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RESEARCH ARTICLE



Immunohistochemical Expression of Xenophagy Proteins in *Helicobacter pylori* and None *Helicobacter pylori* Gastritis

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Abstract

The complex interplay between *H. pylori* and autophagy was elucidated in gastritis in term of host defense and survival of microbe. However, *H. pylori* frequently succeed in their survival and develops more aggressive tissue damage by different ways. A total of 80 gastritis patients undergoing gastroscopy during September, 2013 to August, 2014. Formalin fixed paraffin embedded gastric tissue were prepared for immunohistochemistry evaluation of ATG16L1, LC3C and vacA protein expression. Rapid urease test was used to discriminate between *H. pylori* or none *H. pylori* gastritis. Xenophagy index was elevated among both urease and IgG positive cases (29.86% and 27.13%) compared with negative cases (13.43% and 17.6% respectively (p<0.001). vacA have been found in 62.5% of urease positive cases and 55.88% of IgG positive cases. The higher xenophagy index score found to be associated with vacA positive cases (p=0.002). Xenophagy index to be associated with chronic *H. pylori* infection and this might be related to vacA positivity.

Keywords: H. pylori, Xenophagy and gastritis.

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INTRODUCTION

The Gram-negative bacteria "Helicobacter pylori" being one of the most common bacterial infections worldwide¹. *H. pylori* possess wide range of tissue pathology like: atrophic gastritis, peptic ulcer and neoplasia². Chronic gastritis represents the most frequent gastropathy depending on the intensity and persistence of bacterium³. H. pylori infection activates an inflammatory response in its host which leads to the recruitment of macrophages, neutrophils, and lymphocytes to the gastric tissue⁴. H. pylori can efficiently inhibit its own uptake by these professional phagocytes. This antiphagocytic phenotype which constitute the majority of these bacterium can reside within the mucosa or with the epithelium⁵⁻⁸, macrophages⁹, and dendritic cells (DCs)¹⁰.

Xenophagy is a specific form of autophagy that targets intracellular pathogens as cargo and makes critical contributions to both the innate and adaptive immune response to infection¹¹. In general, the induction of autophagy by pathogenic bacteria is triggered by virulence factors or bacterial components¹². Vacuolating cytotoxin-A, VacA, one of the most important virulence factors produced by H. pylori causing massive cytoplasmic vacuolation in cultured cells¹³. In vitro studies have been reported that vacA can modulate host cell activities including autophagy¹⁴. To the best of our knowledge, no clinical data have reported an association between the presence of vacA and xenophagy markers. This study aims to investigate the possible association between the presence of vacA toxin and "xenophagy index" LC3C/ATG16L1 ratio determined by immunohistochemistry.

PATIENTS AND METHODS

This cross-sectional analytical study conducted in the department Microbiology & Immunology and Pathology Department, College of Medicine, AL-Nahrain University during the period of September, 2013 to August, 2014. A total of 80 patients attending gastroenterology units in Gastroenterology and Hepatology Teaching Hospital and Al-Emamain Al-Kadhemain Medical City suffering from dyspepsia and requiring upper gastrointestinal endoscopy were included. The study was approved by College of Medicine – AL-Nahrain University institutional review board, and written informed consent was obtained from each patient.

From each patient, blood sample was taken and serum was separated and stored in - 20°C for serology (anti *H. pylori* IgM and IgG antibodies) according manufacturer instruction. Two endoscopic biopsies were taken from the lesion of stomach, one of these biopsies was processed as soon as possible for rapid urease test and the second one preserved in neutral buffered formalin for routine Hematoxylin and eosin stain and immunohistochemistry.

The tissue sections were stained by ATG16L1 (bs-4007R) and LC3C (bs-20416R) polyclonal antibodies purchased from Bioss $^{\circ}$ and visualized by LSAB System-HRP Dako (K0679). The immunoreactivity was evaluated by calculating the percentage of positive protein expression. The relative xenophagy index were calculated by dividing LC3C over ATG16L1 percentage of expression, then scoring system were expressed according to decimal intervals as the follow; score 1: \leq 0.1, score 2: 0.11-0.2, score 3: 0.21-0.3, score 4: 0.31-0.4 and so on.

Statistical analysis

The statistical analysis was done by using Graphpad PRISM [®] version 6. Crosstab model used to estimate association of allelic variant among study groups and relative risk (RR) and corresponding 95% CIs were estimated. ANOVA test were used to compare means of numerical variables between more than two groups.

RESULTS

The study subjects comprise 80 patients their mean age was 40.31 and ranged from 15-69 years old. Female were 48 patients, 21 were smokers, only 5 were alcohol drinkers, 64 patients presented with abdominal pain, 44 with nausea, 37 with vomiting. According to rapid urease test, 40 patients were *H. pylori* positive and 40 patients were considered as negative.

The serological evaluation of serum anti-*H. pylori* IgM antibody were detected in 6 patients, while, 34 patients were IgG positive.

LC3C (Xenophagy marker) is differentially expressed in gastritis

The gastric epithelial cells were evaluated for expression of LC3C among gastritis patients.

According to rapid urease results, LC3C WAS highly expressed in tissue biopsies with positive rapid urease 23.78±7.49 than those negative group 8.55±3.8 (p<0.001), while, none significant difference of LC3C molecule between patients whom IgM positive 16.83±10.55 and those with IgM negative 16.11±9.67 (p=0.861). interestingly, IgG positive cases have had statistically significant (p<0.001) LC3C expression in gastric epithelium (20.62±9.16) compared with those IgG negative (12.87±8.75). Autophagy related protein (ATG16L1) were significantly higher in expressed in epithelial cells with positive urease test (79.47±5.94) compared with urease negative group (64.73±11.35).

While, none significant difference in the mean of ATG16L1 expression between IgM positive group (69.33±16.65) and IgM negative group (68.57±11.2), also, between IgG positive group (73.32±10.71) and IgG negative group (69.46±12.18).

These results might indicate an increased expression of xenophagy related molecules in gastric epithelial cells in patients with an evidence of presence of *H. pylori* with active urease test in their tissue or who had been previously infected with the bacterium as indicated by positive serum IgG. This suggestion has been noted when we divided LC3C over ATG16L1 percentage of expression to suggest a relative xenophagy index, accordingly, this index was higher among rapid urease tissue biopsies (29.86±8.94) and IgG positive group (27.13±10.81) compared with negative urease group (13.43±5.96) and IgG negative group (17.6±9.74) p value <0.001) (Table 2).

Table 1. Patients characteristics included in the study

	Value
Age (years) mean±SD	40.31±13.85
Median (Min-Max)	39 (15-69)
Gender male (%)	32 (40)
Smoker (%)	21 (26.2)
Drinker (%)	5 (6.2)
Abdominal pain (%)	64 (80)
Nausea (%)	44 (55)
Vomiting (%)	37 (46.2)
Rapid urease test	40 (50)
IgM	6 (7.5)
IgG	34 (42.5)

Total number= 80.

		Rapid urease test		IgM		lgG	
		No (n=40)	Yes (n=40)	No (n=74)	Yes (n=6)	No (n=46)	Yes (n=34)
LC3C		8.55±3.8	23.78±7.49	16.11±9.67	16.83±10.55	12.87±8.75	20.62±9.16
P value		<0.001**		0.861 ^{NS}		<0.001**	
ATG16L1		64.73±11.35	5 79.47±5.94	68.57±11.2	69.33±16.65	69.46±12.18	73.32±10.71
P value		<0.001**		0.210 ^{NS}		0.144 ^{NS}	
LC3C/ATG16L1	ratio	13.43±5.96	29.86±8.94	21.48±11.29	23.67±10.77	17.6±9.74	27.13±10.81
(Xenophagy Ind	dex)						
	P value	<0.001**		0.649 ^{NS}		<0.001**	
	Score 1	14 (35)	0 (0)	14 (18.9)	0 (0)	12 (26.1)	2 (5.9)
	Score 2	22 (55)	3 (7.5)	22 (29.7)	3 (50)	19 (41.3)	6 (17.6)
Xenophagy	Score 3	4 (10)	20 (50)	22 (29.7)	2 (33.30)	11 (23.9)	13 (38.2)
index	Score 4	0 (0)	12 (30)	12 (16.2)	0 (0)	3 (6.5)	9 (26.5)
	Score 5	0 (0)	5 (12.5)	4 (5.4)	1 (16.7)	1 (2.2)	4 (11.8)
	P value	<(.001** 0.407 ^{NS}		107 ^{NS}	<0.001*	
vacA	Negative	39 (97.5)	15 (37.5)	52 (70.27)	2 (33.33)	39 (84.78)	15 (44.12)
	Positive	1 (2.5)	25 (62.5)	22 (29.73)	4 (66.67)	7 (15.22)	19 (55.88)
	P value	<(0.001	0.084		<0.001	

Table 2. Immunohistochemical evaluation of autophagy related proteins and xenophagy index.

NS: none statistical significance (p>0.05)

*: Statistical significance (p≤0.05).

**: Highly statistical significance (p≤0.001).

Xenophagy index is correlated with vacuolating cytotoxin expression

Furthermore, this study investigates the expression of vacA protein in gastritis cases using immunohistochemistry staining method. vacA positive cases were 32.5% (26 out of 80 patients). Furthermore, its expression found to be associated with active urease positive cases 62.5% were vacA positive among 40 urease positive cases (p<0.001) and IgG positive cases in which 55.88% were vacA positive (p<0.001). however, none significant association had been found with IgM positive cases p=0.084 (Table 2).

Further analysis of results of vacA protein expression and xenophagy score, the results showed that a statistically significant association (p=0.002). in which, higher percentage 30%, 34% of vacA positive were score 3 and 4 respectively while, in vac A negative cases a higher percentage found in were score 2 and 3 in which 38.89% and 29.63% respectively (Table 3).

DISCUSSION

The chronicity of *H. pylori* infection, until last decade had been attributed to the complex interaction between virulence factors produced by *H. pylori* like: cagA and vacA and host innate immune defense mechanisms^{5,12,15}. The current study provides an index for xenophagy process based on relative protein expression of LC3C/ ATG16L1. The measurement of antimicrobial phagocytosis process is essential to describe an innate immune mechanism against microbe (s) in pathological cases¹⁶. Here in this study we try to describe the cross-talk between *Helicobacter*

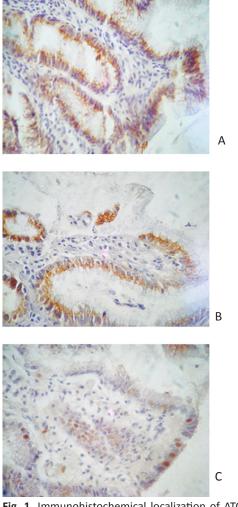


Fig. 1. Immunohistochemical localization of ATG16L1 (A), LC3C (B) and vacA (C), stained by specific primary and visualized by peroxidase staining system on gastritis tissue section. 400X original magnification.

 Table 3. Association between vacA expression and xenophagy index in gastric epithelial cells

		vacA		Total
		Positive	Negative	
	Score 1	2 (7.69)	12 (22.22)	14
	Score 2	4 (7.38)	21 (38.89)	25
Xenophagy index	Score 3	8 (30.77)	16 (29.63)	24
	Score 4	9 (34.62)	3 (5.56)	12
	Score 5	3 (11.54)	2 (3.7)	5
Total P value *: Statistical significanc	26 0.002* e (p≤0.05).	54		

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pylori and gastric epithelial expression of both LC3C and ATG16L1 molecules which are essential for complete phagosome assembly against microbe^{17,18}. The results showed that patients with higher expression of xenophagy index were

In this study, it's a first time that reports an association between the presence of vacA protein and higher index of LC3C/ATG16L1 proteins (xenophagy index) in gastric epithelial cells. The localization of these markers in gastric tissue biopsies (Fig. 1) provide an evidence for this relationship. At experimental level, studies have been reported that vacA producing H. pylori have the ability to invade AGS cell line, forming large vacuoles, which enhance long term survival of bacteria while vacA negative did not⁵. Furthermore, vacA have the ability to reduce cellular activities like: impaired production of cathepsin D that results in inability to eliminate bacteria^{5,19}, inhibition of T cell activation^{20,21}, proliferation²² and induction of apoptosis in gastric epithelial cells²³.

They found that vacA can modulate cellular autophagy process, Jones, et al., showed that vacA can promote survival in gastric epithelial cell line⁵ and macrophages²⁴. Furthermore, *H. pylori* trigger phagosome formation within the cell containing bacterial components which is similar to vacA induced vacuoles that characterized by LC3B puncta formation²⁵.

This study found an association between active urease secretion and IgG positive patients with higher expression of LC3C protein, higher scores of xenophagy index and vacA positive cases. The chronicity of gastritis was achieved by Gram negative bacterium that actively invade gastric epithelium and evasion of host defense mechanisms. The chronic vacA exposure can promote inhibition of autophagy rather than induction and elimination of bacteria(26) inducing accumulation of lysosomes and larger vacuole containing bacteria(27). Moreover, longer persistence of bacterium would results in more advanced gastro-duodenal ulceration(28) and gastric cancer(15).

Clinically, based on immunohistochemical localization of vacA and determination of xenophagy index (LC3C/ATG16L1 ratio) gives further evidence that this bacterium modulates autophagy process and can persist in gastric epithelial cells of chronic gastritis patients.

CONFLICT OF INTEREST

Authors declares no conflict of interest regarding this work.

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REFERENCES

- Moodley Y, Linz B, Bond RP, Nieuwoudt M, Soodyall H, Schlebusch CM, et al. Age of the association between *Helicobacter pylori* and man. *PLoS Pathog.* 2012;8(5).
- Alzahrani S, Lina TT, Gonzalez J, Pinchuk I V., Beswick EJ, Reyes VE. Effect of *Helicobacter pylori* on gastric epithelial cells. *World J Gastroenterol*. 2014;**20**(36):12767–80.
- Ghasemi Basir HR, Ghobakhlou M, Akbari P, Dehghan A, Seif Rabiei MA. Correlation between the intensity of *Helicobacter pylori* colonization and severity of gastritis. *Gastroenterol Res Pract*. 2017;**2017**:10–5.
- Wang Y-H, Gorvel J-P, Chu Y-T, Wu J-J, Lei H-Y. Helicobacter pylori Impairs Murine Dendritic Cell Responses to Infection. PLoS One [Internet]. 2010;5(5):e10844. Available from: http:// dx.plos.org/10.1371/journal.pone.0010844
- Terebiznik MR, Vazquez CL, Torbicki K, Banks D, Wang T, Hong W, et al. *Helicobacter pylori* VacA toxin promotes bacterial intracellular survival in gastric epithelial cells. *Infect Immun*. 2006;**74**(12):6599–614.
- Chu Y-T, Wang Y-H, Wu J-J, Lei H-Y. Invasion and multiplication of *Helicobacter pylori* in gastric epithelial cells and implications for antibiotic resistance. *Infect Immun*. 2010;**78**(10):4157–65.
- Ricci V, Romano M, Boquet P. Molecular cross-talk between *Helicobacter pylori* and human gastric mucosa. *World J Gastroenterol*. 2011;17(11):1383–99.
- Tang B, Li N, Gu J, Zhuang Y, Li Q, Wang HG, et al. Compromised autophagy by MIR30B benefits the intracellular survival of *Helicobacter pylori*. *Autophagy*. 2012;8(7):1045–57.
- Wang Y-H, Wu J-J, Lei H-Y. The autophagic induction in *Helicobacter pylori*-infected macrophage. *Exp Biol Med* (Maywood). 2009;**234**(2):171–80.

- Wang Y-H, Gorvel J-P, Chu Y-T, Wu J-J, Lei H-Y. *Helicobacter pylori* impairs murine dendritic cell responses to infection. *PLoS One*. 2010;5(5):e10844.
- Levine B, Mizushima N, Virgin HW. Autophagy in immunity and inflammation. *Nature* [Internet].
 2011 Jan 20 [cited 2018 May 7];469(7330):323– 35. Available from: http://www.nature.com/ articles/nature09782
- Cemma M, Brumell JHH. Interactions of Pathogenic Bacteria with Autophagy Systems. *Curr Biol* [Internet]. 2012;**22**(13):R540–5. Available from: http://dx.doi.org/10.1016/j. cub.2012.06.001
- Foegeding NJ, Caston RR, McClain MS, Ohi MD, Cover TL. An overview of *Helicobacter pylori* VacA toxin biology. *Toxins* (Basel). 2016;8(6):1– 21.
- 14. Terebiznik MR, Raju D, Vázquez CL, Torbricki K, Kulkarni R, Blanke SR, et al. Effect of *Helicobacter pylori's* vacuolating cytotoxin on the autophagy pathway in gastric epithelial cells. *Autophagy*. 2009;**5**(3):370–9.
- Ki MR, Hwang M, Kim AY, Lee EM, Lee EJ, Lee MM, et al. Role of vacuolating cytotoxin VacA and cytotoxin-associated antigen CagA of *Helicobacter pylori* in the progression of gastric cancer. *Mol Cell Biochem*. 2014;**396**(1–2):23–32.
- Kwon DH, Song HK. A Structural View of Xenophagy, A Battle between Host and Microbes. 2018;8(1):27–34.
- Mao K, Klionsky DJ. Xenophagy: A battlefield between host and microbe, and a possible avenue for cancer treatment. *Autophagy* [Internet]. 2017;13(2):223–4. Available from: http://dx.doi.org/10.1080/15548627.2016.12 67075
- Shpilka T, Elazar Z. Essential Role for the Mammalian ATG8 Isoform LC3C in Xenophagy. *Mol Cell* [Internet]. 2012;48(3):325-6. Available from: http://dx.doi.org/10.1016/j. molcel.2012.10.020
- Necchi V, Sommi P, Vanoli A, Fiocca R, Ricci V, Solcia E. Natural history of *Helicobacter pylori* VacA toxin in human gastric epithelium in vivo: Vacuoles and beyond. *Sci Rep* [Internet]. 2017;7(1):1–11. Available from: http://dx.doi. org/10.1038/s41598-017-15204-z
- 20. Gebert B, Fischer W, Weiss E, Hoffman R, Haas R. *Helicobacter pylori* Vacuolating Cytotoxin

Inhibits T Lymphocyte Activation. *Science* (80-) [Internet]. 2003;**301**(5636):1099–102. Available from: http://www.sciencemag.org/ cgi/doi/10.1126/science.1086871

- Boncristiano M, Paccani SR, Barone S, Ulivieri C, Patrussi L, Ilver D, et al. The *Helicobacter pylori* Vacuolating Toxin Inhibits T Cell Activation by Two Independent Mechanisms. J Exp Med [Internet]. 2003;**198**(12):1887–97. Available from: http://www.jem.org/lookup/doi/10.1084/ jem.20030621
- Torres VJ, VanCompernolle SE, Sundrud MS, Unutmaz D, Cover TL. *Helicobacter pylori* vacuolating cytotoxin inhibits activation-induced proliferation of human T and B lymphocyte subsets. *J Immunol* [Internet]. 2007;**179**(8):5433– 40. Available from: http://www.jimmunol.org/ cgi/content/abstract/179/8/5433
- 23. Zhu P, Xue J, Zhang Z, Jia Y, Tong Y, Han D, et al. Helicobacter pylori VacA induces autophagic cell death in gastric epithelial cells via the endoplasmic reticulum stress pathway. Cell Death Dis [Internet]. 2017;8(12):3207. Available from: http://www.nature.com/articles/s41419-017-0011-x
- 24. Zheng PY, Jones NL. *Helicobacter pylori* strains expressing the vacuolating cytotoxin interrupt phagosome maturation in macrophages by recruiting and retaining TACO (coronin 1) protein. *Cell Microbiol*. 2003;**5**(1):25–40.
- 25. Terebiznik MR, Raju D, Vázquez CL, Torbricki K, Kulkarni R, Blanke SR, et al. Effect of *Helicobacter pylori*'s vacuolating cytotoxin on the autophagy pathway in gastric epithelial cells. *Autophagy*. 2009;**5**(3):370–9.
- Oldani A, Cormont M, Hofman V, Chiozzi V, Oregioni O, Canonici A, et al. *Helicobacter pylori* counteracts the apoptotic action of its VacA toxin by injecting the CagA protein into gastric epithelial cells. *PLoS Pathog*. 2009;5(10).
- 27. Raju D, Hussey S, Ang M, Terebiznik MR, Sibony M, Galindo-Mata E, et al. Vacuolating cytotoxin and variants in Atg16L1 that disrupt autophagy promote *Helicobacter pylori* infection in humans. *Gastroenterology*. 2012;**142**(5):1160–71.
- Wada A, Yamasaki E, Hirayama T. *Helicobacter* pylori vacuolating cytotoxin, VacA, is responsible for gastric ulceration. *J Biochem*. 2004;**136**(6):741–6.