRDC). ARC training 1999.
28. Marjorie, M.C. Plant products as antimicrobial agent. *Clinical Microbiology Reviews* 1999; **12**: 564 – 582.
29. Richard, N.B., Roger, M.W. Secondary metabolites in plant defence mechanisms. *New Phytol* 1994; **127**: 617 – 633.
30. Suarez, H.M., Pallares, J.R., Mesa, D.R., Rodrlguez, I.E.R., Romero, C.D. Variation of the chemical composition of tomato cultivars (*Lycopersicon esculentum* Mill.) according to resistance against the tomato yellow leaf curl virus (TYLCV). *J Sci Food Agric* 2009; **89**: 177 – 194.
31. Balazs, B., Jozef, F., Miklos, P., Zoltan, K. Role of reactive oxygen species and antioxidants in plant disease resistance. *Pest Manag Sci* 2003; **59**: 459 – 464.
32. Grahovac, M., Ini, D., Tanovi, B., Lazi, S., Vukovi, S., Hrusti, J. and Gvozdenac, S. Integrated management of causal agents of postharvest fruit rot of apple. *Pesti. and Phytomed.* 2001; **26** (4): 289-299.
33. Rahjoo, V., Zad, J., Javan-Nikkhah, M., Mirzadi-Gohari, A., Okhovvat, S.M., Bihamta, M.R., Razzaghian, J. and Klemsdal, S.S. Morphological and Molecular identification of *Fusarium* isolated from maize ears in Iran. *Journal of Plant Pathology* 2008; **90** (3): 463-468.
34. Narian, A., Swain, N.C., Sahoo, K.C., Dash, S.K., Shukla, V.D. A new leaf blight and fruit rot of watermelon, *Indian Phytopath.*, 1985; **38**(1): 149-151.
35. Nagrale, D. T., Gaikwad, A. P., Sharma, L. Morphological and cultural characterization of *Alternaria alternata* (Fr.) Keisslerblight of gerbera (*Gerbera jamesonii* H. Bolus ex J.D. Hook) *J. Appli. and Natu. Scie.* 2013; **5** (1): 171-178.
36. Prasad, V., Upadhyay, R. S. *Alternaria alternata* f. sp. lycopersici and its toxin trigger production of H₂O₂ and ethylene intomato. *J. Plant Patho.* 2010; **92**: 103-108.