

Prevalence of *MecC* Gene Among Methicillin Resistant *Staphylococcus aureus* isolated from Patients in Ain-Shams University Hospital

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Abstract

Methicillin-Resistant *Staphylococcus aureus* (MRSA) is an important cause of healthcare associated infections globally. New *mecA* homologue (*mecC*), was first reported in the UK and Denmark. The *mecC* mediated MRSA is resistant only to B-lactams antibiotics and is sensitive to other antibiotics. Detecting the prevalence of *mecC* MRSA provides more options in treatment of MRSA infections. The aim of this study was to prevalence of *mecC* gene in clinical isolates of MRSA in Ain-Shams university hospitals & to correlate Minimal Inhibitory concentration (MIC) of Oxacillin with the *mecC* gene expression in MRSA isolates. Fifty MRSA isolates were collected from different intensive care units (ICUs) of Ain-Shams university hospital from April-December 2018. Methicillin resistance was detected by Cefoxitin disc, and antimicrobial susceptibility testing was done for all isolates and its results were interpreted according to Clinical & Laboratory Standards Institute (CLSI) guidelines 2018. Minimal Inhibitory Concentration of Oxacillin was detected using Oxacillin E-test and the results were interpreted according to the manufacturer's instructions, then Polymerase Chain Reaction was done to detect *mecA* and *mecC* genes among MRSA isolates. Fifty isolates were identified as MRSA by Cefoxitin disc out of 163 samples. Twelve isolates were sensitive to Oxacillin while 38 isolates were resistant to Oxacillin. All isolates were positive to *mecA* gene while only 3 isolates were positive to both *mecA* and *mecC* genes. *MecC* is a new emerging gene responsible for methicillin resistance in *staphylococci* and was detected in 6 % of the isolates in this study.

Keywords: MRSA, *mecC* gene, *mecA* gene, PCR

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INTRODUCTION

Methicillin-Resistant *Staphylococcus aureus* (MRSA) is considered as one of the most common causes of community and hospital-acquired infections, leading to high morbidity and mortality. MRSA poses a serious problem in hospitals and its detection is crucial for infection prevention and control¹.

Resistance to beta-lactams antibiotics is conferred by the presence of *mecA* gene, which encodes a penicillin-binding protein (PBP2'). *mecA* is part of a mobile genetic element called the "Staphylococcal cassette chromosome (SCC) *mec*."²

The report of MRSA carrying a new variant of the *mecA* gene in 2011 in humans was highly significant. This new variant was named *mecC* gene. The presence of this gene poses diagnostic problems due to the probability of misdiagnosis of isolates as methicillin-sensitive *S. aureus* which may affect the results of MRSA surveillance³.

The *mecC* gene shared only 70% DNA identity with the *mecA* gene. MRSA isolates carrying *mecC* gene have been reported in different European countries and from several host species⁴.

This *mecC* MRSA shows a characteristic antimicrobial susceptibility pattern compared to *mecA* MRSA. Where *mecA* MRSA displays resistance to both Oxacillin and Cefoxitin while most of *mecC* MRSA shows resistance to Cefoxitin and susceptibility to Oxacillin⁵.

There is limited available data about the prevalence of MRSA that carry *mecC* gene in Egypt, hence, this study was performed.

METHODOLOGY

Collection of clinical samples

Different clinical samples were obtained from patients admitted to different ICUs of Ain-Shams university hospitals according to the regulations of scientific research Ethical Committee Faculty of Medicine-Ain Shams University during the period from April 2018 to December 2018. The collected specimens include blood, pus, sputum and swabs from burn and surgical wound.

Identification and antimicrobial susceptibility testing of the isolated organism

Sample collection and identification of the isolated organism were performed by

conventional bacteriological methods according to Colle et al, 1996⁶ and Cheesbrough, 2009⁷.

For identification of *Staphylococcus aureus* isolates, the samples were inoculated on Blood agar and Mannitol salt agar, the colonies were identified based on morphology of the isolated colonies, microscopic examination of Gram-stained films, the β -hemolytic effect on blood agar and yellow colonies on mannitol Salt agar.

Then detection of MRSA strains was done by Cefoxitin discs 30 μ g (FOX) supplied by (Oxoid, Basingstoke, UK) using Kirby-Bauer disc diffusion method and the results were interpreted according to CLSI guidelines 2018⁸.

Antimicrobial susceptibility testing for the following antibiotics was done using Muller Hinton agar: Ciprofloxacin 5 μ g (CIP), Cefoxitin 30 μ g (FOX), Chloramphenicol 30 μ g (C), Penicillin 0.6 μ g (P), Linezolid 10 μ g (LZO), Rifampicin 5 μ g (RA), Doxycycline 30 μ g (DO), Erythromycin 15 μ g (E), Clindamycin 2 μ g (DA), Levofloxacin 5 μ g (LEV), Gentamycin 10 μ g (CN), Trimethoprim/ sulfamethoxazole 1.25 + 23.75 μ g (SXT), all antibiotic discs were supplied by (Oxoid, Basingstoke, UK), and the results were interpreted according to CLSI 2018⁸.

Detection of oxacillin minimal inhibitory concentration (MIC)

MIC was done for the Fifty MRSA isolates by using Oxacillin E-test supplied by LIOFILCHEM company, and the interpretation was done according to CLSI guidelines, 2018.

Detection of *mecA* gene and *mecC* gene by Polymerase chain reaction (PCR)

All MRSA strains were subjected to PCR for detection of *mecA* and *mecC* gene, first DNA extraction was done using ultraclean microbial DNA extraction kit supplied by MoBio by QIAGEN, then amplification of *mecA* gene⁹ was performed using the primers forward 5'-AAA ATC GAT GGT AAA GGT TGGC-3' and reverse 5'-AGT TCT GCA GTA CCG GAT TTG C-3' and for *mecC* gene⁴ forward 5'-GAA AAA AAG GCT TAG AAC GCC TC-3' reverse 5' GAA GAT CTT TTC CGT TTT CAG C-3'.

PCR conditions for *mecA* gene⁹

Thermal cycler was adjusted to 2 minutes at 95°C for primary denaturation followed by 40 cycles of 45 seconds denaturation at 95°C then 45 seconds at 50°C for annealing, and 1 min at 72°C

for extension. The amplified product was identified at 530 base pair (bp) by electrophoresis on 1.5% agarose gels and using 100 bp DNA ladder.

PCR conditions for *mecC* gene⁴

Thermal cycler was adjusted to 2 minutes at 95°C for primary denaturation followed by 40 cycles of 45 seconds denaturation at 95°C then 45 seconds at 60°C for annealing, and 1 min at 72°C for extension. The amplified product was identified at 137 bp by electrophoresis on 1.5% agarose gels and using 100 bp DNA ladder (Fig. 1 & 2).

Data analysis

All statistical procedures were carried out using SPSS version 22.0 for Windows (SPSS Inc, Chicago, IL, USA).

RESULTS

In this work, different clinical samples were collected from 163 patients admitted to different ICUs. From these samples, fifty isolates were identified as MRSA in which 25 isolates were collected from males (50%) and 25 from females (50%). Their age ranged from 18-75, and the mean age was 49.6 ± 13.1 years and all patients were under antibiotic therapy.

Most of MRSA isolates were obtained from blood samples (25 isolates (50%)) followed by 9 (18%) sputum and 9 (18%) swabs from surgical and burn wounds and 7(14%) pus samples.

The results of antimicrobial susceptibility of MRSA revealed that the highest sensitivity values were found with Trimethoprim/ sulfamethoxazole

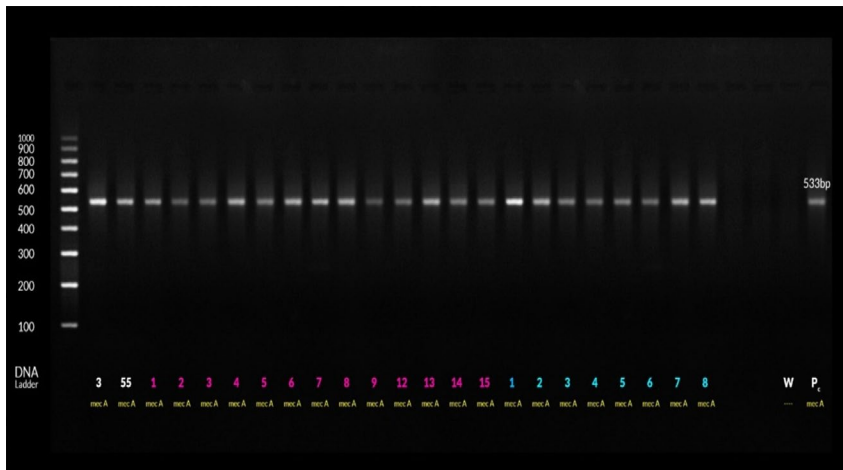


Fig. 1. Gel electrophoresis of *mecA* gene amplification shows *mecA* bands at 533bp

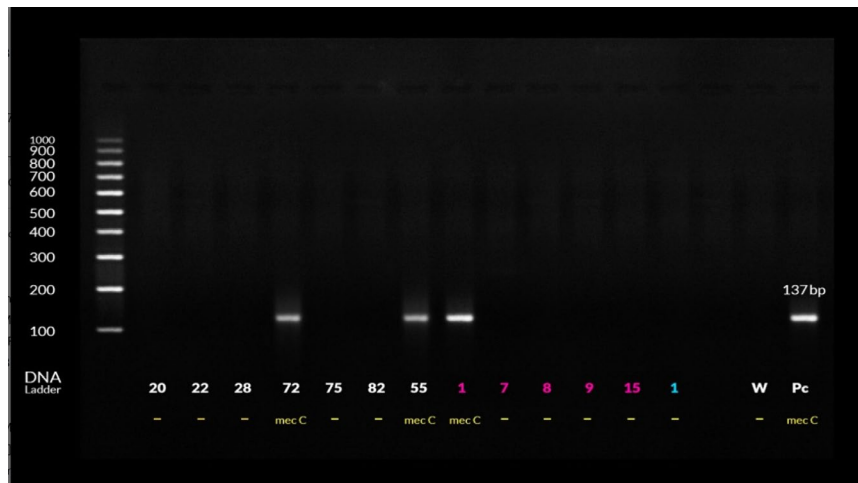


Fig. 2. Gel electrophoresis of *mecC* gene amplification shows *mecC* bands at 137bp

(SXT), linezolid (LZD), and Chloramphenicol (C) (68% for each antimicrobial agent) while lower sensitivity (66%) was found with Rifampin (RA) followed by Levofloxacin (LEV) (62%). Sensitivity values dropped to 38% with Clindamycin (DA) followed by Doxycycline (DO) and Ciprofloxacin (CIP) with sensitivity in 36% of the isolates. The highest resistance pattern was to Erythromycin (E) and Penicillin (P) as 41 isolates (82%) were resistant to erythromycin and 48 isolates (98%) were resistant to Penicillin (P) as shown in Table (1).

Minimal inhibitory concentration (MIC) of MRSA isolates to Oxacillin was done by using E Test and 12 isolates (24%) were sensitive to Oxacillin while there were 38 isolates (76%) resistant to Oxacillin.

As regards PCR results of determining the presence of *mecA* and *mecC* genes, all isolates (100%) were positive to *mecA* gene while only 3 isolates (6%) were positive to both *mecA* and *mecC* genes and the isolates that carrying both genes were isolated from blood samples

The three isolates carrying both *mecA* and *mecC*, two of them were resistant to Oxacillin E-test and one was sensitive, while 47 MRSA isolates carrying only *mecA* gene, 11 (92%) were sensitive to Oxacillin E-test and 36 (95%) were resistant to Oxacillin E-test.

The results of antimicrobial susceptibility of MRSA carrying both *mecA* and *mecC* genes

revealed that the highest sensitivity values were found with (CIP), (SXT), (LZD) and (C) agents (66.7% for each antimicrobial agent). Lower sensitivity was found with (LEV), (RA) and (DA) which was 33.3% for each. There was no sensitivity towards other antimicrobial agents as shown in Table (2).

DISCUSSION

MRSA infections are well established in both the healthcare setting and in the community. *MecC* is a new emerging gene responsible for methicillin resistance in *staphylococci*. The *mecC* MRSA produces a characteristic antimicrobial susceptibility pattern different from *mecA* MRSA as *mecA* MRSA typically displays resistance to both oxacillin and ceftioxin antibiotic. On the other hand, most of *mecC* MRSA show resistance to Cefoxitin and are therefore reported as MRSA, although it shows susceptibility to Oxacillin ⁵.

In this study, the highest susceptibility of MRSA strains was found to Trimethoprim/Sulfamethoxazole, Linezolid and Chloramphenicol antibiotic (68% for each antimicrobial agent) While lower susceptibility was found with Clindamycin (38%) followed by Doxycycline and Ciprofloxacin (36% for each). The highest resistance pattern was to Erythromycin (82%) and penicillin (98%). This goes in accordance with a study done in Egypt by Abdel-Maksoud et al. in 2016¹⁰ who reported that 100% of all tested isolates were resistant to Penicillin and 70.5 % were resistant

Table 1. Frequencies (n) and percentages (%) for antimicrobial susceptibility of MRSA arranged from highest to lowest sensitivity

| Antimicrobial agent | Sensitive | | Intermediate | | Resistant | |
|-----------------------------------|-----------|----|--------------|----|-----------|-----|
| | n | % | N | % | N | % |
| Trimethoprim/ sulfamethoxazole | 34 | 68 | 6 | 12 | 10 | 20 |
| linezolid | 34 | 68 | 1 | 2 | 15 | 30 |
| Chloramphenicol | 34 | 68 | 1 | 2 | 15 | 30 |
| Rifampin | 33 | 66 | 2 | 4 | 15 | 30 |
| Levofloxacin | 31 | 62 | 1 | 2 | 18 | 36 |
| Clindamycin | 19 | 38 | 15 | 30 | 16 | 32 |
| Doxycycline | 18 | 36 | 6 | 12 | 26 | 52 |
| Ciprofloxacin | 18 | 36 | 17 | 34 | 15 | 30 |
| Gentamycin | 16 | 32 | 4 | 8 | 30 | 60 |
| Erythromycin | 9 | 18 | 14 | 28 | 27 | 54 |
| Penicillin | 2 | 4 | 0 | 0 | 48 | 96 |
| Cefoxitin | 0 | 0 | 0 | 0 | 50 | 100 |

Table 2. Frequencies (n) and percentages (%) for antimicrobial susceptibility of MRSA in *mecC* positive cases (n = 3) arranged from highest to lowest sensitivity

| Antimicrobial agent | Sensitive | | Intermediate | | Resistant | |
|-----------------------------------|-----------|------|--------------|------|-----------|------|
| | n | % | n | % | n | % |
| Ciprofloxacin | 2 | 66.7 | 0 | 0 | 1 | 33.3 |
| Trimethoprim/ sulfamethoxazole | 2 | 66.7 | 0 | 0 | 1 | 33.3 |
| linezolid | 2 | 66.7 | 0 | 0 | 1 | 33.3 |
| Chloramphenicol | 2 | 66.7 | 0 | 0 | 1 | 33.3 |
| Levofloxacin | 1 | 33.3 | 0 | 0 | 2 | 66.7 |
| Rifampin | 1 | 33.3 | 0 | 0 | 2 | 66.7 |
| Clindamycin | 1 | 33.3 | 0 | 0 | 2 | 66.7 |
| Erythromycin | 0 | 0 | 1 | 33.3 | 2 | 66.7 |
| Deoxycycline | 0 | 0 | 0 | 0 | 3 | 100 |
| Penicillin | 0 | 0 | 0 | 0 | 3 | 100 |
| Cefoxitin | 0 | 0 | 0 | 0 | 3 | 100 |
| Gentamycin | 0 | 0 | 0 | 0 | 3 | 100 |

to Ciprofloxacin and 64.4% were resistant to Erythromycin and highest susceptibility was to Trimethoprim/ Sulfamethoxazole, and Rifampicin.

Also, Al-Zoubi et al in 2017¹¹, reported that the highest susceptibility of MRSA isolates was found to Linezolid (96.5%) and Chloramphenicol (86.7 %) and medium sensitivity percentages were found to Clindamycin (54.9%), Gentamicin (47%) and Levofloxacin (42.5%) and highest resistance was to Erythromycin and Penicillin.

Regarding the minimal inhibitory concentration (MIC) using E-test, we found that 12 (24%) isolates were sensitive to Oxacillin and there were 38 (76%) isolates resistant to Oxacillin .on the other hand Saeed et al.in 2014¹² reported (1.2%) strains were sensitive to oxacillin using phenotypic methods in the United Kingdom and Cikman et al. in 2019¹³ reported that all tested MRSA isolates were phenotypically resistant to oxacillin and Cefoxitin.

In the current study results of PCR showed that all cases (100%) were positive to *mecA* gene while only three cases (6%) were positive to both *mecA* and *mecC* gene, one of the *mecC* cases was sensitive to Oxacillin E-test with MIC \leq 2 and two *mecC* were resistant to Oxacillin E-test with MIC \geq 4.

One of the first papers published to discuss the emergence of *mecC* gene in MRSA isolates versus *mecA* gene was conducted in

Denmark 2011 by Garcia-Alvarez⁴ who found that *mecC* gene represents 2.8% of MRSA isolates while in the current study we found 6% *mecC* gene of MRSA isolates, the author of the previous study contributed this result to rural areas as the detection of *mecC* gene was linked to livestock. On the other hand, Rania et al. in 2017¹⁴ in Egypt conducted a similar study showed that of 150 isolates (110 MRSA + 40 MR-CoNS) all were subjected to PCR where no MRSA isolates carrying *mecC* gene were reported detected. This was similar to Ganesan¹⁵ who also did not detect *mecC* gene in his study. Another study done in Egypt by Khairalla et al. in 2017¹⁶ reported that all the MRSA isolates included in the study contain the *mecA* gene ($n = 34$, 100%), while *mecC* was not identified.

On the other hand Khan and co-workers¹⁷ reported that the prevalence of *mecA* gene was 54% which is less than reported in this study, they also found 3% *mecA* negative isolates carrying *mecC*, while only one MRSA isolate carrying both *mecA* and *mecC* genes. also, Aklilu and Hui Ying¹⁸ reported first *mecC* and *mecA* positive livestock-associated MRSA in Malaysia, they concluded that out of the 15 *mecC* positive isolates, 12 were positive for both *mecA* and *mecC* genes this concedes with results of the current study concerning the 3 *mecC* positive were *mecA* positive, so we can assume that we can find both

mecA and *mecC* genes in same isolate which needs further studies for confirmation of this finding .

CONCLUSION

In conclusion, this work is one of first report of presence of *mecC* gene in MRSA isolates in Egypt, *mecC* gene was detected in 6% of MRSA isolates included in this study and this isolates also carry *mecA* gene, although this percentage is not alarming, further studies testing for *mecC* gene in larger scale using different primer sets for *mecC*, may increase the probability to detect *mecC* gene especially in rural areas as the presence of *mecC* is linked to livestock.

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

AUTHORS' CONTRIBUTION

All authors designed the experiments. HR performed the experiments. HR and SA drafted the manuscript, compiled information from the literature, and designed the figures and tables.

WK and AN supervised and reviewed the manuscript. All authors read and approved the manuscript for publication.

FUNDING

None.

DATA AVAILABILITY

All datasets generated or analyzed during this study are included in the manuscript.

ETHICS STATEMENT

This article does not contain any studies with human participants or animals performed by any of the authors.

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