

Evaluation of Chemicals and Bioagents for Managing *Phytophthora* Root Rot and Root-knot Nematode Disease Complex in Capsicum under Naturally Ventilated Polyhouse

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Capsicum is an important vegetable crop of Karnataka. Among the various diseases affecting capsicum, *Phytophthora* root rot (*Phytophthora capsici*) and root-knot nematode (*Meloidogyne incognita*) infestation are considered as the most destructive under protected cultivation. Totally six fungicides and seven bioagents were screened against *P. capsici* under *in vitro*. Among the fungicides, Bordeaux mixture and Metalaxyl MZ were highly inhibitory to *P. capsici*. Among the antagonists, *Trichoderma* were most effective in inhibiting the growth of *P. capsici*. Least inhibition was noticed with *Bacillus* sp followed by *Pseudomonas*. Effect of various disease management tools on seedling emergence, *Phytophthora* root rot incidence and root knot index were studied. As part of integrated management strategies an investigation was carried out to manage the disease through soil application of fungicide, nematicide and bioagents. Metalaxyl MZ significantly reduced the *Phytophthora* root rot incidence of 69.23% disease reduction over control under polyhouse condition. Integration of Carbofuran + *P. lilacinus* (0.33) also proved superior in managing root knot disease over control.

Keywords: *Phytophthora capsici*, *Meloidogyne incognita*, *Capsicum annuum*, Management.

Capsicum (*Capsicum annuum* L.) is infected by a number of diseases, among them wilt complex has gained major importance in India and the losses caused by disease are up to the extent of 30 to 40 per cent. The wilt disease, incited by a number of pathogens, is the devastating soil-borne disease and hence it is difficult to manage. The disease has been observed to be caused by *Phytophthora capsici* and Root-knot nematode, *Meloidogyne incognita*. Keeping in view the etiology, exorbitant losses and lack of information regarding integrated management of wilt complex disease using bio-control agents and chemicals, the present studies were conducted to evolve the effective management strategies under naturally ventilated polyhouse condition. The study was

carried out in Zonal Agricultural and Horticultural Research Station, UAHS, Shivamogga, Karnataka.

MATERIALS AND METHODS

The fungus, *Phytophthora capsici* was isolated from capsicum plants affected with wilt complex disease. The pure cultures of pathogen were maintained on potato dextrose agar media at 4°C. Root-knot nematode was extracted from the wilt affected soil and also from affected root galls.

In-vitro Evaluation of Fungicides against *Phytophthora capsici*

The efficacy of fungicides *viz.* Metalaxyl MZ, Alliete, Carbendazim, Mancozeb, Bordeaux mixture and Avatar were tested at three different concentrations 0.05%, 0.075%, 0.1% against each pathogen under *in-vitro* condition by poisoned food technique to find out the effective fungicides. The petri-plates containing Potato dextrose agar

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media amended with desired concentration of fungicides were inoculated with 5 mm discs of pathogen and incubated with $25\pm 1^\circ\text{C}$ temperature. The plates without any fungicides served as check. The radial mycelial growth was recorded in each treatment and percent inhibition over check was calculated.

In-vitro* Evaluation of bioagents against *Phytophthora capsici

Seven Bioagents viz., *Bacillus subtilis*, *Pseudomonas fluorescens*, *Paecilomyces lilacinus*, *Trichoderma harzianum*, *T. viride*, *T. koningii* and *T. virens* were evaluated for their efficacy through dual culture technique. The bioagents and the test fungus were inoculated side by side on a single petridish containing solidified Potato Dextrose Agar. Five replications were maintained for each treatment with one control by maintaining only pathogen separately. The diameter of the colony of both bioagents and the pathogen was measured in two directions and average was recorded. The per cent inhibition of growth of the test fungus was calculated

Evaluation of chemicals and bioagents against capsicum wilt complex under polyhouse condition

The fungicides found effective *in vitro* were tested *in vivo* as soil application to the plant. The studies were carried out in polyhouse. Treatments were imposed to the beds before sowing viz., Metalaxyl MZ (0.1%), Copper oxychloride (0.1%), Carbofuran (10g/plant), *P. lilacinus* (20g/plant), *T. harzianum* (20g/plant), *P.*

fluorescens (20g/plant), Metalaxyl MZ (0.1%) + Carbofuran (10g/plant), Metalaxyl MZ (0.1%) + *P. lilacinus* (20g/plant), Metalaxyl MZ (0.1%) + *T. harzianum* (20g/plant), Metalaxyl MZ (0.1%) + *P. fluorescens* (20g/plant) and Carbofuran (20g/plant) + *P. lilacinus* (20g/plant). The antagonists (20 g each) were properly mixed with 1 kg FYM and then mixed with the top soil layer of plot. The observations on wilt incidence and RKI were recorded in each treatment.

RESULTS AND DISCUSSION

In vitro* evaluation of fungicides against *P. capsici

The present investigation in Table 1 revealed that cent per cent inhibition of mycelial growth of *P. capsici* was recorded with Bordeaux mixture and Metalaxyl MZ at all tested concentrations which are followed by Avatar (75.89%) and Alliete (72.30%) at 0.1%. Least inhibition of 55.93 and 55.03 per cent was recorded in Bavistin and Mancozeb respectively. Similar observations have been recorded by Babadoost and Islam² with respect to metalaxyl MZ as highly inhibitory to the mycelial growth of *P. capsici* under *in-vitro* conditions³. Bordeaux mixture (1%), Blitox and Metalaxyl MZ which were found effective in inhibiting the growth and sporangial formation of *P. capsici* and *P. meadii*^{13,14,16}.

In vitro* evaluation of bioagents against *P. capsici

Efficacy of bioagents was studied under *in vitro* conditions by following dual culture

Table 1. *In vitro* evaluation of fungicides against *Phytophthora capsici*

Fungicides	Percent inhibition				
	Concentration (%)				
Common name	Trade name	0.05	0.075	0.1	Mean
Carbendazim	Bavistin 50 WP	55.75 (48.30) *	55.93 (48.41)	55.93(48.41)	55.87 (48.37)
Aliette	Fosetyl 80 WP	100.00 (90.05)	100.00 (90.05)	100.00 (90.05)	100.00 (90.05)
Metalaxyl MZ	Ridomil 72WP	100.00 (90.05)	100.00 (90.05)	100.00 (90.05)	100.00 (90.05)
Hexaconazole 4%+ Zineb 68%	Avatar72 WP	58.99 (50.18)	71.94 (58.01)	75.89 (60.59)	68.94 (56.13)
Dithane M45	Mancozeb 75WP	58.99 (50.18)	44.42 (41.80)	55.03 (47.89)	52.81 (46.61)
Bordeaux mixture	-	55.03 (47.89)	61.69 (51.76)	72.30 (58.24)	63.00 (52.54)
Mean	71.46 (57.71)	72.33 (58.26)	76.52 (61.02)	73.43 (58.97)	
	Fungicides (F)	Concentration (C)F × C			
S.Em. ±	0.31	0.22	0.55		
C.D @ 1 %	0.84	0.67	1.46		

*Figures in the parenthesis are arcsine transformed values

Table 2. *In vitro* evaluation of bioagents against *Phytophthora capsici*

Bioagents	Per cent inhibition
<i>Paecilomyces lilacinus</i>	44.63 (41.94)*
<i>Trichoderma harzianum</i>	100.00 (90.05)
<i>T. viride</i>	53.70 (47.15)
<i>T. virens</i>	58.15 (49.71)
<i>T. koningii</i>	48.15 (43.96)
<i>Pseudomonas fluorescens</i>	57.86 (51.87)
<i>Bacillus subtilis</i>	45.19 (42.26)
SEm±	0.28
CD@1%	1.17

*Figures in the parenthesis are arcsine transformed values

method against *P. capsici* and the results are presented in Table 2. There was significant difference among all the tested bioagents. *Trichoderma harzianum* (100%) was significantly superior in inhibiting the mycelial growth of *P. capsici* followed by *T. virens* (58.15%), *P. fluorescens* (57.86%) and *T. viride* (53.70%). Further, *T. koningii* (48.15%), *Bacillus subtilis* (45.19%) and *P. lilacinus* (44.63%) remained on par with each other in inhibition. Inhibition of mycelial growth was recorded in *P. fluorescens* (43.81%). This has been reported by many workers^{8,10}. The antagonism of *Trichoderma* spp. against many fungi is mainly due to production of

Table 3. Evaluation of Chemicals and Bioagents against *Phytophthora capsici* and *Meloidogyne incognita*

S. No.	Treatments	% seedling emergence	% Increase over Control	% disease incidence	% disease reduction over control	Nematode Population		RKI (0-5)
						Initial	Final	
T1	Metalaxyl MZ	74.93	39.62	18.02 (25.13)*	69.23	905.33	815.67	4.00
T2	COC	72.67	35.40	25.50 (30.35)	56.45	881.40	791.40	4.00
T3	Carbofuran	76.93	43.35	37.40 (37.72)	36.13	757.00	466.67	2.67
T4	<i>P. lilacinus</i>	76.00	41.61	27.07 (31.37)	53.78	610.00	220.00	1.00
T5	<i>T. harzianum</i>	77.67	44.71	21.29 (27.49)	63.65	655.60	345.60	1.67
T6	<i>P. fluorescens</i>	78.89	46.98	27.43 (31.60)	53.15	745.17	435.17	2.00
T7	T1+T3	76.83	43.16	18.22 (25.28)	68.89	720.00	258.33	2.67
T8	T1+T4	76.67	42.85	20.31 (26.80)	65.31	606.00	294.67	1.33
T9	T1+T5	75.55	40.77	16.20 (23.75)	72.34	660.77	350.33	1.67
T10	T1+T6	73.80	37.51	20.07 (26.63)	65.73	737.00	427.00	2.00
T11	T3+T4	73.33	36.64	35.77 (36.75)	38.92	650.00	205.00	0.33
T12	Control	53.67	-	58.40 (49.86)	-	920.67	975.33	5.00
SEm±0.19		-	1.42	-	-		0.22	
CD@5%		0.56	-	4.20	-			0.65

*Figures in the parenthesis are arcsine transformed values

acetaldehyde¹³. This may also be the reason for antagonistic effect of native isolates of *Trichoderma* against *P. capsici*. The antagonistic nature of *Bacillus subtilis* and *Pseudomonas* sp⁶. Similar results were observed in the efficacy of *Trichoderma* spp. and *Pseudomonas* spp. against the pathogen *P. capsici*^{1, 2, 11}.

Evaluation of chemicals and bioagents against capsicum wilt complex under polyhouse condition

The data presented in Table 3 also revealed that, highest percent seedling emergence over control were recorded in *P. fluorescens* (46.98%), *T. harzianum* (44.71%), Carbofuran (43.35%) and Metalaxyl MZ + Carbofuran (43.16%) treatments respectively. Further, Metalaxyl MZ

(69.23), Metalaxyl MZ + Carbofuran (68.89%), Metalaxyl MZ + *P. fluorescens* (65.73%) and Metalaxyl MZ + *P. lilacinus* (65.31%) showed higher disease reduction over control respectively. The integration of biocontrol agents with Metalaxyl MZ resulted lesser disease incidence. Integration of Carbofuran + *P. lilacinus* (0.33) recorded less gall index of 0.33 followed by *P. lilacinus* (1.00). Further, *T. harzianum* and Metalaxyl MZ + *T. harzianum* showed 1.67 gall index compared to control (5.00 RKI). The present findings are supported by earlier works wherein integration of bio-control agents with fungicides and nematicides gave significantly higher disease control in several crops than that obtained either

by bio-control agent or by fungicide or by nematicide alone^{9,14,15}. Treatment with Metalaxyl MZ may eradicate the wilt causing pathogens or other microflora thereby less competition for biocontrol agents to colonize the seed, root surface and proliferation^{5, 11}. The subsequent application of compatible fungicide/ nematicide may support the growth of antagonists and may also prevent the plant from the attack of pathogens. The other workers also observed the additive effects of compatible fungicide/nematicide and antagonists⁴. Integrated *T. viride* or *T. harzianum* with Metalaxyl MZ and Cymoxanil (8%) + Mancozeb (64%) to control root rot in cotton⁷. Results from the present investigation concluded that, soil application of Metalaxyl MZ or Carbofuran+ bioagents applied simultaneously to manage the root gall and *Phytophthora* rot in Capsicum.

REFERENCES

1. Anandaraj, M., Sarma, Y. R. and Venugopal, M. N., Biological control of foot rot of black pepper. In: *Biological Control of Diseases of Spices* Eds. Anandraj, M. and Peter, Pub. Indian Institute of Spice Research, Calicut, 1995, p.120.
2. Anith, K. N. and Manomohandas, T. P., Combined application of *Trichoderma harzianum* and *Alcaligenes* sp. strain AMB 8 for controlling nursery rot disease of black pepper. *Indian Phytopath.*, 2001; **54**: 335-339.
3. Babadoost, M. and Islam, S. Z., Fungicide seed treatment effects on seedling damping- Off of pumpkin caused by *Phytophthora capsici*. *Plant Disease*, 2003; **87**(1): 65-68.
4. Chattopadhyay, C., and Sen, B., Integrated management of *Fusarium* wilt of Muskmelon caused by *Fusarium oxysporum*. *Indian J. Mycol. Pl. Pathol.*, 1996; **26**(2): 162-170.
5. Chet, I. Elad, Y., Koflan, A., Hadar, Y., and Katan, J., Integrated control of soil borne pathogen in Iris. *Phytoparasitica*, 1982; **10**: 229-231.
6. Filippi, C., Bagnoli, G. and Pices, G., Antagonistic effect of soil bacteria on *Fusarium oxysporum* f.sp. *dianthi*. *Agril. Mediterranea*, 1989; **119**: 327-336.
7. Gaur, R. B. and Sharma R. N., Biocontrol of Root Rot in Cotton and Compatibility of Potential Bioagents with fungicides. *Indian J. Pl. Protec.*, 2010; **38**(2): 176-182.
8. Ghaffar, A., Biological control of white rot of onion. Interaction effect of soil microorganisms with *Sclerotium cepivorum* Berk. *Mycopathology. Myco. Appl.*, 1969; **38**: 101-111.
9. Jayalakshmi, K., Ravindra, H., Soumya D., Narasimhamurthy, H. B., Nagarajappa Adivappar, and Raju J., Evaluation of chemicals for managing *Phytophthora capsici* and *Meloidogyne incognita* disease complex in black pepper under polyhouse condition. *Paper in 3rd Intl. Symposium: Phytophthora: Taxonomy, genomics, Pathogenecity, Resistance and disease management 9 Sep Bengaluru, India.* 2015; 56p.
10. Naik, M. K. and Sen, B., Biocontrol of plant disease caused by *Fusarium* spp. In: *Recent Development in Biocontrol of Plant Diseases*, Eds. Mukerjee, K.G., Tiwari, J.P. and Arora, D.K., Aditya Books, New Delhi, 1995; 37-51.
11. Rajan, P. P., Sarma, Y. R. and Anadaraj, M., Management of foot rot disease of black pepper with *Trichoderma* sp. *Indian Phytopath.*, 2002; **55**: 34-38.
12. Ram, D., Mathur, K., Lodha B. C. and Webster, J., Evaluation of resident biocontrol agents as seed treatments against ginger rhizome rot. *Indian Phytopath.*, 2000; **53**(4): 450-454.
13. Ramachandran, N., Sarma, Y. R. and Anandaraj, M., Management of *Phytophthora* infection in black pepper. In: *Proc. of Int. Pepper Community Workshop. Joint Res. Country. Black Pepper Disease.* 27-29, October, 1988, National Research Centre for Spices, Calicut, India, 1990; 158-174.
14. Raju J., Nagarajappa Adivappar, Jayalakshmi, K., Ravindra, H., Narasimhamurthy, H. B., and Soumya D., Evaluation of chemicals and bioagents for managing *Phytophthora* root rot and root-knot nematode disease complex in capsicum under protected cultivation. *Paper in 3rd Intl. Symposium: Phytophthora: Taxonomy, genomics, Pathogenecity, Resistance and disease management 9 Sep Bengaluru, India.* 2015; 56.
15. Robinson, P. M. and Park, D., Volatile inhibitor spore germination produced by fungi. *Trans British Mycological Society*, 1966; **49**: 639-649.
16. Sastry, M. N. L., Studies on species of *Phytophthora* affecting plantation crops in Karnataka with special reference of Koleroga of arecanut and black pepper. *Ph.D.Thesis*, Univ. Agric. Sci., Bangalore (India) 1982.
17. Sawant, I. S. and Mudhopadhyay, A. N., Integration of metaxyl with *Trichoderma harzianum* for the control of Pythium damping-off in sugar beet. *Indian Phytopath.*, 1990; **43**: 535-541.
18. Veena, S. S. and Sarma, Y. R., Uptake and persistence of potassium phosphonate and its protection against *Phytophthora capsici* in black pepper. *Centennial Conference on Spices and Aromatic Plants*, Calicut, Kerala, 20-23 September, pp. 2000; 243-248.

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