

RESEARCH ARTICLE

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## Temperature Tolerant *Rhizobium leguminosarum* bv. *viciae* Strains with Plant Growth Promotion Traits

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### ABSTRACT

Rhizobacteria (PGPR) that promote the plant growth are essential component of sustainable agriculture. Pea (*Pisum sativum* L.) root nodule *Rhizobium leguminosarum* bv. *viciae* ten strains were cultured at two different temperatures (28°C and 45°C). Out of eight strains screened the three N25, N30 and N40 were temperature tolerant while only one strain (N40) showed tolerance to pH11. The growth of *Rhizobium* strain N40 at 45 °C was 96.8 percent as compared to the growth of the at 28°C. The temperature tolerant strain N40 produced maximum IAA and solubilized insoluble tri calcium phosphate compared to other strains and thus can be used microbial inoculant in biofertilizer technology.

**Keyword:** *Rhizobium leguminosarum* bv. *viciae*, indole acetic production, phosphate solubilization, temperature, pH tolerance

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## INTRODUCTION

Pea (*Pisum sativum* L.), an important leguminous crop belongs to family Fabaceae. China, India, Canada, Russia, France and the United States of America are the main pea countries (Food and Agriculture Organization, 2012). Plants are cultivated in temperate regions and during winter season in tropical countries. In India, the area under pea cultivation is about 275 thousand hectares which accounts 4.5 per cent global area under pea cultivation. Pea is an important nutritional pulse and the green peas contain a form of omega-3 fats alpha-linolenic acid (ALA). Rhizobia are the single largest contributor to the biological N<sub>2</sub> fixation (Jason et al., 2012). Legume-*Rhizobium* interaction is a key model for understanding the molecular basis of communication which represents most studied one but not fully understood symbiosis (Sachs et al., 2013). Plant growth promoting rhizobacteria (PGPR) as a microbial inoculant provide an alternative to chemical fertilizers and pesticides. In recent years, the use of PGPR has increased the productivity tremendously worldwide (Asghar et al., 2002). Combination of PGPR and efficient strains of *Rhizobium* in integrated biofertilizer technology has enhanced the growth of pea by inducing nitrogen fixation and efficient symbiosis (Kaur and Khan, 2013). Inoculation of *Rhizobium* strains with high temperature tolerance and effective nodulation to enhance plant growth would be required to improve crop productivity under climate change scenario (Singh et al., 2013, Gray and Smith, 2005).

High soil temperature poses a major problem for cultivation of legume crop. Temperature of the surface soil (5-10 cm) layer reaches up to 40-50°C in the arid zone of North West India in the summer months and the microbial inoculants are often exposed to high temperatures which adversely affect the survival of rhizobia strains in the soil and establishment rhizobia-legume symbiosis. The prolonged exposure to high temperature reduces the saprophytic survival of rhizobia in soil and effective nodulation (Kiran et al., 2007). Temperature affects *Rhizobium*-legume symbiosis in various ways such as poor infection of root hairs, differentiation of bacteroids in nodule and nitrogen fixation. As an important biotic factor, temperature influences the plant

growth, metabolic enzymes activity and uptake of nutrients and indirectly on the solute solubility, transport of ions, membranes permeability, density and many colloidal properties of matter in soil-water interface (Sardesai and Babu, 2001; Xie et al., 2013). *Rhizobium* is a mesophilic bacterium and the growth optimum temperature ranges from 25°C to 31°C (Somasegaran and Hoben, 1994) with the upper temperature limits ranges between 32 - 47°C (Munevar and Wollum, 1988). The climate of Varanasi is humid subtropical with high temperature ranges between 32 - 46°C in summer and 5 - 15°C during winter (Singh et al., 2012). In the present communication, isolation and screening of high temperature and pH tolerant strains of *Rhizobium* isolated from pea nodules have been described. Further, the growth promoting parameters such as IAA production and solubilization of phosphate were also investigated in selected strains.

## MATERIAL AND METHODS

### *Rhizobium* strains

Healthy nodules from Pea (*Pisum sativum* L.) roots were collected from the 20-25 days old plants growing at different location of Eastern Uttar Pradesh, India. Healthy determinate nodules were washed several times thoroughly with tap water and transferred to sterile water. Flask was shaken on a shaker thoroughly. Surface sterilized nodules with 95% ethanol for 1 min, followed by treatment with 0.1% HgCl<sub>2</sub> for 5 min and cleaned with sterile distilled water (Chaintreuil et al., 2000). A loop full crushed surface sterilized nodule was inoculated on yeast-extract mannitol (YEM) agar plates and incubated in a BOD incubator at 28°C for 2-3 days. The pure colony were selected and incubated on Bakelite screw cap tube slants in same medium at 4°C for maintenance.

The strains were identified by cross inoculation test and based on 16s rRNA gene sequence phylogeny following the standard protocol.

### Temperature tolerance

Temperature tolerance of the strains was investigated by growing the strains at 28°C and 45°C in 10 ml YEM broth with an inoculum of 10<sup>8</sup> CFU/ml. The growth was measured in terms of absorbance at 420 nm in a Spectrophotometer.

Pea plant seeds were inoculated with *R. leguminosarum* bv. *viciae* strains and nodulation was checked on slant culture at 45°C in a Growth Chamber. Also, the bacterized seed were grown in the sterilized soil in earthen pot to observed the nodule formation and growth of the host plant as compared to control.

### Effect of high pH

Isolated *Rhizobium* strains were grown at pH 9 and 11 on YEMA plates to observe their pH tolerance. The pH was maintained with sodium hydroxide (NaOH) and stabilized with Tris-HCl buffer (10mM). The plates were incubated in a BOD incubator at 28°C.

### Indole acetic acid (IAA) production

IAA produced by *Rhizobium* strains was determined in cultures supplemented with 100 µg/ml filter sterilized L-tryptophan at 28°C with continuous shaking for 48 hours. Cell biomass was centrifuged at 10,000 x g and the IAA produced (ml<sup>-1</sup> culture) was measured by mixing 2 ml culture supernatant with 4 ml of Salkowski reagent (1 ml 0.5 M FeCl<sub>3</sub> in 50 ml of 35% perchloric acid). The absorbance of color developed was measured at

530 nm after 30 min (Gordon and Weber, 1951). The amount of IAA produced was determined with the help of standard curve of IAA (10- 100 µg/ml) and calculated by equation,  $y = mx + c$  (in µg/ml) where:

$y$  = O.D. of *Rhizobium* culture.  $m$  = O.D. of blank solution

$x$  = amount of IAA produces by *Rhizobium* isolates  $c$  = Zero (constant)

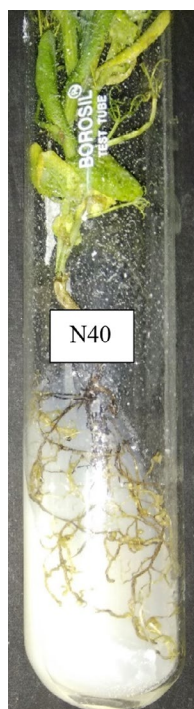
### Phosphate solubilization

Phosphate solubilization of *Rhizobium* strains was determined on Pikovskaya nutrient agar plates (Pikovskaya, 1948). The plates were

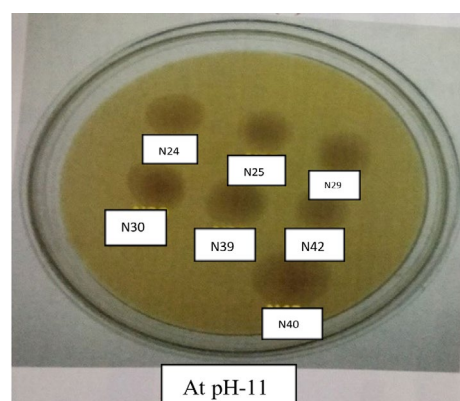
**Table 1.** Growth and survival of *Rhizobium leguminosarum* bv. *viciae* strains at 28°C and 45°C temperatures

Strain	Absorbance (420 nm) of cultures grown		Percent growth at 45°C as compared to 28°C
	at 28 °C 4 DAI	at 45 °C 4 DAI	
P09	2.664	1.987	74.6
N13	1.457	ND*	ND
N16	1.875	ND*	ND
N24	2.473	1.476	59.68
N25	2.785	2.465	88.5
N29	2.598	1.463	56.31
N39	2.645	1.534	60.54
N30	2.975	2.398	80.60
N40	2.987	2.875	96.25
N42	2.527	1.364	57.98

\*ND= Not detectable, DAI= Day after incubation



**Fig. 1.** Nodulation by *Rhizobium leguminosarum* bv. *viciae* strain N40 at 45°C on pea plant



**Fig. 2.** Growth of *Rhizobium* strains at pH-11

inoculated as spots with 10µl of exponentially grown *Rhizobium* strains (Yanni et al., 2001). The radius of the clear zone including colony was measured and phosphate solubilization activity was determined in mm per unit time. The phosphate solubilization index (SI) was determined using the following formula by measuring diameters of halo (clear zone) and colony.

$$SI = \frac{\text{Colony diameter} + \text{halo zone diameter}}{\text{Colony diameter}}$$

## RESULTS

The pea nodulating symbiotic bacterial strain was identified as *Rhizobium leguminosarum* bv *viciae* as revealed by 16s rDNA gene sequence phylogeny and the accession number of the strain is KT150239.1 which has been submitted in NCBI.

### Temperature tolerance of *Rhizobium leguminosarum* bv. *viciae* strains

Ten clones of *Rhizobium leguminosarum* bv. *viciae* were isolated from the pea nodules collected from North-Eastern districts of UP, India. Based on their growth in YEM broth and agar plates at elevated temperature and cross inoculation test these were numbered as N13, N16 (temperature sensitive), N24, N25, N29, N30, N39, N40 and N42 (temperature tolerant) selected for further studies. The strains were grown at two different temperatures viz 28°C and 45°C in YEM broth to observe their temperature tolerance presented in Table1.

The growth of all the strains retorted at 45°C as compared to at 28°C, however, some of them could survive and grow well at 45°C. The

growth recorded at 45°C was in decreasing order of N40>N25>N30>N39>N29>N42>N24 which indicated that strains N40 could be considered as the best temperature tolerant among the strains. The strains N13 and N16 failed to grow at 45°C (Table 1).

A nodulation test whether strain N40 is efficient for effective and competent symbiosis at 45°C was performed. The pea seed inoculated with strain N40 formed pink color nodules on slant culture in growth chamber at this temperature. Also, the bacterized pea seed formed effective nodules in sterilized soil in earthen pot and the growth of host plant was better as compared to control. A photograph showing nodulation on slant culture has been presented in Fig. 1

### pH tolerance of *Rhizobium leguminosarum* bv. *viciae* strains

All the *Rhizobium* strains (N13, N16, N24, N25, N29, N30, N39, N40, and N42) grew well at pH 9. The maximum growth of *Rhizobium* strains at pH 9.0 was in N40, followed by N42, N24, N29, N30, N25, N13 and N16. At pH 11, strain N40 showed maximum growth, followed by N30, N24, N42, N29, N39, N25, N13 and N16. Growth pattern of *Rhizobium* levels. The isolate N40 exhibited maximum phosphate solubilization activity (Figs. 4 and 5).

## DISCUSSION

*Rhizobium* is a mesophilic bacterium which forms determinate (pink) nodules at soil temperature between 15-30°C. Global warming and climate change have necessitated

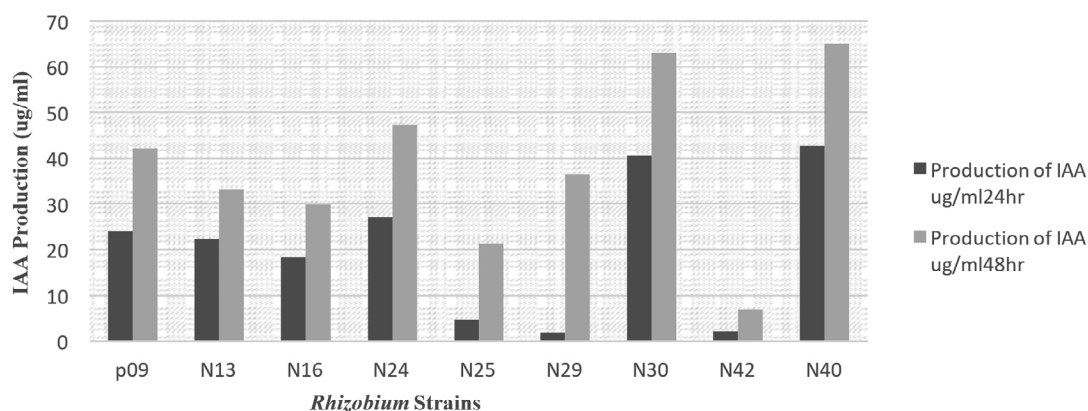


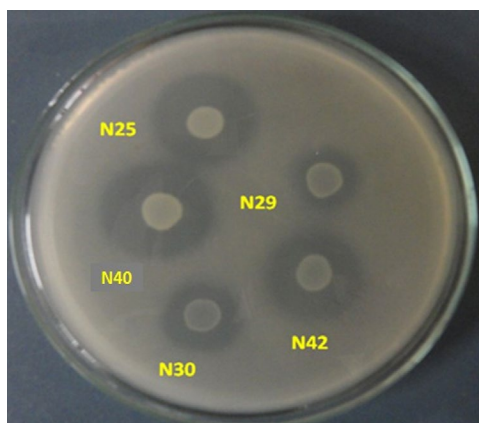
Fig. 3. IAA production by *Rhizobium* strains after 24 and 48 hours of incubation

microbiologist to isolate such strains of rhizobia which could tolerate high temperature with additional characteristics of growth promotion. Growth and survival of three *Rhizobium* strains N25, N30, and N40 at 45°C higher than the other strains and the performance of strain N40 was best. *Rhizobium* strains AER, ANR, ASR, ATR, ANR nodulating legume trees in South Riyad exhibited grow that elevated temperatures of 35°C and 45°C (Shetta et al.,2011). Fifteen *Pseudomonas fluorescense* and *Rhizobium* rhizospheric strains were isolated from Hyderabad soils with a view to screen temperature tolerant isolates(Manasa

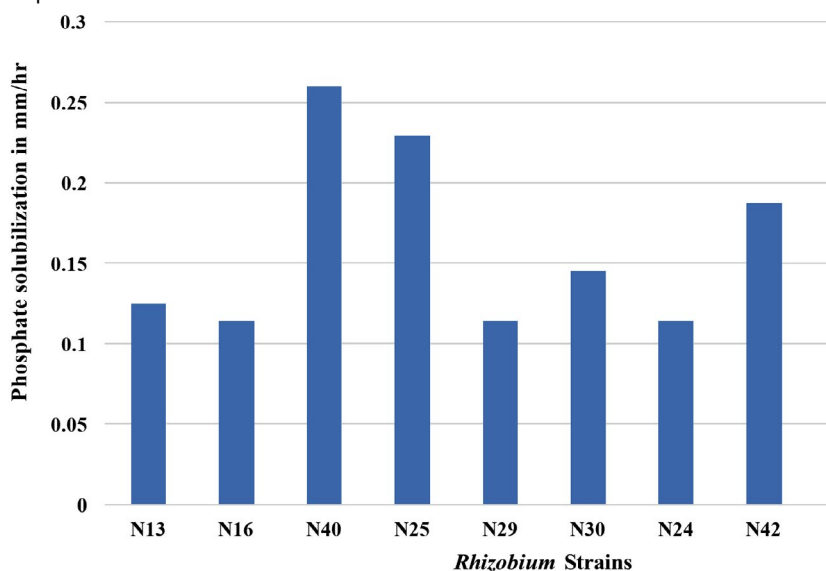
et al.,2017). Two *Rhizobium* isolates RR-1, GNR-1 and *Pseudomonas fluorescense* isolates GGP-1 were temperature tolerant with temperature optima of the growth for most isolates at 40°C. Fast growing *Rhizobium* isolates generally have less temperature tolerance as compared to slow growing (Singh et al., 2001). Isolate NE19 reduced the pH of the medium significantly and exhibited acid phosphatase activity. The strains exhibited different capacity to solubilize P. These strains require test to assess their field performance.

In this present study all the *Rhizobium* strains showed an appreciable amount of phosphate solubilization activity. The maximum phosphate solubilization activity was in N40 (0.260mm/hr). Actinobacteria solubilized insoluble tricalcium phosphate or aluminium phosphate as sole sources of phosphorus. The rhizospheric bacterial strains T1C, T1H, T3A, T3C, P3E, F1A, F2A and V2B expressed phosphate solubilization in the quantitative assay which could be used for further studies to evaluate plant growth promotion activities (Salcedo et al.,2014). Although P-solubilization was not evaluated in N40 strain at 45°C but on the basis of its growth at 45°C it can be speculated that this strain may be efficient P-solubilizer at this temperature.

All the *Rhizobium* strains present study produced IAA using tryptophan as substrate which was maximum (65.13µg/ml) by strain N40. The



**Fig. 4.** Phosphate solubilization by some *R. leguminosarum* bv. *viciae* strains indicated by clear zone formation on nutrient agar plate containing tricalcium phosphate



**Fig. 5.** Phosphate solubilization of *Rhizobium leguminosarum* bv. *viciae* strains

rhizospheric PGPR from Tunjung isolates, UPMB19, endophytic PGPR from Besut, Terengganu UPMB20, rhizobia from soybean UPMR 30 and from Mimosa UPMR31 were screened for plant inoculation trials along with a oil palm PGPR isolate UPMB 10 as the reference strain (Tan et al., 2014). Strain UPMB19 significantly stimulated the seedling (7 days after transplanting) growth which could be attributed to the higher  $N_2$ -fixation and IAA production rates. Karnwal (2012) isolated *Rhizobium* strains from maize and wheat plant which were able to produce different amount of IAA in a range of 0.6 to 2.7  $\mu\text{g/ml}$ . *Rhizobium leguminosarum* bv. *viciae* N40 formed effective pink color nodulation on pea plant and in slant the pod cultures. The growth and nodule formation were superior under both the conditions.

Global warming and frequent climate change are posing serious threat to the farmers world-wide. The farmers need such varieties which could tolerate high temperature and some time temperature and drought both. Thus, the microbial inoculant *Rhizobium* should also possess the temperature tolerance capacity. Our temperature tolerant *R. leguminosarum* bv. *viciae* strain N40 could tolerate 45°C temperature with a slight decrease in growth compared to mesophilic growth condition. Also, this strain has additional PGPR traits activity such as phosphate solubilization and IAA production. However, it is necessary to test the performance of strain N40 for PGPR activity under field condition before it is recommended for use as microbial inoculant. Among the *R. leguminosarum* strains, N40 strain could tolerate the performance to pH tolerance was high. The results suggested that the strain tolerant to one adverse condition may be also tolerant to other stresses. Under the climate change condition  $\text{CO}_2$  fertilization is expected to enhance the tremendous growth of the crop plants while the elevated temperature level may have adverse effect. The temperature tolerance strain under these conditions will be most favoured choice for the farmers.

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#### CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

#### AUTHORS' CONTRIBUTION

Every author has significantly contributed in experimentation and preparation of manuscript.

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#### ETHICS STATEMENT

Not applicable.

#### DATA AVAILABILITY

The datasets generated and/or analysed during the current study are available from the corresponding author on reasonable request.

#### REFERENCES

1. Asghar HN, Zahir ZA, Arshad M, Khaliq A. Relationship between in vitro production of auxins by rhizobacteria and their growth promoting activities in *Brassica juncea* L. *Biol Fert Soils*. 2002;(35):231-237. doi: 10.1007/s00374-002-0462-8
2. Chaintreuil C, Giraud E, Prin Y, et al. Photosynthetic bradyrhizobia are natural endophytes of the African wild rice *Oryza breviligulata*. *Appl Environ Microbiol*. 2000;66:5437-5447. doi: 10.1128/AEM.66.12.5437-5447.2000
3. Edi-Premono M, Moawad AM, Vlek PLG. Effect of phosphate-solubilization *Pseudomonas putida* on the growth of maize and its survival in the rhizosphere. *Indones. J Crop Sci*. 1996;11:13-23.
4. Gordon SA, Weber RP. Colorimetric estimation of indoleacetic acid. *Plant physiology*. 1951;26(1):192. doi: 10.1104/pp.26.1.192
5. Gray EJ, Smith DL. Intracellular and extracellular PGPR commonalities and distinctions in the plant-bacterium signalling processes. *Soil Biology and Biochemistry*. 2005;37:395-412. doi: 10.1016/j.soilbio.2004.08.030
6. Jason CH, Jeffrey HD, Daniel JF, McMahan LG, Genggeng Q, Christopher WJ. Designing Adsorbents for  $\text{CO}_2$  Capture from Flue Gas- Hyperbranched Aminosilicas Capable of Capturing  $\text{CO}_2$  Reversibly. *J Am Chem Soc*. 2012;10(130):2902-2903.
7. Karnwal A. Screening of plant growth promoting rhizobacteria from maize (*Zea mays* L.) and wheat (*Triticum aestivum* L.). *African J food Agric Nutrition Dev*. 2012;12:6170-6185.

8. Kaur S, Khanna V. Effect of temperature-tolerant rhizobial isolates as PGPR on nodulation, growth and yield of Pigeon pea [ *Cajanus cajan* (L) Milsp.]. *Journal of Food Legumes*. 2013;26(3 & 4):80-83.
9. Kiran D, Krishnamoorthy U. Rumen fermentation kinetics and nitrogen degradability of commonly used ruminant feeds tuffs *in vitro*. *Anim Nutr Feed Technol*. 2007;7(1):63-71.
10. Manasa K, Subhash RR, Trivenu S. *In vitro* screening of temperature stress tolerance of Rhizobial and *Pseudomonas fluorescence* isolates. *J Pharmaco Phytochem*. 2017;6(5):764-67.
11. Manivannan M, Ganesh P, Kumar SR, Tharmaraj K, Ramya SB. Isolation, Screening, Characterization and Antagonism Assay of PGPR Isolates from Rhizosphere of Rice Plants in Cuddalore District. *Int J Pharm Biol Arch*. 2012;3:179-185.
12. Munevar FAG, Wollum II. Response of soybean plants to high temperature as affected by plant cultivar and *Rhizobium* strain. *Agronomy Journal*. 1982;74:138- 142. doi: 10.2134/agronj1982.00021962007400010036x
13. Rai R, Dash PK, Mohapatra T, Singh A. Phenotypic and molecular characterization of indigenous rhizobia nodulating chickpea in India. *Indian J Exp Biol*. 2012;50:340-350.
14. Sachs JL, Gano A, Hollowell AC, Regus JU. The Legume-*Rhizobium* Symbiosis. *Ecology*. 2013;54:435-446. doi: 10.1093/obo/9780199830060-0095
15. Salcedo LDP, Prieto C, Correa FM. Screening phosphate solubilizing actinobacteria isolated from the rhizosphere of wild plants from the Eastern Cordillera of the Colombian Andes. *Biol Fertil Soils*. 2014;56:765-773.
16. Sardesai N, Babu CR. Poly- $\beta$ -hydroxybutyrate metabolism is affected by changes in respiratory enzymatic activities due to cold stress in two psychrotrophic strains of *Rhizobium*. *Current Microbiology*. 2001;42(1):53-58. doi: 10.1007/s002840010178
17. ShettaND, Al-Shaharani TS, Abdel-Aal M. Identification and Characterization of *Rhizobium* Associated with Woody Legume Trees Grown under Saudi Arabia Condition. *Am Eurasian J Agric Environ Sci*. 2011;10:410-418.
18. Singh RK. Genetics of thermotolerance in Ciprofloxacin resistance mutant of *Rhizobium leguminosarum* bv. *Phasesl*. *Ind J Expl Boil*. 2001;39:818-820.
19. Singh RK, Malik N, Singh S. Impact of rhizobial inoculation and nitrogen utilization in plant growth promotion of maize(*Zea mays* L.). *Bioscience*. 2013;5:8- 14. doi: 10.13057/nusbiosci/n050102
20. Somasegaran P, Hoben HJ. Quantifying the growth of rhizobia. In Handbook for Rhizobia (pp. 47-57). *Springer*, New York, 1994. doi: 10.1007/978-1-4613-8375-8\_5
21. Tan KZ, Radziah halimi MS, Khairuddin AR, Cabib SH, Khamsuddin ZH. Isolation and characterization of rhizobia and plant growth-promoting rhizobacteria and the effects on growth of rice seedlings. *Am J Agric Biol Sci*. 2014;9(3):342-360. doi: 10.3844/ajabssp.2014.342.360
22. Xie B, Bishop S, Stessman D, Wright D, Spalding MD, Halverson LJ. *Chlamydomonas reinhardtii* thermal tolerance enhancement mediated by a mutualistic interaction with vitamin B12-producing bacteria. *ISME Journal* (online). 2013;7:1544-1555. doi: 10.1038/ismej.2013.43
23. Yanni YG, Rizk RY, El-Fattah FKA, Squartini A. The beneficial plant growth-promoting association of *Rhizobium leguminosarum* bv *trifolii* with rice roots. *Aust J Plant Physiol*. 2001;28:845-870. doi: 10.1071/PP01069