Development of Chitosan Silver Nanocomposites: Its Characteristic Study and Toxicity Effect against 3T3-L1 Cell Line

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Abstract

Nanocomposite from the natural source is opened a wide area for the researchers to find a natural remedy to replace the chemicals or harmful products in all the fields of agricultural, food and medical fields. Here the biopolymer (chitosan) was extracted from the two white rot fungi of Pleurotus floridanus and Pleurotus djamor, and biologically synthesized with 1mM AgNO3 solution. Synthesized chitosan nanocomposite was characterized with UV-Visible study, FTIR, FESEM, XRD, EDAS for the confirmation based upon the peaks, functional group, crystalline nature, size, morphology and the percentage of elements respectively. Toxicity study was carried out using 3T3 L1 (Mouse embryo fibroblast Cell Line) normal Cell Line to find out the cytotoxicity effect of the chitosan nanocomposite and found that the nanocomposites were non toxic to the Cell Line.

Keywords: Nanocomposite, FESEM, XRD, EDAS, 3T3-L1 Cell Line

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INTRODUCTION

From the middle of 20th century, the world’s population has engaged to find out a better remedy to treat various diseases caused by microorganisms. Improper use of the chemicals in clinical treatment in living animals has prompted the different of pathogens that are resistant to the variety of compounds and less effect in treatment. Significant changes in the generation of bacteria to the antibiotic available on the market have been observed.1,2 Pharmaceutical industries are facing issues for finding new or innovative drug making because of the changes of genetic elements of the pathogens. This is opening a new way to find an antimicrobial compound.

Currently, nanotechnology and nano-materials are getting more priority in all the fields of agriculture, pharmacy, food industry, etc. Owing to the unique property of the nanoparticle, various methods and research are going on in all the fields. Nanoparticles are commonly established in the perspective of nanotechnology and Nanoscience is a unit of scientific measurement from the nanometre of 1 – 100nm in size.3 In current occasion, metallic nanoparticles are showing unique properties and activities such as physical, biological and chemical synthesising. Thus this have more subject of extensive research in recent years.4,5

Metallic nanoparticles show signs of different properties in their size and shape dependent. Silver nanoparticle has high therapeutic potential value because of its good antimicrobial activity. This nanoparticle has high antimicrobial activity while using very small concentration. It is also identified as good anti-cancer agent.6 Bio synthesizing methods are employing natural compounds such as microorganism, animal source, plants, polysaccharide etc. are crop up as simple and alternative source as silver nanoparticle.7

Chitosan is a polysaccharide extracted from the chitin by deacetylation process. Most of the chitosan extraction is from crab or shrimp.8 Apart from marine source plants, fungi are rich in chitosan. Pleurotus spp. are most popular edible mushroom because of its temperature from 15-30°C, easy cultivation procedure, different substrate of agro residue, feed, vitamins, waste and weed for production of food, enzymes in addition to their waste detoxification properties and degradation.9 Edible fungi has biological effects as it contains important bioactive compounds. Mushrooms are using as a food before 2000 years ago because of the medicinal importance. 200 species of mushrooms are used as functional food in the world,10 but only 35 to 40 species are commercially cultivated for various purposes.11 The current research was done to synthesize the chitosan silver nanocomposite from the Pleurotus spp. (Pleurotus floridanus and Pleurotus djamor) characterization and cytotoxicity effect against 3T3-L1 Cell Line.

MATERIALS AND METHODS

Sub culturing and extraction of chitosan from Pleurotus spp.

Pleurotus spp. was collected from Tamilnadu Agricultural University and subcultured to Potato Dextrose Agar. Molecular identification was done and submitted to NCBI gene bank (Pleurotus floridanus- MG324371.1 and Pleurotus djamor- MG328900.1).12 After seeding, the plate was incubated at 30°C for 12 days. The grown fungi mycelium was reinoculated in MGYP medium and the chitosan was extracted after deproteinization, demineralisation and deacetylation.13

Preparation of chitosan nanocomposite solution

The extracted chitosan was dried and 1% of the chitosan solution was prepared in 2 % of acetic acid (2ml acetic acid and 98ml distilled water) and glucose (2g in 100 ml distilled water). The mixture was incubated in magnetic stirrer for 30 minutes for mixing and added 1% of NaOH.14 Then it was used for the silver nanoparticle synthesizing by mixing equal volume of 1mM silver nitrate. After blending, sample was incubated for 24 to 48 hrs at room temperature.

Characterisation of chitosan nanocomposites

Synthesized nanocomposites were primarily identified with color change from light brown to dark brown. The nanocomposite was analyzed by UV-Visible spectrophotometer from 300 -600nm using Labtronics LT 291. The functional group responsible for the synthesising was identified with FTIR from 4000cm⁻¹ to 400cm⁻¹ (Shimadzu). The size and morphology were identified with MIRA 3 TESCAN SEM- Field Emission Scanning Electron Microscope.15 Percentage of
the elements and the crystalline nature of the nanocomposites were identified with XRD and EDAS analysis using PAN analytical instrument.\textsuperscript{16}

Toxicity study

MTT Assay method\textsuperscript{12}, (3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenylethrazo-liumbromide) was used for toxicity study against 3T3-L1 (Mouse embryo fibroblast-Normal Cell Line) Cell Line. The cells were grown in DMEM medium supplemented with 10% fetal bovine serum albumin and incubated in CO\textsubscript{2} incubator with 5% CO\textsubscript{2} and 70-80% of humidity at 37°C for 24-72hrs. Cells was primarily confirmed by using inverted microscope (Unicon) and the Cell Line was transferred to 96 well plate. 100µl of the Cell Line was mixed with different concentration of the nanocomposite (25,50,100,200,400 µg/ml) and incubated for 24hrs in CO\textsubscript{2} incubator (5% CO\textsubscript{2}) at 37°C. 100µl of blank (DMSO), control (only Cell Line) and standard control (drug-doxorubicin-12.5 µM) was also maintained in 96well plate (with triplicates). After 24hrs of incubation, the cells was washed with 50µl of DMSO and trypsin. To the washed cells, 20µl of MTT dye was added and again incubated in CO\textsubscript{2} incubator for 24hrs. After incubation, using ELISA (Robonik-Readwell Touch Elisa Plate Analyser) reader the OD value was measured at 570nm, calculated the viability (formula: Viability =100 x (Control OD-Sample OD)/Control OD) and \textit{IC\textsubscript{50}} value using linear regression equation (\textit{Y}=\textit{M}x+\textit{C}, here \textit{Y}=50, \textit{M} and \textit{C} values from the Viability graph).

RESULTS AND DISCUSSION

Characterization of chitosan nanocomposites

UV-Visible study

As shown in Fig. 1, the extracted chitosan (11.5g of chitosan was obtained from 100g of \textit{Pleurotus} mycelium) was used for the synthesizing of silver nanocomposite. After 24 hrs of incubation the synthesized nanocomposite was characterized primarily with the color change (dark brown color observed).

UV-Visible study was done to determine the optical properties of the chitosan nanocomposite, which is widely used technique for the characterization of the various nanoparticle. Silver nanocomposite plasmon peak was observed at 460nm (Fig. 2), which indicated the presence of silver nanocomposite. Similar results were observed for the synthesis of chitosan silver nanoparticle extracted from the crustacean...
sample at 450nm.\textsuperscript{17} It has been reported that the 400nm above peaks may be related to synthesis from natural sources tend to present two or more populations with diverse proportions and the presence of NaOH in the synthesis.\textsuperscript{18}

UV-Visible spectroscopy shown peaks at 390nm and 420nm for the chitin and chitosan silver nanoparticle extracted from fungi \textit{F. solani}.\textsuperscript{19} Honary et al, synthesised with different molecular weight chitosan and observed the plasmon peaks between 400-420nm.\textsuperscript{20} Synthesized chitosan silver nanoparticle using s-triazinyl-bis (2-aminomethylpyridine) and s-triazinylbis (8-oxyquinoline) got a high peak at 420nm.\textsuperscript{21} In another study chitosan silver nanoparticle from \textit{Saccharum officinarum} exhibited peak at 460nm.\textsuperscript{22} These findings are co inside with our results.

**FTIR**

The synthesized silver nanocomposite solution was subjected to Fourier Transform Infrared spectroscopy study, to identify the functional groups. Obtained peaks are

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**Fig. 2.** UV-Visible spectroscopy analysis.

**Fig. 3.** FTIR study of the chitosan nanocomposite.
3340 cm\(^{-1}\) (alcohol OH stretch), 1643 cm\(^{-1}\) (C=C alkene), 678 cm\(^{-1}\) and 601 cm\(^{-1}\) (C-Cl), 555, 455 and 424 cm\(^{-1}\) (C-I strong bond). FTIR spectrum peaks at 3431, 1635, 1390, 1274, 556 cm\(^{-1}\), which represent amide linkage group and band at 1635 cm\(^{-1}\) is an amide I bond. The peak at 1390 cm\(^{-1}\) is a CH\(_3\), and at 1274 cm\(^{-1}\) corresponded to single bond stretching of carboxylic acids, the peak at 3431 cm\(^{-1}\) corresponded to the stretch OH groups. Peak at 556 cm\(^{-1}\) belonged to the single bond Br stretch of alkyl halides (Fig. 3).

In the IR spectrum, 3500 - 3200 cm\(^{-1}\) was more asymmetric and indicated the existence of OH and NH\(_2\) groups. Peak at 1680 cm\(^{-1}\) in was due to oscillations of C=O in amino groups and acetyl amino groups, i.e. a deacetylated appearance of chitin. C-CH\(_3\) deformation in amide group observed at 1384 cm\(^{-1}\). 1050 cm\(^{-1}\) indicated the saccharide after the absorption of the silver ions. FTIR peaks from 650 to 400 cm\(^{-1}\) is denoting the reducing deed of chitosan and precipitation of metallic silver and formation of silver nanocomposites.

**FESEM**

Examination of FESEM image revealing that the synthesized chitosan silver nanocomposite has spherical in shape and the size from 50.24 to 88 nm (Fig. 4).

Sharanya Devadiga et al, found 96.3 nm size of the chitosan silver nanocomposites, extracted from the crustacean sample. Another study reported that the synthesised chitosan composite had 16 nm in size with more porous upon cross linking nature obtained from fungi. Nanoparticles with less than 100 nm in diameter were attracting with the efficiency of the sample in the various field of industry. The nanoparticles size was showing difference in their materials, as a results of quantum confinement, particle size, large surface area etc.

![Fig. 4. FESEM image.](image1)

![Fig. 5. EDAS analysis of chitosan nanocomposite.](image2)
EDAS

EDAS is an important technique to identify the elemental composition of the synthesised sample. All the elements have its own atomic structures producing a unique set of peaks in X-ray spectrum which can be used for the identification of the element composition (Fig.5). The chitosan silver nanocomposites were showing different elements of Carbon (22.36%), Oxygen (38.15%), Sodium (13.66%), Silicon (1.2%), silver (17.85), Calcium (6.78%). Lovejoy et al, reported that the elemental composition of the sample or a material identifying with EDAS analysis denotes the presence unique elements with weight and atomic concentration.27

There for in EDAS study, presence of silver and the other elements of carbon and oxygen signals are due to the chitosan that surrounds the silver nanoparticle while its get synthesised.28 Other study quantified the presence of elements such as silver (1.36%) sulfur (0.84%), silicon (67.74%) and oxygen (30.07%) from the synthesised chitosan composite,29 these resembles our findings.

XRD

Examination of XRD is an essential method to identify the crystalline structure of the nanoparticle along with the identification of phase composition and presence of impurities. Modrzyewska et al stated that the adsorption of Ag+ ions changes crystalline or the fibrous structure of the chitosan polymer when it get synthesised.25 The X-ray analysis revealing the Ag metal ions and the degree of ordering of the tested sample is reduced because of the adsorption and synthesis of silver nanocomposites.23 The numbers of reflections with 2θ values of 38.53°, 44.67°, 65.08° and 78.06° are corresponding to (1 1 1), (2 0 0), (220) and (3 1 1), which indicating the AgNPS spherical structure and crystalline nature.

Fig. 6. XRD analysis of chitosan nanocomposite.

Fig. 7. 3T3 Cell Line under microscope; 1-Control Cell Line, 2-standard drug, 3-25µg, 4-50 µg, 5-100 µg, 6-200 µg, 7-400 µg of sample treated against 3T3 Cell Line.
The sharp peak indicates bioactive compounds in the chitosan nanocomposite. Structural properties of the synthesized chitosan silver nanocomposite is clear from Fig. 6. *2θ* was observed at 38.47 and 77.67 with the height of 77.87 and 27.32 respectively (Fig. 6). Chitosan peaks were obtained at 2h values of 11.7, and 19.8 and silver peaks were pick up at 37.9, 44.0 and 63.9, respectively. In another study, peaks were observed for chitosan at 2θ=10.1° and 2θ=19.9°. The peak at 2θ=19.9° indicated the high degree of chitosan and their crystal lattice constant and answering to 4.4. The numbers of Bragg mirroring of distinct peaks at 38°, 44.1°, 64° and 77° specify the miller indices (111), (200), (220) and (311) reflections of metallic silver represented the face-centered cubic crystalline structure of silver.

**Toxicity study**

The toxicity study of the chitosan nanocomposites was done using the 3T3-L1 Cell Line employing MTT assay method. The increasing interest in the use of natural source as a remedial measure against obesity is because of the presence of chemical compounds in the commercial drugs and leads to some side-effects. For this reason, anti-obesity agents from natural products are being studied and to be commercialised. As for chitin/chitosan-derived biomaterials, conflicting reports have recently been reported because of the anti-obesity effects.

Another study reported that the chitosan and Chitosan Oligosaccharide supplementation are helping to decrease the plasma cholesterol and increased cholesterol excretion in overweight normcholesterolemic humans.

From the Fig. 8, the toxicity effect of the chitosan nanocomposite was found to be non toxic. 25, 50, 100, 200, 400 μg/ml chitosan nanocomposite shown 97.09±0.009, 92.54±0.006, 83.94±0.005, 71.680.004, 69.65±0.001 percentage of cell viability with the 620 µg/ml of IC₅₀ value. The standard doxorubicin (12.5 μg/ml) shows 72% of cell viability.

Sumiyoshi and Kimura investigated the positive roles of chitosan for hypocholesterolemic effects in mice. Ranjitha et al, reported *Terminalia arjuna* extract has been found 98.55 to 90.88% (10 μg to 320 μg/ml) of viable cells compare with untreated 3T3-L1 Cell Line.

**CONCLUSION**

Our previous publication was detailed about the extraction of chitosan from the *Pleurotus floridanus* and *Pleurotus djamor*. The current research work was focused on the biogenic synthesis of chitosan AgNO₃ nanocomposites. Further, it was characterised and tested against the 3T3-L1 Cell Line and found that the synthesised nanocomposite was non toxic. In UV –Visible spectrophotometer study, peak maxima was
observed at 460nm, and FTIR also showed different functional group and confirmed the synthesis of silver nanocomposite. Size and morphology of the nanocomposites were determined with FESEM and XRD, 50.24 to 88nm of size was observed with spherical in shape. Carbon (22.36%), Oxygen (38.15%), Sodium (13.66%), Silicon (1.2%), silver (17.85), Calcium (6.78%) were observed in EDAS analysis. The nanocomposites exhibited 69.65% of viable cells and non toxic to 3T3-L1 Cell Line at the higher concentration (400µg/ml). Further research is needed to improve and find the various application of the chitosan nanocomposites in food industry and other pharmaceutical effects.

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The authors declare that there is no conflict of interest.

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DATA AVAILABILITY
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ETHICS STATEMENT
This article does not contain any studies with human participants or animals performed by any of the authors.

REFERENCES


