

RESEARCH ARTICLE

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Ethnobotanical Study of Underutilized Wild Edible Fruits and their Antibacterial Activity against Human Pathogens

Suresh Ch. Biswas^{1,4} , Pranab Dutta^{2*} , Purnima Barua³ ,
Tarun Kr. Misra⁴  and Subrata Das⁵ 

¹Krishi Vigyan Kendra, Khowai - 799 207, Tripura, India.

²School of Crop Protection, College of Post Graduate Studies in Agricultural Sciences, Central Agricultural University, Umiam - 793 103, Meghalaya, India.

³Department of Microbiology, Jorhat Medical College Jorhat - 785 001, Assam, India.

⁴Department of Chemistry, National Institute of Technology Agartala - 799 046, Tripura, India.

⁵Department of Chemistry, National Institute of Technology Patna - 800 005, Bihar, India.

Abstract

Nine underutilized wild edible fruits collected from four districts of Tripura, a North-Eastern state of India were studied for the antibacterial properties against five (5) human pathogens, *Escherichia coli* (Gram–Ve, ATCC-2522), *Pseudomonas aeruginosa* (Gram–Ve, ATCC-27853), *Salmonella typhi* (Gram –Ve, ATCC-14028), *Staphylococcus aureus* (Gram +Ve, ATCC-25923) and *Klebsiella pneumonia* (Gram–Ve, ATCC-700603) by disk diffusion method. Besides, the folk medicinal values and their ethnobotanical importance are also reported. Out of the nine (9) wild edible fruits, *C. macroptera* and *S. dulcis* showed the least antibacterial effect against all the bacterial pathogens under the present study. On the other hand, *G. gummi-gutta* and *A. lacucha* showed considerable antibacterial effects against all of the bacterial pathogens with the highest zone of inhibition against *E. coli*, followed by *P. aeruginosa*. Selective inhibitory effect of some of the wild edible fruits was also observed against the targeted pathogens. This study paves the way for using the underutilized wild fruits of the North-Eastern region of India as a potential source of antipathogenic activity of human pathogens. Further, a detailed study may help in identifying the bioactive compound of industrial importance.

Keywords: Wild edible fruits, ethnobotanical, antibacterial activity

*Correspondence: pranabdutta74@gmail.com

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INTRODUCTION

The North-Eastern States of India is rich of natural plant products which have tremendous ethnobotanical value but unexplored. Searching for alternatives of antibacterial synthetic drugs is a paramount area of research. The wild eatable fruits have incredible potential for creating new medications to human well-being. There are numerous wild fruits utilized in medicine today, which initially got from plant sources. A large portion of medications' inception from plants of a restorative worth offers new wellsprings of drugs, which have been utilized successfully in daily medicine.^{1,2} Plant origin antibacterial property has massive remedial possibilities and is viable in treating irresistible ailments than engineered antimicrobials, which have antagonistic impacts.^{3,4} Examination laborers announced the elite of dynamic plants having 157 families⁵ and the antimicrobial movement.⁶⁻¹⁰ The issue of microbial obstruction is developing, and the stand point for the utilization of antimicrobial drugs, later on is unsure. Thus, documentation of ethnobotanical value of the plant products and investigation on their antibacterial properties could be beneficial to the society. Tripura, a North-Eastern part of India, is mostly hilly with dense forest. Among the eight (8) districts, four (4) including Khowai, Dhalai, West Tripura and Sipahijala districts were selected for sample collection sites (Fig. 1) as this areas are rich in wild fruit plants which have been used

by the tribal people to meet their nutrition and medicines. The huge knowledge on the prevailing wild fruits helped us to choose the different fruits for the present study.

Recently, we have documented the ethnobotanical value of some fruits.¹¹ Nevertheless, the restorative estimation of wild edible fruits lies in their ingredients like alkaloids, flavonoids, tannins and phenolic derivatives, and that produce a specific physiological activity on the human body.¹² Polyphenols from the wild fruits have a few advantages, including antimicrobial properties against pathogenic and waste microorganisms and varieties in these mixes' concoction creations to prompt marked contrasts in their antimicrobial action.¹³⁻¹⁵ In this paper we have reported the antibacterial properties of the nine (9) underutilized wild fruits viz., *G. gummitutta*, *P. serratum*, *S. suaveolens*, *F. jangomas*, *S. assamicum*, *S. dulcis*, *A. lacucha*, *F. hispida*, and *C. macroptera* against 5 human pathogens viz., *E. coli*, *P. aeruginosa*, *S. typhi*, *S. aureus* and *K. pneumonia*. Additionally, the ethnobotanical value of the four (4) unexplored fruits: *F. jangomas*, *S. dulcis*, *A. lacucha* and *F. hispida* has also been documented.

MATERIALS AND METHODS

Sample collection

Field visits were made to conduct surveys to different villages and forest areas of four (4)



Fig. 1. Sample collection site (encircled with green colour) of Tripura, India.¹¹

Table 1. List of wild edible minor fruits of Tripura, India considered for the study

No.	Botanical name *(IPNI)& family name	Voucher specimen number	Local name	Time of availability	Taste	Uses: folk medicines	Ref.
1	<i>Garcinia gummi-gutta</i> (L.) N. Robson; Clusiaceae	SB-28, 23.06.2015, Tablabari, Khowai	B- <i>Kowagota</i> K- <i>Kouk</i> R- <i>Akouk</i>	May - July	Sour and sweet	Raw fruits as juice, pickles, use treatment for Diarrhoea as folk medicine.	[11]
2	<i>Protium serratum</i> Engl.; Burseraceae	SB-21, 12.06.2015, Tablabari, Khowai	B- <i>Niyar</i> K- <i>Thychrm</i> R- <i>Thaichramm</i>	May - July	Sour	Fruits are used for treatment of mouth ulcer, fruits have antioxidant properties	[11]
3	<i>Stixis suaveolens</i> (Roxb.) Pierre.; Capparaceae	SB-22, 4.06.2016, Tablabari, Khowai	B- <i>Madhabi</i> K- <i>Majeelota</i> R- <i>Mooni</i>	May - July	Taste like ripen date palm	Fruits are used as consumption, also use folk medicine for heart disease, asthma.	[11]
4	<i>Flacourtia jangomas</i> (Lour.) Raeusch Salicaceae	SB-64 23.8.2016 Ajagartilla, Khowai	B-Tipafol / Paniol K- Tekroi R- Tekrui	July - Nov	Mild sour and tangy taste	Consumed as raw and cooked, salt-dried, jam, marmalade, leaves and fruits used against diarrhea, Dried leaves effectively for bronchitis.	Present study
5	<i>Syzygium assamicum</i> (Biswas & C. S.Purkay.) Raizada; Myrtaceae	SB-46, 12.07.2016, Tablabari, Khowai	B- <i>Khudikalajam</i> K- <i>Jamuk</i> R- <i>Jamuk</i>	May - Sept	Slightly bitter in taste and sweet	For treating diabetes (seeds)	[11]
6	<i>Spondias dulcis</i> L. Anacardiaceae	SB-66 20.6.2016 Champahar, Khowai	B-Amra K-Thaitwi R- Thaitu	June - Aug	Sour in Taste	Consumed as raw, Prepared pickle, flavoring for sauces, stews, Fruits has been used as diuretic and Febrifuge, bark is astringent, dysentery etc.	Present study
7	<i>Artocarpus lacucha</i> Buch.-Ham Moraceae	SB-54 12.7.2016 Tablabari, North Chebri	B-Dowa K-Dawa R-Dua	May- July	Slightly sweets sour	As raw ripen fruits, seeds consumed by roasted, seeds have medicinal properties	Present study
8	<i>Ficus hispida</i> L. Moraceae	SB-65 12.7.2016 Tulashikhar, Khowai	B- Jagadamru K-Khuichang R-Khoichawng	May- July	Slightly sweet	Consumed as raw, used for Jaundice, edema, anemia, leprosy, ulcers, epitasis.	Present study
9	<i>Citrus macroptera</i> Montrouz.; Rutaceae	SB-34, 10.10.2015, Tulasikhar, Khowai	B-Sathkora K-Chatukora R-Saataokra	Sept- Nov	Sour	Fruits are eaten raw. Peels of fruit used for tenderization of meat, treating gastritis, kidney stone treatment.	[11]

*IPNI: International Plant Names Index, Local names: B-Bangla; K- Kakborak; R- Reang.

districts of Tripura viz., Khowai, Dhalai, West Tripura and Sipahijala during different seasons in Feb-2018 and July-2019 and live specimens were collected (Table 1). One hundred (100) nos fruits per species were collected from 10 different plants. All the plant species selected for the present study were scientifically identified by the taxonomists, Prof. B.K. Datta, Department of Botany, Tripura University, Tripura, India and Dr. A. Kaur, The Energy and Resources Institute, NER Center, Guwahati, Assam, India.

Bacterial strains

Five bacterial strains viz. *Escherichia coli* (Gram-Ve, ATCC-2522), *Pseudomonas aeruginosa* (Gram-Ve, ATCC-27853), *Klebsiella pneumoniae* (Gram-Ve, ATCC-700603), *Salmonella typhi* (Gram -Ve, ATCC-14028), and *Staphylococcus aureus* (Gram +Ve, ATCC-25923) (Plate 1a & b) were collected from the Institute of Jorhat Medical College and Hospital, Jorhat, Assam, India for the present study.

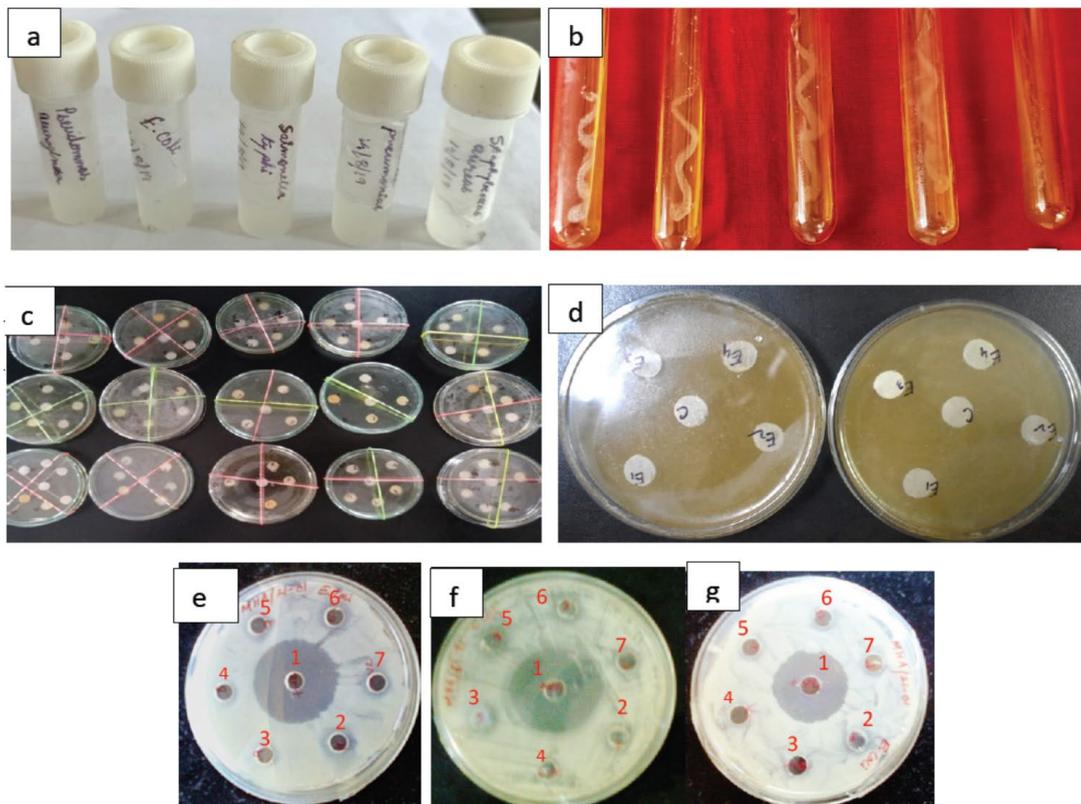


Plate 1(a-i). a. Microorganism in used in the study (L-R) :*Pseudomonas auriginosa* (ATCC27853), *Eschericia coli* (ATCC-2522), *Salmonella typhi* (ATCC-14028), *Klebsiella pneumoniae* (ATCC-700603), *Staphylococcus aureus* (ATCC-25923), b. Pure culture of (L-R): *Pseudomonas auriginosa* (ATCC-27853), *Eschericia coli* (ATCC-2522), *Salmonella typhi* (ATCC-14028), *Klebsiella pneumoniae* (ATCC-700603), *Staphylococcus aureus* (ATCC-25923), c. General view of *in vitro* assay on antibacterial assay of fruit extract against the targeted pathogens, d. Closer view of *in vitro* assay on antibacterial assay of fruit extract against the targeted pathogens, e. Inhibition of growth of *E. coli* by different plant extract (1: *G. gummi-gutta* at 100 mg/ml, 2. *S. suaveolens* at 100 mg/ml, 3. *G. gummi-gutta* at 25 mg/ml, 4. Control, 5. *S. suaveolens* at 25 mg/ml, 6. *G. gummi-gutta* at 50 mg/ml, 7. *G. gummi-gutta* at 50 mg/ml), f. Inhibition of growth of *S. typhi* by different plant extract (1. *G. gummi-gutta* at 100 mg/ml, 2. *S. assamicum* at 100 mg/ml, 3. *G. gummi-gutta* at 75 mg/ml, 4. Control, 5. *S. assamicum* at 25 mg/ml, 6. *A. lackocha* at 75 mg/ml, 7. *A. lackocha* at 50 mg/ml, g. Inhibition of growth of *S. aureus* by different plant extract (1. *G. gummi-gutta* at 100 mg/ml, 2. *S. suaveolens* at 100 mg/ml, 3. *P. serratum* at 100 mg/ml, 4. Control, 5. *S. dulcis* at 100 mg/ml, 6. *F. jangomas* at 100 mg/ml, 7. *F. hispida* at 100 mg/ml

Table 2. Antibacterial activity of ethanol extracts of wild fruits against five human bacterial pathogens

Plant name	Concen. (mg/mL)	Inhibition zone (mm)*				
		<i>E. coli</i>	<i>S. aureus</i>	<i>P. auriginosa</i>	<i>K. pneumoniae</i>	<i>S. typhi</i>
<i>G. gummi-gutta</i>	25	7.0±1.05	5.0±1.12	5.5±0.98	6.5±1.21	4.5±1.59
	50	16.5±1.10	13.5±0.99	13.5±1.92	11.0±1.32	10.0±1.78
	75	23.4±1.23	21.6±0.87	20.5±1.26	19.0±1.44	17.5±1.40
	100	35.0±1.06	32.0±1.12	33.5±1.62	31.5±1.52	32.5±1.38
	25	-	-	4.0±1.71	3.5±1.39	-
<i>P. serratum</i>	50	-	-	7.5±1.29	7.0±1.83	-
	75	-	-	13.0±0.78	12.0±1.52	-
	100	-	-	21.0±1.93	18.0±1.84	-
<i>S. suaveolens</i>	25	0.0±1.00	0.0±1.54	3.0±0.91	0.0±1.96	-
	50	3.0±1.21	3.0±1.62	7.5±0.99	0.0±1.20	-
	75	6.5±1.09	5.0±1.23	15.0±1.74	5.0±1.36	-
	100	10.0±1.21	9.0±1.41	21.0±1.65	9.0±1.94	-
	25	-	0.0±1.22	0.0±1.65	0.0±0.79	-
<i>F. jangomas</i>	50	-	3.0±1.32	4.0±1.72	0.0±0.82	-
	75	-	6.5±1.07	6.5±1/25	4.5±1.64	-
	100	-	11.0±1.17	10.5±1.71	8.5±1.72	-
<i>S. assamicum</i>	25	5.5±0.74	4.0±1.25	-	-	4.5±1.28
	50	9.0±1.72	7.5±1.42	-	-	7.5±1.42
	75	17.5±1.48	13.5±1.52	-	-	12.0±1.63
	100	32.5±1.30	23.0±1.33	-	-	19.0±1.72
	25	-	-	0.0±1.77	-	-
<i>S. dulcis</i>	50	-	-	0.0±1.62	-	-
	75	-	-	3.5±1.52	-	-
	100	-	-	9.0±1.64	-	-
	25	6.5±1.24	4.5±1.71	9.5±1.76	0.0±0.56	7.50±1.94
<i>A. lackocha</i>	50	12.5±1.73	9.5±1.10	15.5±1.82	4.5±1.62	14.0±1.37
	75	19.0±1.86	15.5±1.26	21.0±1.43	9.5±1.80	19.5±0.81
	100	33.5±1.52	24.0±1.62	34.5±1/23	15.0±1.88	32.0±1.82
<i>F. hispida</i>	25	-	-	3.0±1.49	-	-
	50	-	-	6.5±1.71	-	-
	75	-	-	12.5±1.64	-	-
<i>C. macroptera</i>	100	-	-	18.0±1.93	-	-
	25	-	-	-	-	-
	50	-	-	-	-	-
	75	-	-	-	-	-
	100	-	-	-	-	-

*Data are mean of five replicates; '-': No inhibition was observation at the tested concentration.

Preparation of Extracts

The wild fruits samples were collected and washed independently in two fold sterile distilled water and freeze dried for 72 hours, followed by drying in hot air oven (Labotech Solutions, India) at 50 °C consecutively for four days. The dried fruits were ground to a fine powder and stored completely dried in a cleaned dry glass holder and protected from the microbial contaminations. One gram of the powder (fine) was transferred

into each of the four sterile Falcon tubes (cap. 15 mL) and added 10 mL of sterile water and ethanol(absolute) respectively. Extraction was prepared by continuous vigorous shaking and sonicating (4X15 min in a 24 h period) in sonicator water bath. On fourth day, tubes were centrifuged for 15 min at 2058 g and the supernatant was transferred to 1 mL aliquots to various Eppendorf containers of limit 1.5 mL. After the evaporation of the solvent, the dried residues of 1mL extract

were re-dissolved in 200 μ L DMSO for the ethanol extracts. The sediments were refrigerated at 4°C until further use.

Preparation of inoculum

Inoculum of the bacterial strains were prepared by moving a loopful of cells from the way of life to test containers of Mueller-Hinton Broth (MHB) and were brooded without disturbance for 24 hrs at 37°C. The way of life was additionally diluted with new MHB to get optical densities of 2×10^6 cfu/mL.

Antibacterial activity of fruits extract

The antibacterial activity of each fruit sample was determined using the disk diffusion method with need based modification.¹⁶ The disk diffusion method was performed using Mueller-Hinton Agar (MHA). The ethanol extracted (100 mg) was fragmented up in 2 mL of dimethyl sulfoxide and sanitized through a film channel (0.22 μ m) and tested at four (4) concentrations. viz., 100, 75, 50, and 25 mg/mL by pouring 5 μ L over the 6-mm distance across of disinfected channel paper plates. At that point, 10-15 mL of agar medium was filled disinfected Petri-dishes and represented setting for 15 minutes, followed by the expansion of 100 μ L of bacterial culture on the plate's head, spread with sterile L-circle and permitted to dry for few moments. Finally, the sterile channel paper circle stacked with various groupings of organic products removed was set on the head of MHA plates and represented brooding for 24 hours. The antibacterial action was assessed by estimating the width of the inhibitory zone for the test microorganisms. The inhibition zone developed due to the action of extract were measured by subtracting disk diameter (6-mm). The potential extracts were chosen based on the highest inhibition zone developed.

Determination of minimum inhibitory concentration (MIC)

MIC of the fruits extracts was measured by following the tube dilution technique.¹⁷ Two-fold dilution of the fruits extracts (25, 50, 75 and 100 mg/mL) were set up in tubes containing nutrient broth. Equal volume of extract and nutrient broth were mixed in each test tube. After that, 0.1 mL of adjusted inoculum of the bacteria (1×10^8 cfu/mL) was added to each tube. All plates were incubated for 24 hours at 37°C. Two control tubes were maintained, one for

positive control containing anti-toxins, and the other was a negative control containing bacterial inoculum, physiological saline and stock medium. Antibiotic disk of ciprofloxacin (1 μ g) was used for *E. coli*, *P. aeruginosa*, *K. pneumoniae*, *S. aureus* and azithromycin for *S. typhi* as reference antibiotic. Turbidity was adjusted to 0.5 McFarland standard and speeded on plates.

Determination of minimum bactericidal concentration (MBC)

MBC was determined by sub-culturing the MIC test dilutions followed by Abdullah and Elsharkawy.¹⁷ For this 50 μ L of incubated test tubes from the MIC, an assay was pipette carefully and transferred to sterile nutrient agar plates. Plates were then incubated overnight at 37 °C and were observed for any visible growth. The lowest MIC concentration that exhibited no visible bacterial growth on the agar plate was taken as MBC.

Statistical analysis

The results obtained from the experiments of the present study were expressed as mean \pm SD. Data recorded from dose-response experiments were represented as the percentage of inhibition and analyzed with Prism TM (Graph Pad Prism 5.0 Software Inc; San Diego, CA). The MIC₅₀ for each growth condition was estimated by fitting the data to a non-linear least- squares sigmoid regression curve.

RESULTS

Ethnobotanical documentation

Ethnobotanical uses of nine (9) plant species belonging to eight (8) families (Rutaceae, Burseraceae, Capparaceae, Myrtaceae, Cluceae, Salicaceae, Moraceae, Anacardiaceae) were recorded from tribal areas of Tripura (Table 1). Among the nine (9) wild fruits species, five (5) fruits, viz., *G. gummi-gutta*, *P. serratum*, *S. assamicum* and *C. macroptera* were reported in our earlier investigation¹¹ and the four (4) fruits: *F. jangomas*, *S. dulcis*, *A. lacucha*, and *F. hispida* are documented in the present study (Table 1). The table provides information on the botanical name, family, local name (Bangali, Kokbork, Reang), time of availability, taste and use in folk medicine. Tribal people use these wild fruit plants for a different therapeutic purpose like treating kidney stones, antioxidant property, heart diseases, asthma, diabetes, diarrhea, bronchitis, jaundice, edema,

Table 3. MIC and MBC value of selected fruits extracts against five bacterial strains

Plant name	Concen. (mg/ml)	Bacterial pathogens									
		<i>E. coli</i>		<i>S. aureus</i>		<i>P. aeruginosa</i>		<i>K. pneumoniae</i>		<i>S. typhi</i>	
		MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC
<i>G. gummi-gutta</i>	25	+	+	+	+	+	+	+	+	+	+
	50	-	+	-	+	-	+	-	+	-	+
	75	-	-	-	-	-	-	-	-	-	-
	100	-	-	-	-	-	-	-	-	-	-
<i>P. serratum</i>	25	-	-	-	-	+	+	+	+	-	-
	50	-	-	-	-	+	+	+	+	-	-
	75	-	-	-	-	-	-	-	+	-	-
	100	-	-	-	-	-	-	-	-	-	-
<i>S. suaveolens</i>	25	+	+	+	+	+	+	+	+	-	-
	50	+	+	+	+	-	-	+	+	-	-
	75	-	+	+	+	-	-	-	+	-	-
	100	-	-	-	-	-	-	-	-	-	-
<i>F. jangomas</i>	25	-	-	+	+	+	+	+	+	-	-
	50	-	-	+	+	+	+	+	+	-	-
	75	-	-	-	+	-	+	-	+	-	-
	100	-	-	-	-	-	-	-	-	-	-
<i>S. assamicum</i>	25	-	-	-	+	-	-	-	-	-	+
	50	-	-	-	-	-	-	-	-	-	-
	75	-	-	-	-	-	-	-	-	-	-
	100	-	-	-	-	-	-	-	-	-	-
<i>S. dulcis</i>	25	-	-	-	-	+	+	-	-	-	-
	50	-	-	-	-	+	+	-	-	-	-
	75	-	-	-	-	+	+	-	-	-	-
	100	-	-	-	-	-	-	-	-	-	-
<i>A. lackocha</i>	25	-	-	-	+	-	-	+	+	-	-
	50	-	-	-	-	-	-	-	+	-	-
	75	-	-	-	-	-	-	-	-	-	-
	100	-	-	-	-	-	-	-	-	-	-
<i>F. hispida</i>	25	-	-	-	-	-	+	-	-	-	-
	50	-	-	-	-	-	-	-	-	-	-
	75	-	-	-	-	-	-	-	-	-	-
	100	-	-	-	-	-	-	-	-	-	-

Red color means: MIC was MBC were not performed as these concentrations do not have any inhibitory effect (Please refer Table 2)
 Yellow color means; MIC and MBC were analyzed as at these concentrations the botanicals showed inhibitory effect (Please refer Table 2).

anemia, leprosy, ulcers, epitis, dysentery, etc. All these plants and fruits are found to have high popularity among the tribes due to their value added efficiency and awareness by the native tribes for their everyday use as an herbal medicine. But the ethnobotanical data on the bio-prospecting of traditional plants for medicinal uses is always a controversy owing to a different report by different workers. Soejarto et al.¹⁸ reported that less than 15% of plant species traditionally used

for medicinal purpose is subjected to clinical trials. Still, the advantages of lying in plant selection based on traditional knowledge can't be ignored. This is because it guides us to target the parts to be used for testing against a disease, avoiding wastage of time and resources in unnecessary testing of inactive plant parts. It also guides us in the steps of processing the materials and the route of administration. Research on natural resources under ethnobotanical guidance could generate

more information for the more significant benefit of human beings and the associated ecosystem. But, the traditional knowledge and practice of apply traditional medicine are disappearing slowly that call the urged need of ethnobotanists to create mass awareness and capacity building for the conservation of the traditional plant species. Saslis-Lagoudakis et al.¹⁹ reported that conventional knowledge that enhances health throughout the world is crucial consideration in preventing worsening health. New research initiatives demonstrate the revival of interest in the subject.

Antibacterial activity

Extract prepared from the wild fruit was tested for antibacterial activity in dilution broth assay against the five bacterial microorganisms, viz., *E. coli*, *P. aeruginosa*, *K. pneumoniae*, *S. typhi*, and *S. aureus* (Plate 1 c-g). The antibacterial assay was done by disk diffusion technique at four (4) concentrations viz., 100, 75, 50 and 25 mg/mL. All the wild fruits extract except *C. macroptera*, showed antibacterial activity against the bacterial strains and with highest zone of inhibition (35.0 mm) against *E. coli* at 100 mg/ml concentration of ethahnol extract of *G. gummi-gutta* (Table 2). This was followed by a zone of inhibition of 34.50 mm by ethanol extract of *A. lakoocha* against *P. auriginosa*. Interestingly, out of all the fruit extracts only *G. gummi-gutta* and *A. lakoocha* showed their antibacterial activities against all the bacterial pathogen under this study. Therefore, these plants may be suitable candidates for discovering board-spectrum antibiotics against the tested bacteria. *G. gummi-gutta* causes zone inhibition of 33.5 mm, 32.5 mm, 32.00 and 31.5 mm against *P. auriginosa*, *S. typhi*, *S. aureus* and *K. pneumonia* respectively at 100 mg/ml of concentration and with decrease of concentration the zone of inhibitions was found to decreases. In case of fruit extract of *A. lakoocha*, the zone of inhibition of 33.5 mm, 32.0 mm, 24.0 mm and 15.0 mm was recorded for *E. coli*, *S. typhi*, *S. aureus* and *K. pneumonia* respectively at 100 mg/ml of concentration. For the fruit extract of *P. serratum* no antibacterial activity was observed against *E. coli*, *S. aureus*, and *S. typhi*. Similarly, no antibacterial activity of *S. suaveolens* was observed against *S. typhi*. On the other hand *S. dulcis* and *F. hispida* showed antibacterial activity against only

P. auriginosa. Similarly, *F. jangomas* was found to have antibacterial activity against *S. aureus*, *P. auriginosa* and *K. pneumonia* with varying degree of zone of inhibition at different concentration but no inhibitory effect of was found against *E. coli* and *S. typhi*. Fruit extracts of *S. suaveolens* and *F. jangomas* at concentration of 25 and 50 mg/ml could not able to inhibit the growth of *K. pnemonie* but at higher concentration i.e., 50 and 100 mg/ml it could able to inhibit the growth significantly. Several scientists have reported the composition, antimicrobial, antifungal and antioxidant activities of essential oils, especially from different fruits of the citrus genus.²⁰⁻²² It has been recognized for centuries that essential oils possessed antimicrobial property. Citrus oils have the GRAS status for food and drug applications possessing the inhibitory effects against a range of both gram positive and gram negative bacteria both as in the form of direct oil and in vapor form.²³ Antimicrobial activities of both the fixed and essential oils were also evaluated earlier against different pathogenic bacterial strains following disk diffusion method.

The zones of inhibition of microbial growth are expressed a function of antimicrobial activity of the extracts.²⁴ Data presented in Table 3 showed that *S. suaveolen* may be a potential source of drug to fight against all the tested organism in the study except *S. typhi*. Earlier, Saklani et al.²⁵ studied the anti-bacterial activity of fruit extract of *Rubus ellipticus* and reported significant inhibitory effect against *E. coli*, which is practically identical to the current investigations. Correspondingly, Babahmad et al.²⁶ revealed the impact of inhibitory action of numerous botanicals against gram-positive and gram-negative microscopic organisms with shifting degrees of hindrance zone, which are similar with the result of the current examination. In inhibition zone of 5 ± 1.10 to 20 ± 0.31 mm was reported by Qadir et al.²⁷ for seed extract of *Anamirta cocculus* against *P. vulgaris*, *S. aureus*, *E. coli*, *K. pneumonia* and *S. typhi*.

Eight plant removes at four novel centers (25, 50, 75 and 100 mg/mL) were attempted against five particular bacterial microorganisms and results are summarized in Table 3. The MIC and MBC assessments of bacterial strains were found to the extent of 50 - 100 mg/mL and 75-100 mg/mL respectively. The MIC and MBC value

for *G. gummi-gutta* was recorded to the extent of 75-100 mg/mL, against all the five bacterial strains. However, the value was 100 mg/mL for *S. suaveolens* against *E. coli*, *S. aureus*, *P. aeruginosa* and *K. pneumoniae*. Lowest MIC and MBC values of 25 mg/mL were recorded for *S. assamicum*, against *E. coli* and *A. lacucha* against *E. coli*, *S. aureus*, *P. aeruginosa*, and *S. typhi*.

Most of the test extracts inhibited the growth of the pathogens and the present studies are in agreement with the earlier worker^{28,29} who reported greater antibacterial activity of Indian Medicinal plants on Gram positive and Gram negative bacteria. But the efficacy was more in Gram positive bacteria than the Gram negative bacteria, which may be due to the nature of cell structure.³⁰ Kitzberger et al.³¹ defined MIC as inhibition of microbial growth after 24 hours of incubation when exposed to the lowest concentration of the plant extract and the minimum amount of plant extract that completely kill the respective microorganism after 24 hours of incubation on the freshly inoculated agar plates is called MBC. The lower extent of MIC and MBC regard suggests a different degree of antimicrobial development. The MIC and MBC value recorded in the current examination is higher than the assessment of wild berry natural item species, which was reported by Radovanovic et al.³² who got the value to the extent of 15.6 to 500 µg/ml against *S. aureus*. In another assessment by Islary et al.³³ reported an MBC assessment of 15 mg/mL for fruit extract of *Aporosa dioica* against *S. aureus* and *P. vulgaris* and MBC assessment of < 30 mg/mL was recorded for fruit extract of *Antidesma bunius* against all the four targeted strains.

DISCUSSION

In this research, out of nine (9) distinctive wild edible fruits species, two (2) viz., *Garcinia gummi-gutta* (L.) N. Robson; and *Artocarpus lacucha* Buch.-Ham, were discovered to be successful in stifling the development of the apparent multitude of bacterial microorganisms viz., *E. coli*, *S. aureus*, *P. aeruginosa*, *K. pneumonia* and *S. typhi* at a convergence of 75 - 100 mg/mL. We found the most minimal MIC and MBC estimations of 25 mg/ml were recorded for *Syzygium assamicum* (Biswas & C. S. Purkay.) Raizada; against *E. coli* and *Artocarpus lacucha*

Buch.-Ham against *E. coli*, *S. aureus*, *P. aeruginosa*, and *S. typhi* showing the concrete antibacterial exercises. Our exercise has detailed four wild, edible fruits their society medication utilizes where included *Flacourtia jangomas* (Lour.) Raeusch, *Spondias dulcis* L., *Artocarpus lacucha* Buch.-Ham and *Ficus hispida* L., and five wild fruits sample have been reported ethnobotanical information in our past investigation. From the current examination, antibacterial exercises give a premise to tranquilize revelation from the indigenous wild eatable fruits devoured by the ancestral network of the North-Eastern province of India where especially, Tripura. Further exploration of the phytochemistry and pharmacological property of the wild consumable organic products must be done; investigating bioactive mixes and their activity method with the target site should also be examined.

CONCLUSION

The underutilized wild fruits are not used for commercial purpose due to lack of proper scientific study and documentation, although have high nutritional values. In light of this investigation, it is evident that further studies to find new, unconventional underutilized fruits from the wild is an urgent necessity. Which will enrich the fruits basket of the different states of India and it is hoped that popularization of these fruits for the benefit of the rest of the communities will gain momentum. We believe that our findings will open a new chapter for commercialization of those wild minor fruits for both cultivation and the mass uses to all communities. Further, this study paves the way of using the underutilized wild fruits of North Eastern region of India as a potential source of antipathogenic activity of human pathogen. Further details study may help in identifying the bioactive compound of industrial importance.

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

AUTHORS' CONTRIBUTION

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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DATA AVAILABILITY

All datasets generated or analyzed during this study are included in the manuscript

ETHICS STATEMENT

Not applicable.

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