# Effect of *Trichoderma harzianum* on *Meloidogyne javanica* in Tomatoes as Influenced by Time of the Fungus Introduction into Soil

# Muhammad TariqJaveed\* and Ahmad Saad Al-Hazmi

Department of Plant Protection, King Saud University, P.O Box 2460 Riyadh 11451, Saudi Arabia.

(Received: 07 February 2015; accepted: 10 March 2015)

A greenhouse pot experiment was conducted to determine the effect of time of introduction of *Trichoderma harzianum* on its biocontrol efficacy against *Meloidogyne javanica* in tomatoes. The experiment consisted of all possible combinations of individual, concomitant and sequential inoculations of *T. harzianum* and *M. javanica*. Results indicated that time of the fungus introduction into the soil proved to be important. Introduction of *T. harzianum* to *M. javanica*-infested soil at or prior (one or two weeks) to nematode inoculation suppressed the nematode reproduction and increased host growth. However, such effects were increasingly greater when the fungus was introduced two or one week prior to nematode inoculation.

**Key words:** Antagonistic effect, Application time, Biological control, Nematophagous fungi, Root-knot nematodes, *Solanum lycopersicum*.

The free-living soil fungus *Trichoderma* spp. is an important biological control agent of plant-parasitic nematodes <sup>1,2,3</sup>. Control of the root-knot nematodes (*Meloidogyne* spp.) by different species of *Trichoderma* has been reported by several scientists <sup>1,3,4,5,6,7,8,9,10</sup>.

The antagonistic organisms that are transferred with the soil must be given time to reproduce to suppressive levels<sup>11</sup>. *Trichoderma harzianum* (Rifai) was found to be more effective against *Meloidogyne javanica* (Treub) when both organisms were applied 18 days before transplanting of tomato seedlings than when both applied at transplanting<sup>12</sup>. *Paecilomyces lilacinus* (Thom) was more effective against *M. incognita* (Kofoid and White) Chit. when it was delivered into the nematode-infested soil ten days before planting of tomato<sup>13</sup>. However, Walia *et al.*<sup>14</sup>

reported that application of *P. lilacinus* 10 days before or after sowing of okra seeds were equally effective against *M. javanica*. When the algae *Microcoleus vaginatus* (Vauch.) Gomont was introduced 10 days prior to the *M. incognita*, damage of tomato plants was reduced and nematode density was suppressed<sup>15</sup>.

In previous *in vitro* and greenhouse tests (un-published data), we found that a local isolate of *T. harzianum* (isolate no. 27) was the most effective isolate, among eight tested isolates and species of *Trichoderma*, against *M. javanica*. This present study was conducted to determine the effect of time of introduction of *T. harzianum* into soil on its biocontrol efficacy against *M. javanica* on tomato.

#### MATERIALS AND METHODS

This study was conducted in the greenhouse (24±2°C). Thirty-day-old seedlings of tomato (cv. Sultana-7) were used, one seedling per pot (15 cm diam.). The soil of each pot (1500 g), a

E-mail: tariqjaved22@gmail.com

 $<sup>^{*}</sup>$  To whom all correspondence should be addressed. Tel: +966509720562;

mixture of sand, sandy loam and peat moss (2:1:1), was previously steam-sterilized with an autoclave for 30 minutes.

The experiment consisted of individual, concomitant and sequential inoculation of M. javanica and T. harzianum. The experiment included eight treatments (table 1), namely: 1) M. javanica alone (N); 2) T. harzianum alone (T); 3) both organisms simultaneously (N + T); 4) nematode first and then fungus one week later; 5) nematode first and then nematode one week later; 7) fungus first and then nematode two weeks later; and 8) control (non-inoculated seedlings).

The inoculum of *M. javanica* consisted of eggs which were extracted by the NaOCl method (Hussey and barker, 1973<sup>16</sup>) from the roots of a pure greenhouse culture of *M. javanica* on tomato. The egg suspension was adjusted to contain 2000 eggs/ml.

The fungus *T. harzianum* (isolate no. 27) was obtained from the mycological unit (Prof. Younes Yousef Molan), Department of Plant Protection at King Saud University, Riyadh, Saudi Arabia. This species was originally isolated, along with other species and isolates of Trichoderma, from soil samples collected from different agricultural fields in Riyadh region, Saudi Arabia, using dilution plate method onto Trichoderma selective media (TSM) according to Elad and Chet<sup>17</sup>. These fungal isolates were purified through subculturing from single spores and, then, identified to Species level by Prof. Younes Molan based on sequences of the Internal transcribed spacer regions 1 and 2 (ITS1 and ITS2) of the ribosomal DNA<sup>18,19</sup>.For inoculum preparation, T. harzianum (isolate no. 27) wasre-cultured on Potato Dextrose Agar (PDA) on petri plates and incubated at 24°C for 14 days. The produced conidia were collected from the culture surfaces by flooding with sterile distilled water and gently scraping the colony surface with a sterile scrapper. The suspension was, then, passed through cheese cloth in sterile distilled water<sup>20</sup>.

At inoculations with the nematode and/ or the fungus, each seedling was inoculated with 10000 eggs of M. javanica and/or $1 \times 10^{10}$  conidia/ g soil of the fungus T. harzianum. The nematode egg inoculum, suspended in 5 ml of water, was equally distributed through three small holes made

in the soil around the seedling stem. Inoculation with the fungus was made by distributing and mixing the fungal inoculum thoroughly with the soil surface of the designated pots. The eight treatments (table 1) were arranged on greenhouse bench in a complete randomized design. Seedlings were irrigated, and fertilized as needed, till the end of the test.

Fifty five days after last inoculation, the test was terminated. Fresh weight of plant shoots and rootswere recorded. Numbers of root galls, egg masses and eggs were counted. Second-stage juveniles (J2) in the soil were extracted by the modified centrifugal floatation method<sup>21</sup>. Final population densities of nematodes were determined and the reproduction factor (Rf)<sup>22</sup> was calculated. Data were statistically analyzed using analysis of variance (ANOVA), and treatments means were separated by protected Fischer's least significant difference (LSD) using SAS<sup>23</sup>.

#### RESULTS

Trichoderma harzianum increased (P< 0.05) shoot and total plant weights, whether introduced at or prior (1 and 2 weeks) to nematode inoculation, compared to inoculation with nematode alone (table 1). However, effects of the fungus introduction at/and prior to nematode inoculation were not different (P  $\leq$  0.05). Similarly, introduction of *T. harzianum* at all times reduced the number of root galls (table 1). However, such reduction of galls was greater when the fungus was introduced two or one weeks prior to nematode inoculation.

Reproduction of M. javanica was suppressed ( $P \le 0.05$ ) by T. harzianum at all times of the fungus introduction (table 2). However, the greatest suppression of the nematode reproduction was obtained when T. harzianum was introduced twoor one weeks prior to nematode inoculation. The least reproduction was achieved when the fungus was introduced two weeks before nematode inoculation (table 2).

### DISCUSSION

The introduction of *T. harzianum* to *M. javanica*-infested soil at or prior to nematode inoculation increased host growth and suppressed

nematode reproduction compared to controls. Our results support previous reports on the efficacy of different species and isolates of *Trichoderma* against *Meloidogyne* spp. <sup>4,7,24,25,26,27</sup>. The suppressive effects of *T. harzianum*, reported in this study, on nematode population and disease severity are strong evidence that considerable parasitism was occurring.

Our results indicate that time of the fungus introduction into the soil proved to be very important. Greater control effects were achieved

when *T. harzianum* was introduced two or one week prior to soil infestation with the nematode. Similar evidence was previously reported<sup>12</sup>. The extra time given to *T. harzianum* was very advantageous to have an established fungal population. A similar conclusion was suggested by Al-Hazmi *et al.*<sup>28</sup> working with the nematode-trapping fungus of *Arthrobotrys conoides* against *M. incognita* on corn.

Although *M. javanica* caused considerable crop damage to tomatoes in Saudi

<b>Table 1.</b> Effect of time of introduction of <i>Trichoderma harzianum</i> (T)	
on host response of tomato inoculated with <i>Meloidogyne javanica</i> (N).	

Treatment	Root weight	Shoot weight (g)	Total plant weight (g)	Gall/root system	Gall/g root
Healthy seedlings (control)	5.01 a	38.20 bc	43.21 bc	_	-
M. javanica alone (N)	5.18 a	35.05 c	40.23 c	816.00 a	157.45 a
T. harzianum alone (T)	5.21 a	41.20 ab	46.41 ab	-	-
N + T	5.80 a	41.10 ab	46.90 ab	492.25 c	86.86 b
N_1wk_T	5.33 a	37.91 bc	43.24 bc	539.50 bc	103.30 b
N² wk,T	5.81 a	38.67 abc	44.48 abc	600.75 b	103.96 b
T <del>1 wk</del> N	6.02 a	41.79 ab	47.80 ab	380.00 d	63.46 c
TwkN	5.25 a	43.15 a	48.40 a	308.50 d	61.96 c

Data are means of four replicates. Means, in each column, followed by the same letter(s) are not significantly different according to Fischer's protected least significant difference (LSD) test ( $P \le 0.05$ ).

 $N \rightarrow T = M$ . javanica followed by T. harzianum,  $T \rightarrow N = T$ . harzianum followed by M. Javanica, wk = week (s).

 $\begin{tabular}{ll} \textbf{Table 2.} Effects of time of introduction with $\mathit{Trichoderma}$ \\ \textit{harzianum}$ (T) on reproduction of $\mathit{Meloidogyne javanica}$ (N) on tomato \\ \end{tabular}$ 

Treatments	Egg mass/g	Eggs/g(X1000)	J2/100g soil	Rf*
Healthy seedling (control)				
M. javanica alone (N)	100.44 a	21.79 a	201.3 a	11.6 a
T. harzianum alone (T)				
N+T	21.061 d	07.74 d	125.8 bcd	04.7 d
N_1wkT	34.595 c	12.70 c	132.3 bc	06.9 с
$N \xrightarrow{2 \text{ w/k}} T$	55.752 b	17.86 b	138.8 b	10.5 b
$T \xrightarrow{1 \text{ wk}} N$	19.509 d	05.81 d	122.0 cd	03.6 e
Twk_₃N	20.208 d	05.77 d	116.8 d	03.1 e

Data are means of four replicates. Means, in each column, followed by the same letter(s) are not significantly different according to Fischer's protected least significant difference (LSD) test  $(P \le 0.05)$ .

<sup>\*</sup>Rf: Reproduction factor = Final nematode density (Pf)/Initial inoculum density (Pi).

 $N \rightarrow T = M$ . javanica followed by T. harzianum,  $T \rightarrow N = T$ . harzianum followed by M.Javanica, wk = week (s).

Arabia, no accurate yield losses were reported. *Meloidogyne javanica* is widespread in our agricultural soils<sup>29</sup> may our native *T. harzianum* isolates negate some of the damage on tomatoes caused by *M. javanica*.

#### CONCLUSION

Our results conclude that time can be an important factor in success of fungal biocontrol agent. Results showed that when *T. harzianum* isolate are introduced into soil at or prior to nematode inoculation they can be more effective in managing *M. javanica* because they need certain time to be established in soil.

# **ACKNOWLEDGEMENTS**

The authors are very thankful to Prof. Y. Yousef Molan for providing the pure culture of *T. harzianum*, and Dr. Mahmood El-Komi for assistance in the laboratory. The authors also gratefully acknowledge their appreciation to the Agriculture Research Center at the College of Food and agricultural Sciences and Deanship of Scientific Research, King Saud University for supporting this research work.

# REFERENCES

- Spiegel, Y., Sharon, E., Bar-Eyal, M. Evaluation and mode of action of *Trichoderma* isolates as biocontrol agents against plant-parasitic nematodes. *IOBC WPRS Bull*, 2007; 30: 129– 133.
- Chen, S., Dickson, D.W. Biological control of nematodes by fungal antagonists. In: Nematology Advances and Perspectives. Nematode Management and Utilization, vol. II. (Chen ZX, Chen SY, Dickson DW, eds). Wallingford: CAB International, UK, 2004; pp. 977-1039.
- 3. Sharon, E., Chet, I., Spiegel. Y. Trichoderma as a biological control agent. In: Biological Control of Plant-Parasitic Nematodes: Building Coherence between Microbial Ecology and Molecular Mechanisms, Progress in Biological Control (Davies K, Spiegel Y, (eds). Springer Netherlands, 2011; pp. 183-201.
- 4. Sharon, E., Bar-Eyal, M., Chet, I., Herrera-Estrella, A., Kleifeld, O., Spiegel, Y. Biological control of the root-knot nematode *Meloidogyne*

- *javanica* by *Trichoderma harzianum*. *Phytopathology*, 2001; **91**: 687–693.
- 5. Sharon, E., Chet, I., Viterbo, A., Bar-Eyal, M., Nagan, H., Samuels, G.J., Spiegel, Y. Parasitism of *Trichoderma* on *Meloidogyne javanica* and role of the gelatinous matrix. *Eu. J. Plant Pathol.*, 2007;**118**: 247-258.
- Affokpon, A., Coyne, D.L., Htay, C.C., Agbèdè, R.D., Lawouin, L., Coosemans, J. Biocontrol potential of native *Trichoderma* isolates against root-knot nematodes in West African vegetable production systems. *Soil Biol. Biochem.*, 2011; 43: 600-608.
- 7. Mascarin, G.M., Junior, M.F.B., Filho, J.V. de A. *Trichoderma harzianum* reduces population of *Meloidogyne incognita* in cucumber plants under greenhouse conditions. *J. Entomol. Nematol.* 2012; **4**: 54-57.
- 8. Naserinasab. F., Sahebani, N., Etebarian, H.R. Biological control of *Meloidogyne javanica* by *Trichoderma harzianum* BI and salicylic acid on tomato. *Afric. J. Food Sci.*, 2011; **5**(3): 276 280.
- 9. Rao, M.S., Reddy, P.P., Nagesh, M. Evaluation of plant based formulations of *Trichoderma harzianum* for the management of *Meloidogyne incognita* on eggplant. *Nematol. Medit.*, 1998; **26**: 59–62.
- AL-Shammari, T.A., Bahkali, A.H., Elgorban, A.M., El-Kahky, M.T., Al-Sum, B.A. The use of *Trichoderma longibrachiatum* and *Mortierella* alpina against root-knot nematode, *Meloidogyne* javanica, on tomato. J. Pure appl. Microbiol., 2013; 7: 199-207.
- 11. Timper, P. Conserving and enhancing biological control of nematodes. *J. Nematol.*, 2014; **46**(2): 75–89.
- Golzari, H., Panjeh, N., Ahmadzadeh, M., Salari, M., Sedaghati-khoravi, E. Elucidating the parasitic capabilities of *Trichoderma* against *Meloidogyne javanica* on tomato. *Insight Plant Dis.*, 2011; 1(1): 12-19.
- Cabanillas, E. Barker, K.R. Impact of Paecilomyces lilacinus inoculum and application time on control of Meloidogyne incognita on tomato. J. Nematol., 1989; 21(1):115-120.
- Walia, R.K., Bansal, R.K. and Bhatti, D.S. Effect of *Paecilomyces lilacinus* application time and methods in controlling *Meloidogyne javanica* on okra. *Nematol. Medit.* 1991; 19: 247-249.
- 15. Khan, Z., Park, S.D. Effects of inoculum level and time of *Microcoleus vaginatus* on control of *Meloidogyne incognita* on tomato. *J. Asia-Pacific Entomol.* 1999; **2**(2):93-96.
- 16. Hussey, R.S, Barker, K.R. A comparison of

- methods of collecting inocula for *Meloidogyne* spp., including a new technique. Plant Dis. Rep. 1985; **57**: 1025-1028.
- 17. Elad, Y., Chet. I. Improved selective media for isolation of *Trichoderma* spp. or *Fusarium* spp. *Phytoparasitica*, 1983; **11**: 55-58.
- Maymon, M., Minz, D., Barbul, O., Zveibil, A., Elad, Y., Freeman, S. Identification of *Trichoderma* biocontrol isolates to clades according to ap-PCR and ITS sequence analyses. *Phytoparasitica*, 2004; 32(4):370-375.
- Hermosa, M.R., Grondona, I., Iturriaga, E.A., Diaz-Minguez, J.M., Castro, C., Monte, E., Garcia-Acha, I. Molecular characterization and identification of biocontrol isolates of *Trichoderma* spp. *Appl. Environ. Microbiol.*, 2000; 66(5): 1890–1898.
- Jansson, H-B., Jeyaprakash, A., Zuckerman, B.M. Control of root-knot nematodes on tomato by the endoparasitic fungus *Meria coniospora*. *J. Nematol.*, 1973; 17: 327-329.
- Barker, K.R.: Nematode extractions and bioassays. In: An advanced treatise on Meloidogyne Vol. II. Methodology (Barker KR, Carter CC, Sasser JN, eds). Raleigh: North Carolina State University Graphics, N. C. USA, 1985; pp. 19-35.
- Oostenbrink, M. Major characteristics of the relation between nematodes and plants. Mededlingen Landbouwhoge School. Wageningen, 1966; 66-4: 1-46.

- SAS Institute. SAS/STAT. User's Guide version. 13.0 Cary, NC, USA, SAS Institute.2013.
- Siddique, I.A., Amer-Zareen, Zaki, M.J., Shaukat, S.S. Use of *Trichoderma* species in the control of *Meloidogyne javanica*, rootknot nematode of in okra and mungbean. *Pak.J. Biol. Sci.*, 2001; 4: 846-848.
- Dababat, A.A., Sikora, R.A. Use of *Trichoderma harzianum* and *Trichoderma viride* for the biological control of *Meloidogyne incognita* on Tomato. *Jord. J. Agri. Sci.*, 2006; 3: 953-961.
- 26. Windham, G.L., Windham, M.T., Williams, W.P. Effect of *Trichoderma* spp. on maize growth and *Meloidogyne arenaria* reproduction. *Plant Dis.* 1989; **73**: 493-494.
- Pandey, G., Pandey, R.K., Pant, H. Efficacy of different levels of *Trichoderma viride* against root-knot nematode in chickpea (*Cicer arietinum* L.). *Annals Plant Protection Sci.*, 2003; 11: 96-98
- 28. Al-Hazmi, A.S., Schmitt, D.P., Sasser, J.N. The effect of *Arthrobotrys conoides* on *Meloidogyne incognita* population densities in corn as influenced by temperature, fungus inoculum density, and time of fungus introduction in the soil. *J. Nematol.*, 1982; **14**(2):168-174.
- 29. Al-Hazmi, A.S. Host-index of plant-parasitic nematodes in the Kingdom of Saudi Arabia. *J. Coll. Agric.*, King Saud Univ., 1984; **6**: 69-85.