



Green Synthesis and Antimicrobial Potential of Silver Nanoparticles with *Boerhavia procumbens* Extract

Muhammad Rizwan¹ , Said Amin², Bates Kudaibergenova Malikovna³ , Abdur Rauf⁴ , Muhammad Siddique⁵ , Kamran Ullah⁶ , Saud Bawazeer⁷ , Umar Farooq⁸ , Yahia Naseer Mabkhot^{9,10}  and Mohamed Fawzy Ramadan^{11,12*} 

¹Centre for Biotechnology and Microbiology, University of Swat, KPK, Pakistan. ²Department of Microbiology and Biotechnology, Abasyn University Peshawar, Pakistan. ³Department of Chemistry and Chemical Technology, Al-Farabi Kazakh National University, Almaty, Kazakhstan. ⁴Department of Chemistry, University of Swabi, KPK, Pakistan. ⁵PCSIR Complex Peshawar, KPK, Pakistan. ⁶Department of Zoology, University of Swabi, KPK, Pakistan. ⁷Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Umm Al-Qura University, Makkah, P.O. Box 42, Saudi Arabia. ⁸Department of Chemistry, University of COMSATS, Abbottabad, Pakistan. ⁹Department of Pharmaceutical Chemistry, College of Pharmacy, King Khalid University, Abha, Saudi Arabia. ¹⁰Research Center for Advanced Materials Sciences (RCAMS), King Khalid University, Abha-6113, Saudi Arabia. ¹¹Deanship of Scientific Research, Umm Al-Qura University, Makkah, P.O. Box 715, Saudi Arabia. ¹²Department of Biochemistry, Faculty of Agriculture, Zagazig University, Zagazig 44519, Egypt.

*Correspondence: mframadan@zu.edu.eg

(Received: April 02, 2020; accepted: June 05, 2020)

Citation: Rizwan M, Amin S, Malikovna BK, et al. Green Synthesis and Antimicrobial Potential of Silver Nanoparticles with *Boerhavia procumbens* Extract. *J Pure Appl Microbiol.* 2020;14(2):1437-1451. doi: 10.22207/JPAM.14.2.42

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Abstract

The goals of the present study were to use silver nitrate (AgNO₃) solution to synthesize plant-mediated silver nanoparticles (AgNPs) using *Boerhavia procumbens* extract, to evaluate the antimicrobial potential of crude *B. procumbens* extracts as well as the antimicrobial potential of synthesized AgNPs. The antimicrobial activity was tested against ten pathogenic bacterial strains including *Klebsiella pneumonia*, *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Citrobacter braakii*, *Providentia spp.*, *Salmonella typhi*, *Salmonella para typhi*, *Vibrio cholera*, and *Proteus vulgaris* and seven fungal species; *Rhizopus stolonifer*, *Candida albican*, *Alternaria alternata*, *Aspergillus flavus*, *Verticillium chlamydosporium*, *Penicillium chrysogenum*, and *Aspergillus oryzae*. The methanol extract was fractionated using several solvents and subjected to phytochemical analysis along with FTIR. Phytochemical analyses revealed flavonoids, tannins, saponins, steroids, quinones, and phenols in the crude plant extract. AgNPs were synthesized using *B. procumbens* extract and characterized by UV-Vis, Fourier-transform infrared spectroscopy (FTIR), and Scanning Electron Microscopy (SEM). Synthesized AgNPs were spherical, with 20-80 nm diameter. The absorption peak of synthesized AgNPs was observed at 392 nm. AgNPs have significant antimicrobial potential against selected pathogenic bacterial and fungal species as compared to different fractions of crude *B. procumbens* extract. The current study suggests that green synthesis is a useful technique and can be used as an alternative to antimicrobial agents against pathogenic organisms.

Keywords: *Boerhavia procumbens*, Nyctaginaceae, scanning electron microscopy, FTIR, silver nanoparticles, Quinones

INTRODUCTION

Medicinal plants have been widely used and considered the most common sources of medicine in the pharmaceutical industry (Ramadan 2008; Ramadan et al. 2008; Mohdaly et al., 2015; Abo El-Maati et al. 2016; Shakya, 2016; Rauf et al., 2018). These plants produced a wide range of bioactive compounds (Bajguz, 2007; Cushnie et al., 2014). The composition of plant secondary metabolites depends on the species, association with microbes, and soil type (Zhao et al., 2011; Morsy, 2014). These bioactive compounds have potential against a wide range of bacterial infections and the best source for the formulation of drugs, declared by the World Health Organization (Iwu et al., 1999; WHO Traditional Medicine Strategy, 2002).

Nanotechnology or nanoscale technology is a strong field of novel research dealing with design, manipulation, and synthesis of particles with 1-100 nm (Sultana et al., 2019). Nanoparticles (NPs) have several uses in cosmetics, health care, food and feed, mechanics, optics, environmental health, drug-gene delivery, biomedical sciences, electronics, space industries, chemical industries, optoelectronics, energy science, catalysis, single-electron transistors, nonlinear optical devices and photo-electrochemical applications (Iravani et al., 2014). Nanotechnology finds its importance by

generating nano-products and NPs with novel and size-related physicochemical traits in contrast to bigger molecules (Chouhan, 2018).

The most important branch of nanotechnology is plant-mediated green synthesis. Due to the environmentally friendly and practical cost, it has gained more importance (Chanel et al., 2017). Green synthesis has many advantages, i.e., less toxicity, cost-effective and straightforward, less energy consumption, perform activities under moderate conditions, and combined the effect of nanoparticles and plant bioactive constituents. Furthermore, green synthesized silver nanoparticles possess more biological potential than nanoparticles synthesized by chemical methods (Choudhury et al., 2016).

Silver NPs (AgNPs) are of interest due to their novel chemical, physical, and biological traits compared to their macro-counterparts (Sultana et al., 2019). AgNPs possess distinct physicochemical properties, comprising a high thermal and electrical conductivity, surface- chemical stability, enhanced Raman scattering, catalytic activity, and non-linear optical properties (Khattak et al., 2019). Those characteristics are responsible for their role in the formation of inks, in medical imaging and microelectronics. Besides, AgNPs reveal wide-spectrum fungicidal and bactericidal potential, making them highly accessible in functional

products (Tran and Le, 2013).

Boerhavia procumbens belong to the family Nyctaginaceae, nearby termed *Biskhpara/Jangli*, and *spreading hogweed*. Leaves are large, broad, and rounded beneath with a vigorous shape. Its branches (about 3 ft.) are glabrous and slender. The branches are long, glabrous, pubescent, stout, or viscous, with 1-2 inches leaves. They are oval, dull-witted, acute, and ordinarily cordate green beneath with panicle inflorescence (Fig. 1). Its habitat Africa, Pakistan, the USA, and India. In Pakistan, it could be found in Peshawar, Attock, Hazara, Sind, Baluchistan, Multan, Thal to Kurram, and Rawalpindi. Its color is purplish-red, and the blooming period is January to August (Hussain et al., 2012). *Boerhavia procumbens* has a lot of pharmaceutical and health-related benefits. Medicinal applications of *Boerhavia procumbens* include diaphoretic, diuretic, expectorant, emetic, laxative, stomachic, rejuvenates and purgative (Khalil et al., 2017; Wajid et al., 2017; Khan et al., 2020). It is mentioned to be effective against cardiac disorders, asthma, hemorrhoids, edema, eye diseases, insomnia, jaundice, kidney stone, nervous system related disorders, cough, against snake venom, urethritis, skin related diseases, intestinal, anemia, and colic kidney disorders (Hussain et al., 2012; Ali et al., 2019).

The purpose of the current investigation was to carry out green synthesise of AgNPs using aqueous extract of *Boerhavia procumbens* and to study the antimicrobial potential of different fractions and synthesized AgNPs against pathogenic bacteria and fungal species.

MATERIAL AND METHODS

Plant collection

B. procumbence was collected from the botanical garden of the Medicinal Botanic Centre of Pakistan Council for Scientific and Industrial Research (PCSIR), Peshawar, Pakistan. The plant was identified in the Department of Botany, University of Peshawar. The leaves having a voucher name BP554 was submitted for future correspondence. The freshly collected plant was washed carefully, then air-dried for 15 days under the shed (Pirtarighat et al., 2019). The dried plant was grounded to make a coarse powder using an electric grinding machine.

Preparation of plant extract

The crude extracts were prepared with different solvents, including methanol, water, ethanol, ethyl acetate, chloroform, and *n*-hexane. All solvents were taken into flasks, and plant material was soaked in flasks for three days. The solvents were decanted, and the procedure was repeated four times for each solvent. All extracts were concentrated using a rotary evaporator at 45°C under vacuum. The extracts were kept in the refrigerator for further analysis (Pirtarighat et al., 2019).

Moisture determination

Dried powdered *B. procumbens* (5 g) was pre-weighed in the Petri dish and kept at 105-110°C for three h. Moisture (%) was calculated regarding the air-dried sample (Pirtarighat et al., 2019).

$$\% \text{ Moisture} = \frac{\text{Wight of moisture in g}}{\text{Wight of plant material (g) before heating}} \times 100$$

Determination of extractive value

The total weight of 5 g of each dried powder of plant material was extracted for about 24 h with 100 mL of respective solvent (methanol, ethanol, ethyl acetate, chloroform, and *n*-hexane). The flasks were shaken, and the mixture allowed to stand overnight, followed by recovery of solvents by decantation and filtration. The filtrates were dried, and the extract was weighted in percentage (Singh et al., 2013).

$$\% \text{ Extract} = \frac{\text{Wight of the extract (g)}}{\text{Wight of plant material (g)}} \times 100$$

Determination of ash

The total weight of 5 g of dried powder of *B. procumbens* was burnt at 600°C in a furnace for three h followed by cooling. The ash was weighed, and the percentage calculated regarding the air-dried *B. procumbens* sample (Premnath et al., 2012).

$$\% \text{ Ash} = \frac{\text{Wight of plant material after burning}}{\text{Wight of plant material before burning}} \times 100$$

Phytochemical analyses

B. Procumbens fractions were analyzed for the presence of phytochemical constituents, according to Singh et al. (2013).

Preparation of silver nanoparticles (AgNO₃)

Ten g of the plant powder was boiled with 100 mL deionized distilled water. The resultant aqueous solution was filtered using Whatman filter paper three times to remove all the insoluble material. This extract was added to a 10⁻³ M silver nitrate (AgNO₃) solution. Analytical grade AgNO₃ (Molecular weight 169.87 g/mol) was purchased from Merck (Darmstadt, Germany). The solution was heated for three h in a water bath at 60°C. The solution color changed from light yellow to brown-reddish. It showed the reduction of pure Ag (I) ions. The reduced ions were monitored and measured using UV-visible spectra of the mixture of plant extract and AgNO₃ solution at regular intervals. Peaks were measured on UV-Vis spectra from 200 nm and 700 nm (Shakeel et al., 2015).

Evaluation of antimicrobial activity

Antimicrobial activities were performed using well diffusion method for both *B. procumbens* crude extracts and AgNPs. The ethyl acetate, methanol, ethanol, water, chloroform, and *n*-hexane extracts of the plant and AgNPs solutions of *B. procumbens* were applied against ten pathogenic bacteria (*Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Escherichia coli*, *Klebsiella pneumoniae*, *Citrobacter braakii*, *Proteus vulgaris*, *Salmonella Typhi*, *Salmonella Para typhi*, *Vibrio cholerae*, and *Providencia stuartii*) and five fungal strains (*Candida albicans*,

Alternaria alternata, *Aspergillus flavus*, *Aspergillus niger*, and *Aspergillus oryzae*). The agar media was prepared then autoclaved and dispensed into sterilized plates in Laminar flow hood (LFH) and left for the solidification. The test microorganism uniform lawn was arranged on the surface of nutrient agar media in *Petri* plates with the help of a sterile swab. Uniform wells of 6 mm were bored in each plate, with sterile metallic borer. Ten mg of test samples (fraction and AgNPs) were prepared in 1.0 mL of DMSO (10%) and added into particular wells. *Petri* dishes were incubated for 16 h at 37°C. The zones were determined after the incubation period from the width of the wells. The formula documented the percent inhibition:

$$\text{Percent inhibition} = \frac{\text{Zone of inhibition of test sample}}{\text{Zone of inhibition of standard}} \times 100$$

RESULTS

The current study was performed to evaluate the antimicrobial potential of *B. procumbens* crude extract, to synthesize AgNPs from *B. procumbens* extract and to determine their biological effects against pathogenic bacteria and fungi. Leave extracts were prepared in methanol, ethyl acetate, ethanol, chloroform, *n*-hexane, and water. Ash (19.38%), moisture (8.2%), and extractive values were also determined (Table 1). The FTIR of all extracts was performed

Table 1. Ash, moisture and extractive value (%) of *B. procumbens*

Ash (%)	Moisture (%)	Extractive value (%)					
		MeOH	EtOH	EtOAc	CHCl ₃	n-hexane	Aqueous
19.38	8.206	13	12	8	7	5	18

Table 2. Phytochemical analysis of *B. procumbens*

No.	Test name	MeOH	EtOH	CHCl ₃	n-hexane	EtOAc	H ₂ O
1	Tannins	++	+++	++	+	++	++
2	Flavonoids	++	++	++	-	++	+
3	Alkaloids	-	-	-	-	-	-
4	Terpenoids	-	-	-	-	-	-
5	Saponins	++	++	+	-	++	+++
6	Steroids	+++	+++	+++	++	++	-
7	Quinones	++	++	+	-	+	+
8	Phenols	+++	+++	+	++	++	+

+++ : strongly, ++moderately, + weak

Table 3. Zone of inhibition by *B. procumbens* crude extract

Bacterial Species	Zone of inhibition (mm)							AgNPs	Positive Control	Negative Control
	H ₂ O	MeOH	EtOH	EtOAc	CHCl ₃	n-hexane				
<i>E. coli</i>	09±0.38	0±0	13±0.73	12±0.44	0±0	0±0	23±0.46	16±0.94	-	
<i>K. pneumoniae</i>	17±0.85	11±0.75	14±0.19	13±0.84	10±0.66	0±0	22±0.28	11±0.26	-	
<i>S. aureus</i>	11±0.02	0	14±0.33	14±0.95	0±0	0±0	22±0.18	15±1.03	-	
<i>P.aeruginosa</i>	0±0	12±0.66	15±0.85	14±0.28	12±0.92	0±0	19±1.03	18±0.37	-	
<i>C. braakii</i> ,	13±0.38	12±1.03	16±0.86	14±1.06	11±0.02	09±0.55	20±0.44	13±0.68	-	
<i>Providentia Spp.</i>	10±0.28	12±0.23	15±0.90	13±0.46	09±0.28	0±0	17±0.90	18±0.39	-	
<i>S. typhi</i>	15±0.55	09±0.35	12±1.00	14±0.94	09±1.04	09±0.99	20±0.23	15±1.02	-	
<i>S. para typhi</i>	0±0	10±0.69	15±0	13±0.28	0±0	0±0	23±0.84	14±0.33	-	
<i>V. cholerae</i>	12±0.53	0±0	13±1.02	11±0.49	0±0	0±0	15±0.92	19±0.91	-	
<i>P. vulgaris</i>	09±0.18	08±0.39	14±0.29	12±0.84	0±0	07±0.18	15±0.49	13±0.25	-	

DMSO was used as -ve while gentamicin was used as +ve control

using FTIR Model IR Pretige-21 (Shimadzu, Japan). Different peaks of functional groups were detected in each fraction (Fig. 2-6).

Phytochemical analysis of *B. purocumbens* extract

The results of the phytochemical analyses, shown in Table 2, reveal that tannins were present in methanol, ethanol, ethyl acetate, chloroform extracts. Flavonoids were absent in the *n*-hexane extract, while present in all other extracts. Alkaloids and terpenoids were absent in all extracts. Saponins were absent in the *n*-hexane extract, while present in methanol, ethanol, ethyl acetate, chloroform, and water extracts. Steroids were present is less quantity in *n*-hexane extract, while strongly found in all the remaining extracts. Quinones was absent in *n*-hexane extract, while

detected in chloroform, ethyl acetate, ethanol, methanol, and aqueous extracts. Phenols were present in much higher concentration in methanol and ethanol extracts, while moderately present in *n*-hexane and EtOAc extracts, and least detected in chloroform and water extracts.

FTIR analysis of *B. purocumbens* extract

All fraction of crude extract of *B. purocumbens* were evaluated for the presence of functional groups using FTIR (Fig 2-7). The Fig. of each fraction was analyzed from its IR values and presented in a tabulated form (Table 5-10).

Characterization of AgNPs

FTIR of AgNPs

The FTIR analysis was carried out to identify AgNPs associated biomolecules. The



Fig. 1. Representative picture of *B. procumbens*

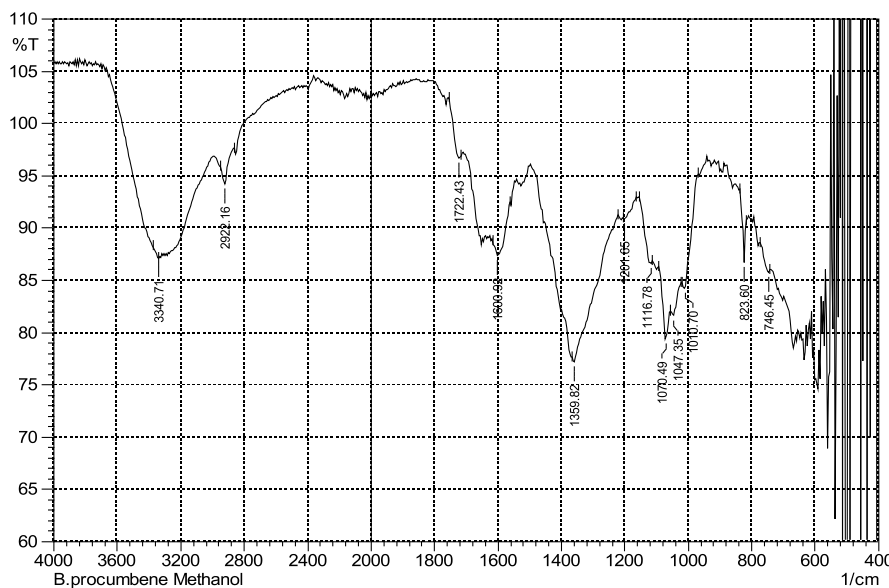


Fig. 2. FTIR analysis of MeOH fraction of *B. procumbens*

results showed that different compounds were detected in the AgNPs sample, i.e., phenols, alcohols, carboxylic acid, ketones, alkenes, nitro group, and ethers. After comparing the spectra of AgNPs and aqueous extract, it has been confirmed that AgNPs are synthesized. The differences of IR spectra of AgNPs and aqueous extract were due to the compounds of aqueous extracts that have been oxidized by silver ions. Some of the peaks present in aqueous IR spectra were absent

in AgNPs extracts. This means that AgNPs have reacted with these groups (Fig. 7-8). The FTIR analysis of AgNPS was interpreted using IR values and presented in Table 11.

pH and color

Synthesized AgNPs can also be confirmed physically by pH and color changes. AgNPs change their color from yellowish to dark brown, and pH changes from 6 to 5.5. The change in pH and color confirms the presence of AgNPs.

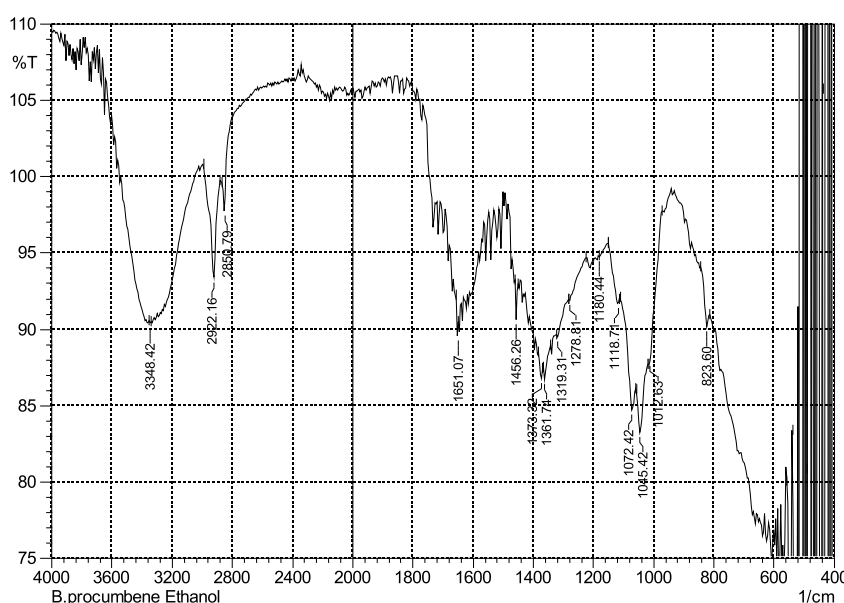


Fig. 3. FTIR analysis of EtOH fraction of *B. percombens*

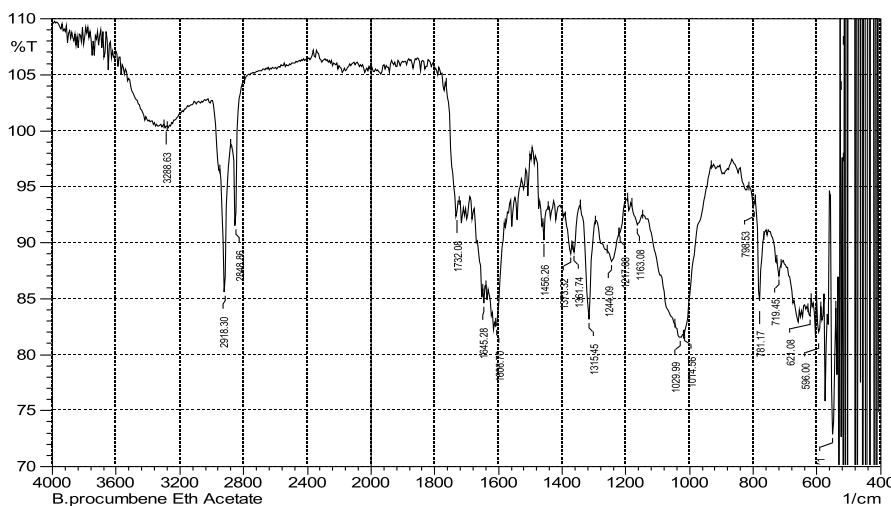


Fig. 4. FTIR analysis of EtOAc fraction of *B. percombens*

Table 4. Zones of inhibition of *B. percumbence* extracts

Fungal Species	H ₂ O	MeOH	EtOH	EtOAc	CHCl ₃	n-hexane	AgNPs	Positive Control	Negative Control
<i>A. oryzae</i>	13±0.23	21±0.65	17±0.24	15±0.84	13±0.84	00±00	22±0.38	14±0.48	-
<i>C. albicans</i>	10±0.39	17±0.22	20±0.84	18±0.37	16±0.58	16±0.65	20±0.74	17±0.65	-
<i>A. niger</i>	15±0.18	19±0.26	22±0.26	18±0.74	18±0.49	19±0.77	24±0.97	14±0.25	-
<i>A. alternata</i>	9±0.73	20±0.83	21±1.33	20±0.64	19±0.65	19±0.84	23±0.88	13±0.79	-
<i>A. flavus</i>	14±0.02	17±0.99	18±0.83	14±0.29	14±0.26	14±0.26	20±0.38	17±0.84	-

DMSO was used as -ve while fluconazole was used as +ve control

UV-Vis Spectroscopy of AgNPs

The UV-Vis spectroscopy is performed to characterize the AgNPs by the surface plasmon resonance (SPR) with a spectrum of 200 to 700 nm. The maximum peak absorbance with the high band was noted at 395 nm (Fig. 9).

Scanning Electron Microscopy (SEM) of AgNPs

The SEM picture shows the shape and size of the AgNPs. The SEM images showed spherical shape AgNPs in scattered form focused on 0.5 µm of area, with diameter ranges from 20-80 nm (Fig. 10).

Antimicrobial activity

For antimicrobial activities, *B. procumbens* extracts obtained from different solvents, as well as synthesized AgNPs were screened against bacterial and fungal pathogens (Table 3 and 4). *B. Percumbens* plant extracts prepared using different solvents, including water, MeOH, CHCl₃, EtOH, EtOAc, and *n*-hexane. MeOH and EtOH fractions resulted in significant activity against pathogenic bacteria and fungi. The extracts of the remaining fraction have no or very few activities. Furthermore, it was also clear that *B. Percumbens* synthesized AgNPs were more active than crude extracts (Tables 3 and 4). *B. Percumbens* synthesized AgNPs can be used to enhance the antimicrobial potential of plant constituents against a wide range of infections.

DISCUSSION

Medicinal plants have attracted significant attention to science communities due to the promising potential in life science (Antony and Sing., 2011). Pakistan is very rich in natural wealth because of it is excellent geographical condition, multiple ecological niches, and soil conditions. It

Table 5. FTIR analysis of MeOH fraction of *B. percumbens*

IR values	Bond	Functional group
3340	O-H Stretch	Phenols & Alcohols
2922	O-H Stretch	Carboxylic acid
1722	C=O Stretch	Ketones
1600	C=C Symmetric	Alkenes
1359	N=O Bend	Nitro group
1201	C-O Stretch	Ethers
1116	C-O Stretch	Ethers
1070	C-O Stretch	Ethers
1047	C-O Stretch	Ethers
1010	C-O Stretch	Ethers

is estimated that 400-600 species out of 5700 are present in Pakistan (Ahmad et al., 2007).

Nanotechnology continues to allow researchers and technologists to find new materials and essential benefits to people. However, it is also an essential part to consider the potential harmful impacts of nanoscale generated products on health as well as our environment (Hull and Bowman, 2018). Due to these facts, it is essential to determine how to decrease

dangerous impacts interred by new materials. In the present study, green synthesis of AgNPs was adopted to reduce the negative impact on health and environment because this is the safest method of preparation of AgNPs. The procedure used by Ahmed et al. (2016) for the synthesis of AgNPs by the chemical method was followed. The methodology for the synthesis of AgNPs was the same, but instead of chemical synthesis, the green synthesis method was used in the present study.

Table 6. FTIR analysis of EtOH fraction of *B. percumbers*

Values	Bond	Functional group	values	Bond	Functional
3348	C-C=C	Amines secondary	1278	C-O Stretch	Ethers
2922	H-C-H	Alkanes	1180	C-O Stretch	Esters
2850	C-H Stretch	Aldehydes	1118	C-O Stretch	Ethers
1651	C=O Stretch	Ketones	1072	C-O Stretch	Ethers
1485	H-C-H	Alkanes	1045	C-O Stretch	Esters
1375	N=O Bend	Nitro group	1012	C-O Stretch	Ethers
1361	N=O Bend	Nitro group			
1319	N=O Bend	Nitro group			

Table 7. FTIR analysis of EtOAc fraction of *B. percumbers*

Values	Bond	Functional group	values	Bond	Functional
3288	C-C=C	Amines primary	1373	N=O Bend	Nitro group
2918	H-C-H	Alkanes	1361	N=O Bend	Nitro group
2848	C-H Stretch	Aldehydes	1315	N=O Bend	Nitro group
1732	C=O Stretch	Esters	1244	C-O Stretch	Ethers
1645	C=O Stretch	Ketones	1217	C-O Stretch	Esters
1606	C=O Stretch	Ketones	1263	C-O Stretch	Ethers
1456	H-C-H	Alkanes	1029	C-O Stretch	Esters
			1014	C-O Stretch	Ethers

Table 8. FTIR analysis of EtOAc fraction of *B. percumbers*

Values	Bond	Functional group	values	Bond	Functional
3346	C-C=C	Amines secondary	1242	C-O Stretch	Esters
2918	H-C-H	Alkanes	1161	C-O Stretch	Ethers
2848	C-H Stretch	Aldehydes	1033	C-O Stretch	Esters
1738	C=O Stretch	Esters			
1732	C=O Stretch	Esters			
1602	C=O Stretch	Ketones			
1558	N=O Stretch	Nitro group			
1539	N=O Stretch	Nitro group			
1456	H-C-H	Alkanes			
1373	N=O Bend	Nitro group			
1315	N=O Bend	Nitro group			
1269	C-O Stretch	Ethers			

The most valuable NPs for medical uses are AgNPs, which are well known for their high antimicrobial activity. AgNPs has a remarkable option for the development of bioactive and novel drugs due to their broad-spectrum activities. Silver ion is famous as a metal ion that shows antimicrobial, antifungal, and anti-algal properties. Mainly, it has been widely used as silver nitrate solution in water, which has sterilizing and also disinfecting activities (Kheybari, 2010). Hussain et al. (2012) revealed that roots containing oil of *B. procumbens* are composed of methyl ester

derivatives of fatty acids. Eighteen types of different components were found and quantified as well. The most important and abundant was the methyl ester of linoleic acid (9%). In the present study, the phytochemical analysis revealed different types of phytochemicals, in which steroids are of great interest nowadays. *B. procumbens* was used as an entire plant for the process of phytochemical analysis, the plant contains eight important phytochemicals and having good antimicrobial activity. Furthermore, this ability was enhanced by synthesizing the

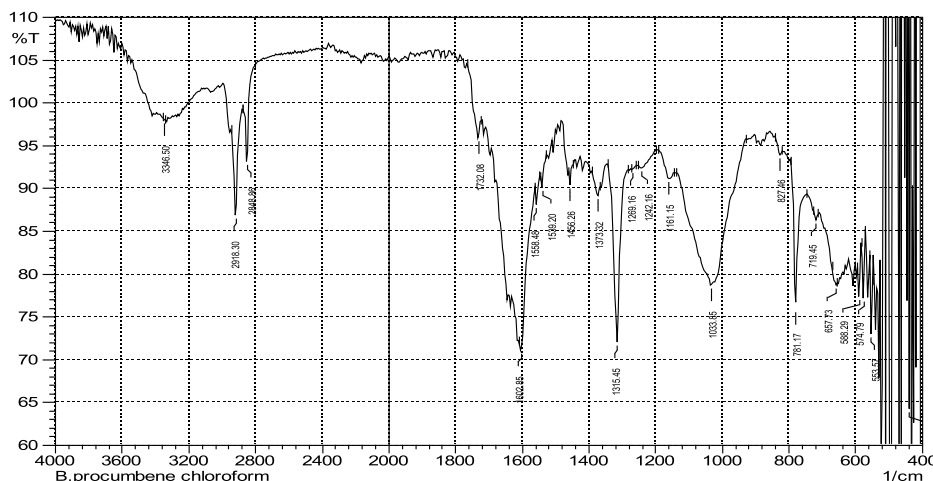


Fig. 5. FTIR analysis of CHCl₃ fraction of *B. procumbens*

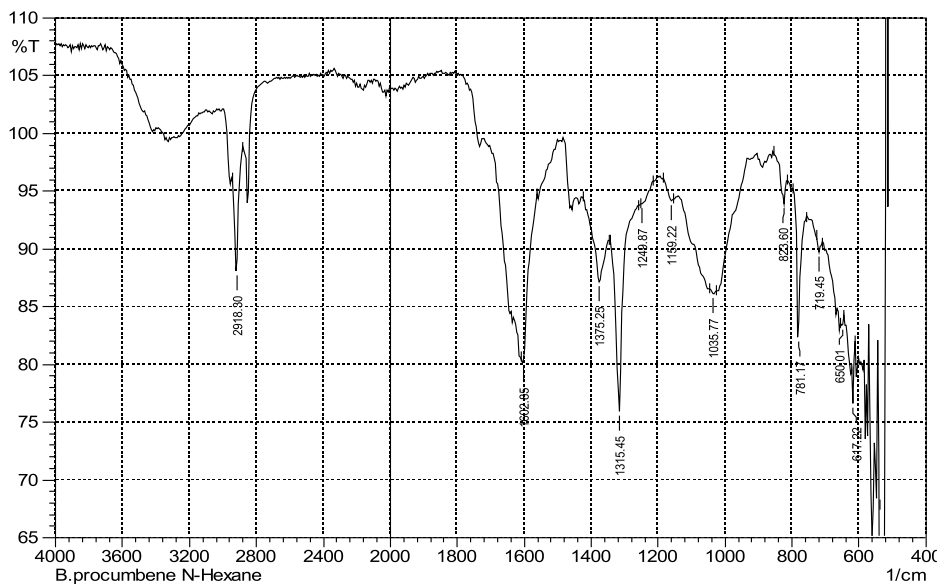


Fig. 6. FTIR analysis of n-hexane fraction of *B. procumbens*

AgNPs and transferring nanomaterials and nanoparticles through a cost-effective route.

Plants having different medically-imported bioactive components could be used to treat severe human pathogenic diseases. There are two types of categories of phytochemicals: i.e., the

primary and secondary compounds. The primary compounds contain proteins, chlorophyll, sugar, and amino acids. Secondary compounds include alkaloids and terpenoids (Wadood et al., 2013). The objective of the present study was the same as both studies investigated the phytochemical

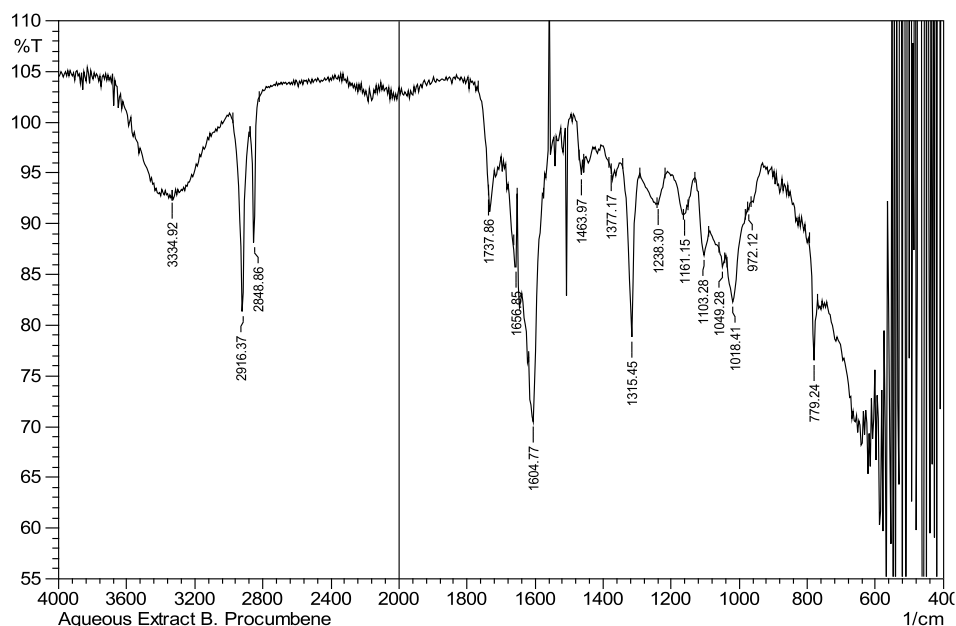


Fig. 7. Fourier Transformed Infrared Spectroscopy (FTIR) of aqueous extract

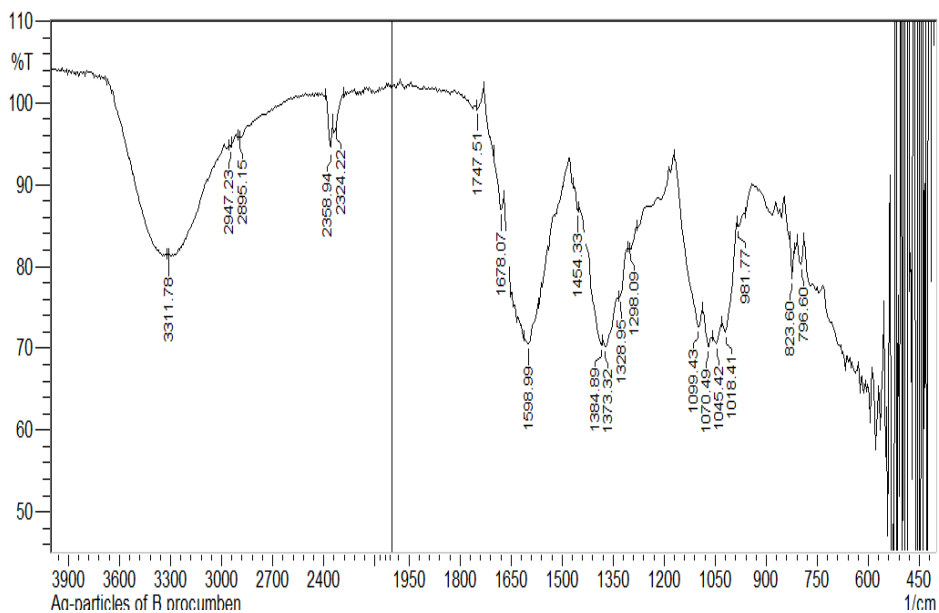


Fig. 8. FTIR of synthesized AgNPs

profile. Important phytochemicals were identified, among them are terpenoids, flavonoids, reducing sugars, and alkaloids.

These compounds serve as a plant defense against microorganisms, insects, and

herbivores. For example, phenolics are the simplest phytochemicals, which are effective against viruses (Wild, 1994), bacteria (Thomson, 1978), and fungi (Duke, 1985). Catechol and

Table 9. FTIR analysis of CHCl₃ fraction of *B. percumberns*

IR values	Bond	Functional group
2918	O-H Stretch	Carboxylic acid
1602	C=C Symmetric	Alkenes
1375	N=O Bend	Nitro group
1315	N=O Bend	Nitro group
1249	C-O Stretch	Ethers
1159	C-O Stretch	Esters
1035	C-O Stretch	Ethers

Table 10. FTIR analysis of an aqueous extract of *B. percumberns*

IR values	Bond	Functional group
3340	O-H Stretch	Phenols & Alcohols
2922	O-H Stretch	Carboxylic acid
1722	C=O Stretch	Ketones
1600	C=C Symmetric	Alkenes
1359	N=O Bend	Nitro group
1201	C-O Stretch	Ethers
1116	C-O Stretch	Ethers
1070	C-O Stretch	Ethers
1047	C-O Stretch	Ethers
1010	C-O Stretch	Ethers

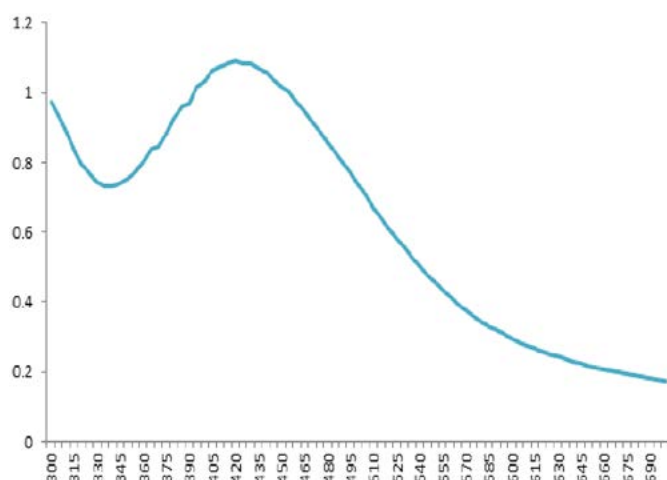


Fig. 9. UV Spectroscopy of synthesized AgNPs

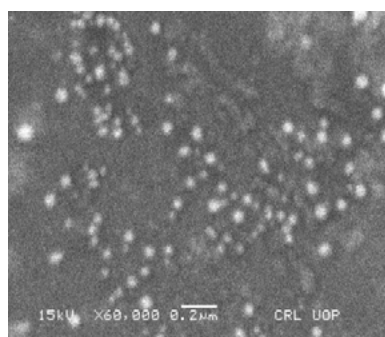


Fig. 10. Scanning electron microscope (SEM) of synthesized AgNPs

Table 11. FTIR analysis of AgNPs of *B. percumbens*

IR values	Bond	Functional group
3340	O-H Stretch	Phenols & Alcohols
2922	O-H Stretch	Carboxylic acid
1722	C=O Stretch	Ketones
1600	C=C Symmetric	Alkenes
1359	N=O Bend	Nitro group
1201	C-O Stretch	Ethers
1116	C-O Stretch	Ethers
1070	C-O Stretch	Ethers
1047	C-O Stretch	Ethers
1010	C-O Stretch	Ethers

pyrogallol are both hydroxylated phenols, toxic to microorganisms. The mechanism of action is enzyme inhibition in different ways. Flavonoids are effective antimicrobial due to their ability to complex with extracellular and soluble proteins and to complex with bacterial cell walls (Tsuchiya et al. 1996). Furthermore, quinones cause inactivation of the (Kazmi et al. 1994), tannins could inactivate microbial adhesins, enzymes, cell envelope transport proteins (Nonaka et al., 1990) and alkaloids intercalate with DNA (Phillipson and O'Neil, 1987).

The clinical trial concerning the antimicrobial significance of AgNPs was carried out with the zone of inhibition produced by pathogenic organisms (gram-negative and gram-positive bacteria) and some disease-causing fungus. (Geoprincy, 2013). The present study aimed to evaluate the antimicrobial activity of *B. procumbens*. Among extracts used, methanol and ethanol showed significant activity against targeted pathogenic bacteria and fungi. The extract of the remaining fractions did not show or showed less activity. Interestingly, the prepared AgNPs from *B. procumbens* showed better results as compared to crude extracts.

ACKNOWLEDGMENTS

The authors are grateful for the financial support provided by the Higher Education Commission of Pakistan for awarding a startup grant No (21:619/SRGP/R&D/HEC/2014). The authors extend their appreciation to the Deanship of Scientific Research at King Khalid University

for its funding this prolific research group no. (R.G.P.2.115/41).

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

AUTHORS' CONTRIBUTIONS

MR, SA, BKM, AR, MS, KU, SB, UM, YNM drafted the manuscript, compiled information from the literature, and designed the Fig. and tables. MR, SA, BKM, AR, MS, KU, SB, UM, YNM drafted the manuscript and gathered information from the literature. MR, SA, BKM, AR, MS, KU, SB, UM, YNM supervised, and reviewed the manuscript. MFR supervised and reviewed the manuscript.

FUNDING

This work was supported by the Deanship of Scientific Research at King Khalid University (Grant number R.G.P. 2/23/41/2019).

ETHICS STATEMENT

This article does not contain any studies with human participants or animals performed by any of the authors.

DATA AVAILABILITY

All datasets generated or analyzed during this study are included in the manuscript.

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