Detoxification of Carcinogenic Cr (VI) by Combined Action of Bacillus pumilus-S4 and Pseudomonas doudoroffii-S5 in Associated with Hydrophytes

Muhammad Faisal

Department of Microbiology and Molecular Genetics, University of the Punjab, Quaid-e-Azam Campus, Lahore-54590, Pakistan.

(Received: 26 June 2014; accepted: 21 August 2014)

Two chromium resistant bacterial strains (*Bacillus pumilus*-S4 and *Pseudomonas doudoroffii*-S5) isolated from tanneries effluent of were reported in this study. Strains resist up to 20 mg K₂CrO₄ ml⁻¹ on nutrient agar and 2.2 mg ml⁻¹ in acetate minimal medium. Chromate uptake by heat killed cells of *Bacillus pumilus*-S4 was (5.3 mg g⁻¹ dry wt) 90 min as compared to living cells (25.4 mg g⁻¹ dry wt). *Pseudomonas doudoroffii*-S5 reduced 14.2 mg Cr (VI) 36 h⁻¹ at an initial concentration of 200 μ g K₂CrO₄ ml⁻¹. Both strains also release extra cellular proteins in the medium which results extra cellular Cr (VI) detoxification. Cr (VI) removal efficiency of these strains was accelerated by *Pistia stratiotes* and *Eichornia crassipes* which provide energy source to strains. Hence these strains could be utilized for the treatment of chromium polluted industrial wastewater in association with metal hyperaccumulator hydrophytes.

Key words: Bacillus pumilus; Chromium; Eichornia crassipes; heavy metals; Pistia stratiotes; Pseudomonas doudoroffii.

Industrialization is although a premium for modern development, yet its effects on surroundings are equally hazardous¹. Major waste water effluents such as antibiotics, drugs and heavy metals are carcinogenic, mutagenic and teratogenic². Exposure or ingestion of contaminated food and water with heavy metals leads to metal intoxication³. Chromium is most toxic heavy metal that is generated from electroplating, leather tanning and in air craft industry⁴. Chromium occurs in many oxidation forms but the two important from are trivalent Cr (III) and hexavalent Cr (VI) chromium. Trivalent chromium is essential micronutrient in animals for proper glucose utilization⁵. Hexavalent chromium is mutagenic⁶ and carcinogenic⁷. Conventional methods for treatment of toxic chromate include chemical reduction followed by precipitation, ion exchange, and adsorption on activated kaolinite, alum and ash⁸. Chemical detoxification of chromium is more expensive and poisonous for the environment. Biological methods of chromium removal are less expensive are environment friendly⁹. Detoxification of chromium with algae¹⁰, fungi¹¹ plants¹² is well established. Many genera of bacteria have been reported to reduce Cr (VI) under either aerobic¹³ or anaerobic¹⁴ conditions. Beside this hydrophytes also play a significant role in metal removal via filtration, cation exchange, adsorption and through plant-induced chemical alteration in the root zone¹⁵. The free floating hydrophytes can be a better alternative technology for the extraction and removal of toxic chromium from contaminated waste water¹⁶. The present study focus on the

^{*} To whom all correspondence should be addressed. Tel: 92-42-35952811; Fax: 92-42-99230481; Email: mohdfaysal@yahoo.com

utilization of chromium resistant bacteria and hydrophytes in combination for the removal of toxic chromium.

MATERIALS AND METHODS

Strains characterizations

Polluted samples were used for the isolation of chromium resistant bacteria. Strains were maintained at 4°C on nutrient agar plates supplemented with 1000 µg ml⁻¹ of K₂CrO₄. Temperature and pH were noted at the spot. Both these strains were characterized morphologically, biochemically, physiologically and genetically following Gerhardt et al¹⁷.

Collections of hydrophytes

Free floating hydrophytes (Pistia stratiotes and Eichhornia crassipes) which were not previously exposed to heavy metals were collected from the Botanical Garden, Department of Botany, University of the Punjab, Lahore, Pakistan. Plant nurseries were maintained and equal sized plats were selected in the chromium removal experiments in combination with bacterial strains. **16S rRNA ribotyping**

For the molecular taxonomic point of view, strains S4 and S5 which can resist K₂CrO₄, 16S rRNA gene sequencing was carried out. 16S rRNA gene (1300 bp) was amplified and the amplicon sequenced using fluorescent di-deoxy terminator cycle sequencing chemistry. The extension product was then separated on an ABI PRISM automated DNA sequencer and the data was compared to the MicroSeq® databases.

Chromate accumulation

For chromate accumulation experiments, both strains were grown in nutrient broth supplemented with two concentration of K₂CrO₄ (200 and 400 µg ml⁻¹). The inoculum was prepared as follow. i) One gram of pellet was taken and heatkilled at 121°C for 15 minutes (Heat killed) and ii) same amount was used untreated (live cell mass). Inoculum was given in 250 ml flask and was incubated at 37°C. After different time incubation (30 min and 720 min), cells were harvested by centrifugation and washed thrice with autoclaved distilled water. Pellets obtained were digested and the total amount of accumulated chromium was determined by first oxidizing any trivalent chromium into hexavalent with KMnO₄ and

analyzed spectrophotometrically at 540 nm in a spectrophotometer using diphenylcarbazide as the complexing agent¹⁸. All experiments were conducted in triplicate.

Cr (VI) reduction experiments

Bacterial Cr (VI) reduction was monitored at two different concentrations of K₂CrO₄ (200 and 400 µg ml⁻¹). The Cr(VI) reduction medium consist of following ingredients (grams per liter: Tryptone 10, yeast extract 5, NaCl 5, citric acid 1 and Na₂HPO₄ 6.9)¹⁹. Medium was prepared in 250 ml flask and after sterilization the medium was supplied with 200 and 400 µg ml⁻¹ of K₂CrO₄. Fresh inoculum was given in these flasks and incubated at 37°C at 150 rpm. After 24 h, samples were withdrawn aseptically and were analyzed for Cr (VI) reduction. The analysis of Cr (VI) reduction by these strains was checked by using the classical spectrophotometeric method¹⁸ using Diphenylecarbazide. The absorption was monitored at 540 nm.

Cr (VI) reduction by bacterial supernatant

To check the Cr (VI) reduction efficiency of bacterial supernatant, cell were grown and centrifuged at 10,000 x g for 10 min. The supernatant from 24 h old cultures were obtained and utilized both as such and after autoclaving (to denature the protein involved in Cr (VI) reduction). Initial K_2CrO_4 concentration used for this experiment was 200 and 400 µg ml⁻¹. The ratio of Cr (VI) reduction medium and supernatant was 1:1.

Cr removal by bacteria in combination with hydrophytes

To investigate the conjunctive effect of both plants and bacteria on the Cr removal ability, equal sized plants were selected and were exposed to an initial K_2 CrO₄ concentration of 50 µg ml⁻¹. Bacterial inoculum was given from freshly prepared overnight cultures. Inoculum of 100 µl of both strains was added in the solution. The incubation was done at plant growth chamber with controlled light and temperature for 15 days. After 15 days incubation, plants were harvested and Cr concentration in both plants was assayed by utilizing oven dried plant material. Cr left in the supernatant and accumulated by bacterial strains was also analysed.

Statistical analysis

Standard errors of the means were calculated following Steel and Torrie²⁰.

RESULTS

Strains characterizations

Both these strains could resist high concentration of K_2CrO_4 both on nutrient agar (up to 20 mg ml⁻¹) and acetate minimal medium (2.2 mg ml⁻¹). Strain S4 is a gram positive aerobic spore forming rod while strain S5 is gram negative facultative anaerobic motile rod. The results of 16Sr RNA gene sequencing revealed that strains S4 and S5 identified as *Bacillus pumilus*-S4 and *Pseudomonas doudoroffii*-S5, respectively. Both strains cloud resist multiple heavy metals and

antibiotics. The optimum temperature and pH for maximum growth of these strains is 37°C and 7, respectively.

Cr accumulation

Fig. 1 revealed that Cr accumulation in these strains is metabolically linked. After 30 min of incubation, heat killed cells of strain *B. pumilus*-S4 accumulated 5.7 and 8.9 μ g K₂CrO₄ g⁻¹ dry weight, at an initial K₂CrO₄ concentration of 200 and 400 μ g ml⁻¹, respectively. While in the same time period living cells of this strain accumulated 6.8 and 9.2 μ g K₂CrO₄ g⁻¹ dry weight (Fig. 1). But after 720 min of incubation, the amount of Cr



Fig. 1. Chromium accumulation potential of bacterial strains (*B. pumilus*-S4 and *P. doudoroffii*-S5) at different initial K_2CrO_4 concentrations of 200 and 400 µg ml⁻¹ of K_2CrO_4 after contact time of 30 min and 720 min at 37°C and pH 7. Cells were used as heat killed and live

accumulated by living cells ($45.6 \,\mu g \, K_2 Cr O_4 g^{-1} \, dry$ weight) was much higher (5 to 6 times) as compared to heat killed cells ($7.2 \,\mu g \, K_2 Cr O_4 g^{-1} \, dry$ weight) (Fig. 1). Almost same trend was observed in case of strain *P. doudoroffii*-S5.

Cr (VI) reduction

Cr (VI) reduction in these strains was monitored at various initial Cr (VI) concentrations (200 and 400 μ g ml⁻¹) and incubation times (12, 36 and 72 h). Strain *B. pumilus*-S4 reduced showed 28, 69 and 85% of Cr (VI) after 12, 36 and 72 h, respectively, at an initial K_2CrO_4 concentration of 200 (Fig. 2). At higher initial Cr (VI) concentration (400 µg ml⁻¹), the reduction % age decreased but overall more amount of chromate was reduced after 72 h of incubation (Fig. 2). After 72 h of incubation, strain *B. pumilus*-S4 reduced 64% while strain *P. doudoroffii*-S5 reduced 52% of Cr (VI) at an initial K_2CrO_4 concentration of 400 µg ml⁻¹ (Fig. 2).

Cr (VI) reduction by bacterial supernatant

To check the effect of bacterial supernatant on chromium reduction, the



Fig. 3. Hexavalent chromium reduction potential of bacterial supernatant (*B. pumilus*-S4 and *P. doudoroffii*-S5) in the presence of 200 and 400 μ g ml⁻¹ of K₂CrO₄ at pH 7 and 37°C for 24 h incubation period. Bacterial supernatant was used i) as such ii) autoclaved at 121°C for 15 min to denature the protein

supernatant from 24 h old cultures were obtained and utilized both as such and after autoclaving (to denature the protein involved in Cr (VI) reduction). Two different initial Cr (VI) concentrations (200 and 400 μ g ml⁻¹) were used in this experiment. Unautoclaved supernatant of strain *B. pumilus*-S4 was able to reduced 11% of Cr (VI) supplied after 24 h at an initial K₂CrO₄ concentration of 200 μ g ml⁻¹ (Fig. 3). While in the same time interval, autoclaved supernatant of this strain reduced 2.4% of the 200 μ g ml⁻¹ of K₂CrO₄ which is much less compared to un-autoclaved supernatant.

Effect of hydrophytes on Cr removal

After two weeks of incubation, in the

presence of *B. pumilus*-S4, *Eichornia crassipes* accumulated 24% alone and 47% in bacterial presence at an initial K_2CrO_4 concentration of 50 µg ml⁻¹ (Fig. 4). In the same time period and metal concentration bacterial strain *B. pumilus*-S4 removes 38% of Cr alone. While in the same time period, *Pistia stratiotes* removes 43% of Cr in the presence of *B. pumilus*-S4.

Almost same trends were seen in case of *P. doudoroffii*-S5 where strain alone removes 32% while in the presence of *Eichornia crassipes* and *Pistia stratiotes* removes 44% and 37% of Cr, respectively, at an initial K_2CrO_4 concentration of 50 µg ml⁻¹ after two weeks (Fig. 4).



Fig. 4. Chromium removal potential of bacterial strains (*B. pumilus*-S4 and *P. doudoroffii*-S5) in the presence of *Eichornia crassipes* and *Pistia stratiotes* at an initial K_2CrO_4 concentration of 50 µg ml⁻¹ for 15 days at 37°C. Plants and bacterial strain were used alone and in combinations

DISCUSSION

Increased pollution with an alarming rate not only affects our daily life but also changed the quality of ecosystem²¹. Hazardous chemicals released from industries are directly thrown in their wastewaters which get their way into fresh water²². Basically the present work deals with the detoxification/removal of carcinogenic Cr (VI) both by bacteria (*B. pumilus*-S4 and *P. doudoroffii*-S5) alone and in combination with free floating hydrophytes (*Eichornia crassipes* and *Pistia stratiotes*). Both these strain could resist Cr (VI) both on acetate minimal (2.2 mg ml⁻¹) and rich media (up to 20 mg ml⁻¹). These strains also show multiple heavy metal and antibiotic resistances. This resistance is a positive point because it will help the strain for their survival in the waste water treatment plant when ever they will be used for bioremediation purpose.

The present Cr accumulation study revealed that the accumulation is metabolically energy dependent mechanism. In the first 30 min the amount of Cr accumulated in the heat killed and living cells was not much different. But with the passage of time the heat killed cells become saturated and no further accumulation occurred. Also some Cr accumulation/deposition apartment

within the cell is also deformed to the heat. But in living cells the Cr accumulation was observed even after 720 min. Pawel *et al*²³ also observed that in yeast cells the amount of Cr accumulation was occurred in two phase, a rapid accumulation phase was followed by a slow time dependent increase. Ozdemir *et al*²⁴ while working on *Pantoea* sp. TEM18 showed that the metal adsorption increased during the first 15 min and after that its goes in to an equilibrium period.

Both these strains reduced Cr (VI) aerobically using citrate as one of the efficient electron donor. The %age of chromate reduction increased with increased in time period. P. doudoroffii-S5 reduced 14.2 mg Cr (VI) 36 h⁻¹ at an initial concentration of 200 µg K₂CrO₄ ml⁻¹. It was observed that at 400 µg K₂CrO₄, the %age of Cr (VI) reduced was less but overall more amount of Cr (VI) was reduced in both strains. Verma et al²⁵ observed that strain Bacillus brevis isolated from tannery effluent could reduced 75.8% of Cr (VI) at an initial concentration of 180 µg Cr ml⁻¹ within 28 h of incubation. Bacteria can reduce Cr (VI) both by soluble protein fractions and membrane fractions. In present case we have also checked the Cr reduction ability of soluble protein present in the supernatant and this activity was significantly decreased by de-nature these proteins. It means in both theses strains (B. Pumilus-S4 and P. doudoroffii-S5) a well developed chromate reduction system associated with soluble protein exists. Beside microorganisms, certain plants are known to accumulate heavy metals, trace elements, organic or radioactive compounds in soils, groundwater, and industrial waste from their environment²⁶. The wetland plants including free floating hydrophytes gained much attention because of their rhizofiltration and phytofiltration of toxic metals²⁷. In the present work it was observed that the amount of Cr removal by bacterial strains was accelerated in the presence of hydrophytes (Pistia stratiotes and Eichhornia crassipes). Besides this they will support each other growth. Because the plants release carbon and energy source in the form of root exudates which help the bacteria strains. In turn these bacterial strains produce and liberate phytohormones which support hydrophytes growth. Becerra-Castro et al28 also study the interaction of bacterial strains and plants to solubilise Ni in the soil and potentially improve phytoextraction strategies.

CONCLUSIONS

These results suggest that both these strains efficiently uptake and reduce toxic Cr (VI) from the solution and this process is augmented in the presence of *Pistia stratiotes* and *Eichornia crassipes*. So the present bioremediation method is a promising step towards the treatment of industrial waste water through the use of bacterial strains along with metal accumulator hydrophytes.

ACKNOWLEDGMENTS

Higher Education Commission, Pakistan is greatly acknowledge for the financial support for this work.

REFERENCES

- Hoffmann, B., Moebus, S., Dragano, N., Mohlenkamp, S., Memmesheimer, M., Erbel, R., Jockel, K.H. Residential traffic exposure and coronary heart disease: results from the Heinz Nixdorf Recall Study. *Biomarkers*, 2009; 14: 74-78.
- 2. Emenike, C.U., Fauziah, S.H., Agamuthu, P. Whole effluent toxicity from active and inactive landfills: Variation of leachate impact on fish (*Pangasius sutchi*). *Adv. Sci. Lett.*, 2013; **19**: 2681-2689.
- Bhattacharya, P., Banerjee, P., Mallick, K., Ghosh, S., Majumdar, S., Mukhopadhyay, A., Bandyopadhyay, S. Potential of biosorbent developed from fruit peel of *Trewia nudiflora* for removal of hexavalent chromium from synthetic and industrial effluent: Analyzing phytotoxicity in germinating *Vigna* seeds. J. Environ. Sci. Hlth, Part A., 2013; 48: 706-719.
- Wurm, J., Morrell, P. Use of platinised titanium anodes in chrome plating applications in aircraft industry. *Trans. Institute Metal Finish.*, 2010; 88: 17-20.
- Aghdassi, E., Arendt, B.M., Salit, I.E., Mohammed, S.S., Jalali, P., Bondar, H., Allard, J.P. In patients with HIV-infection, chromium supplementation improves insulin resistance and other metabolic abnormalities: a randomized, double-blind, placebo controlled trial. *Curr. HIV Res.*, 2010; 8: 113-120.

Arakawa, H., Wu, F., Costa, M., Rom,

6.

W., Tang, M.S. Sequence specificity of Cr(III)– DNA adduct formation in the p53 gene: NGG sequences are preferential adduct-forming sites. *Carcinogenesis.*, 2006; **27**: 639-645.

- Kerger, B., Butler, W., Paustenbach, D., Zhang, J.D., Li, S. Cancer mortality in Chinese populations surrounding an alloy plant with chromium smelting operations. *J. Toxicol. Environ. Hlth Part A.*, 2009; **72**: 329-344.
- Camargo, F.A., Okeke, B.C., Bento, F.M., Frankenberger, W.T. *In vitro* reduction of hexavalent chromium by a cell free extracts of *Bacillus* sp. ES 29 stimulated by Cu (2+). *Appl. Microbiol. Biotechnol.*, 2003; 62: 569-573.
- Mnif, S., Chamkha, M., Sayadi, S. Isolation and characterization of Halomonas sp. strain C2SS100, a hydrocarbon-degrading bacterium under hypersaline conditions. J. Appl. Microbiol., 2009; 107: 785-794.
- Doshi, H., Ray, A., Kothari, I.L. Bioremediation Potential of Chlorella: Spectroscopic, Kinetics, and Sem Studies. *Int. J. Phytoremediat.*, 2008; 10: 264-277.
- Kim, G.H., Choi, Y.S., Kim, J.J. Improving the efficiency of metal removal from CCA-treated wood using brown rot fungi. *Environ. Technol.*, 2009; **30**: 673-679.
- Antiochia, R., Campanella, L., Ghezzi, P., Movassaghi, K. The use of vetiver for remediation of heavy metal soil contamination. *Anal. Bioanal. Chem.*, 2007; **388**: 947-956.
- Rehman, Y., Rizwan, M., Faisal, M., Hasnain, S. Seasonal effects of domestic wastewaters on the Cr (VI) reduction potential of *Bacillus cereus* S-6 and *Ochrobactrum intermedium* CrT-1. *Biol. Environ.*, 2011; **110B**: 33-40.
- Bhowmick, D.C., Bal, B., Chatterjee, N.S., Ghosh, A.N., Pal, S. A low-GC Grampositive *Thermoanaerobacter*-like bacterium isolated from an Indian hot spring contains Cr(VI) reduction activity both in the membrane and cytoplasm. *J. Appl. Microbiol.*, 2009; **106**: 2006-2016.
- Hanc, A., BaraBkiewicz, D., Piechalak, A., Tomaszewska, B., Wagner, B., Bulska, E. An analysis of long-distance root to leaf transport of lead in *Pisum sativum* plants by laser ablation-ICP-MS. *Int. J. Environ. Anal. Chem.*, 2009; 89: 65-659.
- Danh, L.T., Truong, P., Mammucari, R., Tran, T., Foster, N. Vetiver grass, *Vetiveria zizanioides*: a choice plant for phytoremediation of heavy metals and organic wastes. *Int. J. Phytoremediat.*, 2009; **11**: 664-691.

- Gerhardt, P., Murry, R.G.E., Wood, W.A., Kreig, N.R. Methods for general and molecular bacteriology. Washington, DC, USA, 1994.
- Clesceri, S.l., Greenberg, A., Eaton, A.D. Standard methods for the examination of water and wastewater (20th ed.). Washington, DC, USA.
- DeLeo, P.C., Ehrlich, H.L. Reduction of hexavalent chromium by *Pseudomonas fluorescens* LB 300 in batch and continuous cultures. *Appl. Microbiol. Biotechnol.*, 1994; 40: 756-759.
- Steel, R.G.D., Torrie, J.H. A Biometrical Approach. In: Principles and Procedures of Statistics. (2nd ed.). McGraw Hill International Book Company, 1981.
- Lone, M.I., He, Z.L., Stoffella, P.J., Yang, X.E. Phytoremediation of heavy metal polluted soils and water: Progresses and perspectives. J. *Zhejiang Uni. Sci.*, 2008; 9: 210-220.
- Ajmal, M., Mohammad, A., Yousuf, R., Ahmad, A. Adsorption behavior of Cadmium, Zink, Nickel, and Lead from aqueous solution by *Mangifera indica* seed shell. *Indian J. Environ. Hlth.*, 1998; 40: 15-26.
- Pawel, K., Daria, F., Helena, K., Lyuba, B., Dorota, W., Henryk, K. Chromium accumulation by living yeast at various environmental conditions. *Microbiol. Res.*, 2004; 159: 11-17.
- Ozdemir, G., Ceyhan, N., Ozturk, T., Akirmak, F., Cosar, T. Biosorption of chromium(VI), cadmium(II) and copper(II) by *Pantoea* sp. TEM18. *Chem. Eng. J.*, 2004; **102**: 249-253.
- 25. Verma, T., Garg, S.K., Ramteke, P.W. Genetic correlation between chromium resistance and reduction in *Bacillus brevis* isolated from tannery effluent. *J. Appl. Microbiol.*, 2009; **107**: 1425–1432.
- Li, S., Liu, R., Wang, H., Shan, H. Accumulations of Cadmium, Zinc, and Copper by rice plants grown on soils amended with composted pig manure. *Commun. Soil Sci. Plan.*, 40: 2009; 1889-1905.
- 27. Uysal, Y., Taner, F. Effect of pH, temperature, and lead concentration on the bioremoval of lead from water using *Lemna minor*. *Int. J. Phytoremediat.*, 2009; **11**: 591-608.
- Becerra-Castro, C., Monterroso, C., Garcia-Leston, M., Prieto-Fernandez, A., Acea, M J., Kidd, P S. Rhizosphere microbial densities and trace metal tolerance of the nickel hyperaccumulator *Alyssum serpyllifolium* Subsp. Lusitanicum. *Int. J. Phytoremediat.*, 2009; 11: 525-541.