

Antibacterial Study of Gellan Gum (GG) Film Incorporated Norfloxacin

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Abstract

A series of norfloxacin-loaded films were fabricated by combining with gellan gum (GG) using evaporative casting technique. The films were evaluated in terms of morphological, water vapour transmission rates (WVTR), water uptake, release study and antibacterial study. The prepared films contained smooth appearance of cross-section distributed by good film of materials. The results indicated that the film swelling were increased at higher concentration of norfloxacin loaded, however decreased the WVTR properties. All films showed almost 100 % of drug release within 5 - 20 minutes. It was found that, the antibacterial activity was directly proportional to the release rate which is at higher concentration of norfloxacin resulted in stronger antibacterial properties.

Keywords: Gellan gum; Film; Antibacterial.

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INTRODUCTION

Gellan gum (GG) is a microbial exopolysaccharides produced from *Sphingo-monas* group and holds a vital role in therapeutic and manufacturing. It has been broadly used as stabilizers, thickening and gelling agent, emulsifiers and etc.¹ in food industry since it has been approved by US FDA (United States Food and Drug Administration) and EU (European Union) regulation labelled as E 415 for food applications². GG are among of the biopolymers that are currently used for commercial product and have been subjected for extensive study³. It was attract attention among the researcher since 1980 regarding to their unique structure and rheology properties that can form pseudo-solid structures or transparent hydrogelen at low concentrations of their usage⁴. Not limited to that, it is a multi-functional gelling agent⁵, and has high potential to be apply in industrial as a high-viscosity biogum, a suspending agent and acts as a gelling agent⁶. Previously, GG has been use in development of scaffold material in tissue engineering application⁷, as a vehicle for ophthalmic drugs⁸, and in eye drops⁹. It is also reported to biocompatible on live cells *eg.* mouse fibroblast (L929 cell line)¹⁰, human dermal fibroblasts (HDFs), human fetal osteoblasts (hFOBs 1.19)¹¹, human nasal cartilage¹², rat bone marrow cells (rBMC) [7] and has been used in treating defects for rabbit knee joints¹³. Besides that, GG also presented their potential for wound dressing application since it is favorable for cell proliferation¹⁴⁻¹⁶. The gellan gum ability also has been used in production of an easy-to-swallow solid dosage forms likes gels and coated tablets. This is due to the release rate of active ingredients inside the tablets and capsules can be modified by using the gellan gum. So, it is favorable to use in controlled and sustained the releasing of variety types of drugs^{1,17} as well as for microencapsulation preparation. In this study, norfloxacin were selected as an antibiotic model due to its strong antibacterial properties. Norfloxacin is a fluoroquinolone with a broad-spectrum activity against gram-positive and gram-negative bacteria, including *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Escherichia coli*, *Salmonella*, *Campylobacter*, *Vibrio cholera* and *Shigella*¹⁸⁻²⁰. Norfloxacin effectively execute gram-positive and gram-negative bacteria, however it

has limitation against anaerobes bacteria²¹. Since few years ago, norfloxacin was become subject of interest for drug delivery study especially in biological and pharmaceutical field²²⁻²⁵. Norfloxacin has been used in treated common and complicated urinary tract infections such as cystitis and prostatitis^{26,27} as well as effectively in treated gonococcal urethritis and infectious diarrhea²⁸. Currently, it was effectively used as antibacterial in wound dressing materials^{29,30}. So, in this work, norfloxacin were incorporated into gellan gum to develop medicated film for wound dressing application. This medicated film was importance to enhance the healing process by controlling the moisture around the wound area as well as to protect the skin from infections and dehydration that could be led to the tissue damaging and producing scar.

MATERIALS AND METHODS

Low-acyl gellan gum (Gelzan™ CM, Mw H^o 2-3 x 10⁵ Da, product number-G1910, lot number SLBB0374V) was purchased from Sigma-Aldrich, Malaysia. Norfloxacin (product number-N9890) used in this work was supplied from Fluka, USA. All materials were used as received without further purification and commercially available.

Preparation of Film

Gellan gum (GG) solution was prepared by dissolving 1% (w/v) of GG in 100 mL deionized water (18.2 MΩ) with continuous stirring for 2 h at 70°C. GG-NOR solutions were made by adding 0.01%, 0.1%, 0.5% and 1% (w/w) norfloxacin (NOR) into the GG solution. Then, the GG and GG-NOR solutions were deposited into petri dishes (90 mm x 15 mm) and were put in Venticell oven at 30°C for 24 h. All films were pre-conditioned in a desiccator (24°C, 50 % relative humidity (RH)) for at least 2 days prior to testing.

Characterization of Film

The Ultraviolet (UV-vis) absorption spectra of solutions and films were collected via spectrophotometer (Varian, Cary 50) with data interval = 5 nm, scan rate = 24 000 nm/min at 200-800 nm of wavelength. Quartz cells of path length at 10 mm were applied. Scanning electron microscopy (SEM) images were obtained by using a JOEL JSM 6360 LA electron microscope. SEM images of cross-sections were obtained by freeze-drying the samples in liquid nitrogen (-160°C) and

were fractured at -150°C . Prior to observe via SEM, the samples were coated with Auto Fine Coats (JFC-1600). Swelling performance was evaluated by weighing the dried films (W_{dry}) before soak into solutions at pH 1, pH 7, and pH 12 in water bath ($37 \pm 0.5^{\circ}\text{C}$). Then, after 24 h, prior to weighed (W_{wet}) the films were removed from the solutions and were wiped gently with a tissue to expel any solution on the film surfaces. Swelling percentage was calculated from the equilibrium-swelling ratio ($(W_{\text{wet}} - W_{\text{dry}})/W_{\text{dry}}$). The water vapor transmission rates (WVTR) was determined based on ASTM International standard method by some of the modification. The films were fixed on the circular opening of a permeation bottle with diameter at 1.5 cm, height at 5.0 cm, and effective transfer area (A) at 1.33 cm^2 and placed in a desiccator (25°C and $50 \pm 5\% \text{ RH}$) for controlled the environment moisture. The WVTR was then measured by determined the rate of change of mass (m) in these water-filled permeation bottles at 24 h exposure time of 24 h using Equation 1:

$$\text{WVTR} = (m / A \Delta t) \quad \dots(1)$$

where $m/\Delta t$ is water gain per unit time of transfer and A is the area exposed to water transfer (m^2).

Antibacterial Assay

The antibacterial activity was tested using Gram-positive (*Staphylococcus aureus* - *S. aureus*) and gram-negative (*Escherichia coli* - *E. coli*) bacteria. The standard growth medium of Muller-Hinton, (MH) Difco™ agar was sterilized using autoclave at 120°C for 15 min. The bacteria were grown in MH agar and incubated aerobically at 37°C for 24 h. The suspensions were measured by using spectrophotometer (Biomerieux Densicheck

Plus, USA) at 600 nm with optical density at 0.5. The antibacterial activity of GG and GG-NOR film samples were tested against Gram-positive (*Staphylococcus aureus* - *S. aureus*) and gram-negative (*Escherichia coli* - *E. coli*) bacteria. The assessment was conducted based on the disc diffusion method. Inoculants of both *S. aureus* and *E. coli* were spread using sterile cotton swab over the surface of the agars in sterile petri plates. Penicillin was used as a control for the bacteria. Each of the specimens and controlled drugs were placed on the standard growth medium of Muller-Hinton, MH, Difco™ agar in sterilized petri plates and then incubated for 24 h at 37°C in triplicates. Prior to gently press of the films in 6 mm of their diameter, the plates that contained both of the bacteria were let to dry. After 24 h of aerobically incubation, the plates were examined for the presences of clear zone around the disc of specimens were then photograph using Samsung 13-megapixel Samsung camera for further evaluation.

RESULTS AND DISCUSSION

UV-visible spectroscopy of gellan gum (GG) with norfloxacin films at different concentrations are shown in Fig. 1. An absorption peak for free-standing GG was observed at 260 nm due to the presence of glucuronic acid group³¹⁻³². The pure norfloxacin band at 272 nm was shifted in GG-NOR blends ($\geq 0.1\%$ of norfloxacin added) to 280 nm, indicating an interaction between both polymers³³⁻³⁵. The absorption intensity of the peak was increased depending on the concentration of norfloxacin included in the GG film. The UV-vis absorption peak of GG-NOR001 which contains the

Table 1. The water vapour transmission rates (WVTR) and swelling properties of GG and GG-NOR films (mean \pm SD) (n=3)

Sample	WVTR		Swelling (%)	
	($\text{g m}^{-2} \text{ d}^{-1}$)	pH1	pH7	pH12
GG	422 \pm 113	308 \pm 15	987 \pm 113	724 \pm 11
GG-NOR001	320 \pm 35	310 \pm 12	1199 \pm 55	814 \pm 37
GG-NOR01	353 \pm 2	307 \pm 20	1123 \pm 47	791 \pm 24
GG-NOR05	344 \pm 63	281 \pm 18	1219 \pm 81	809 \pm 44
GG-NOR1	337 \pm 7	263 \pm 16	1284 \pm 18	859 \pm 30

Table 2. Inhibition zone of gellan gum and gellan gum incorporated with norfloxacin against *Staphylococcus aureus* and *Escherichia coli* (mean \pm SD) (n=3)

Test bacteria	Inhibition zone (mm)	
	<i>S.aureus</i>	<i>E.coli</i>
Material		
GG	-	-
Penicillin	-	-
GG-NOR001	5.3 \pm 0.06	5.0 \pm 0.01
GG-NOR01	11.0 \pm 0.06	11.3 \pm 0.06
GG-NOR05	12.7 \pm 0.12	13.0 \pm 0.10
GG-NOR1	13.0 \pm 0.01	14.3 \pm 0.06

lowest content of norfloxacin at 275 nm shifted to longer wavelength for GG-NOR05 and GG-NOR1 to 280 nm with increased absorption indicating the increase in cross-linking network of the composite. From the scanning electron microscopy photographs as shown in Fig. 2, smooth cross-section of GG-NOR films were observed which shows that the norfloxacin was distributed well in

GG solution. In addition, the formation of smooth cross-section is caused by the good film forming property³⁶⁻³⁸ between gellan gum and norfloxacin.

In Table 1, the water vapour transmission rate (WVTR) values are shown for gellan gum (GG) and GG incorporated with norfloxacin (NOR). The WVTR values were decreased with addition of norfloxacin into the gellan gum film. The water

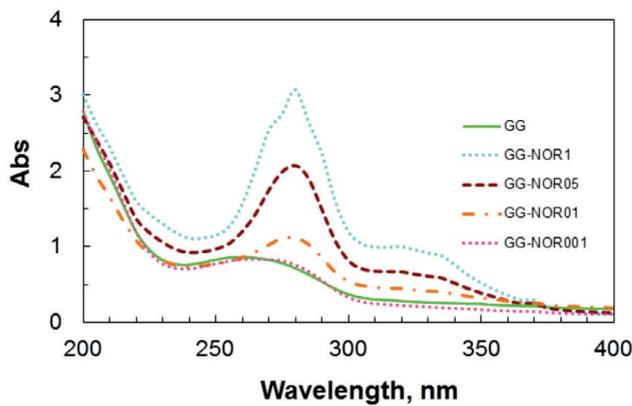


Fig. 1. UV-Visible absorbance of pure GG and GG-NOR films.

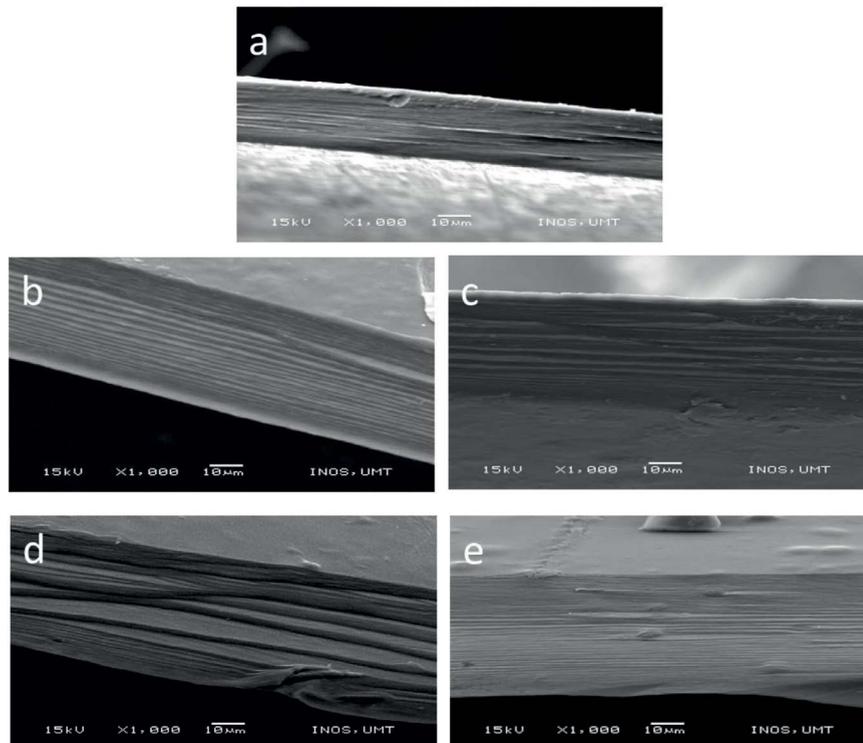


Fig. 2. Scanning electron microscopy images of cross-sectional area of (a) GG, (b) GG-NOR001, (c) GG-NOR01, (d) GG-NOR05, and (e) GG-NOR1 films.

vapour transmission rates of GG-NOR001 have decreased about 24% compared to GG. Further, the water vapour transmission rates of GG-NOR01, GG-NOR05, and GG-NOR1 were decreased to

353±2 g m⁻² d⁻¹, 344±63 g m⁻² d⁻¹, and 337±7 g m⁻² d⁻¹, respectively compared to GG films.

Table 1 presents the swelling properties of gellan gum and gellan gum inclusion with

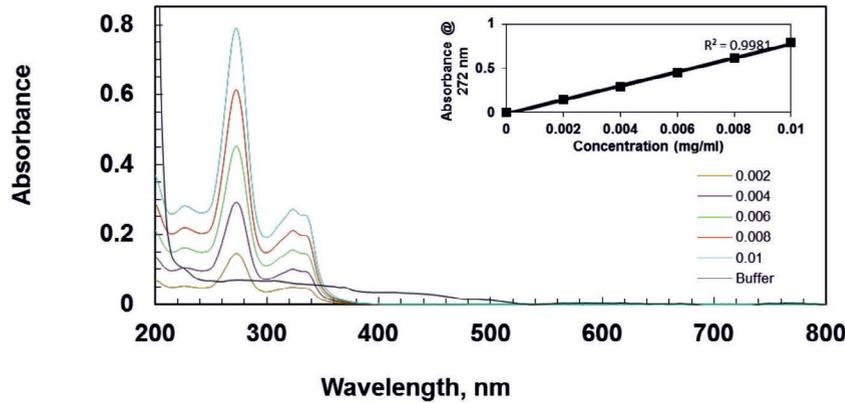


Fig. 3. Spectra of norfloxacin at different concentration.

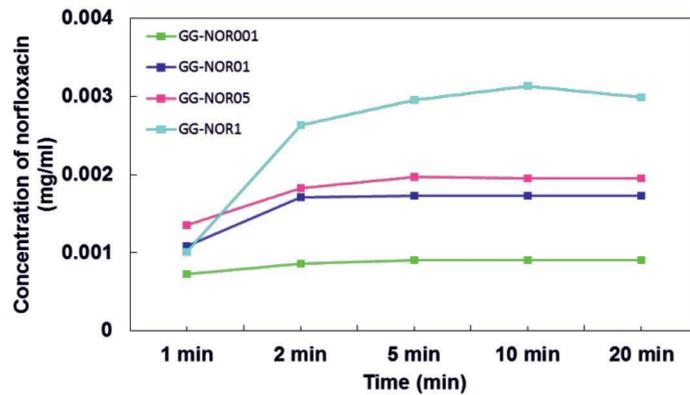


Fig. 4. Graph of releasing property through GG-NOR001, GG-NOR01, GG-NOR05, and GG-NOR1 films.

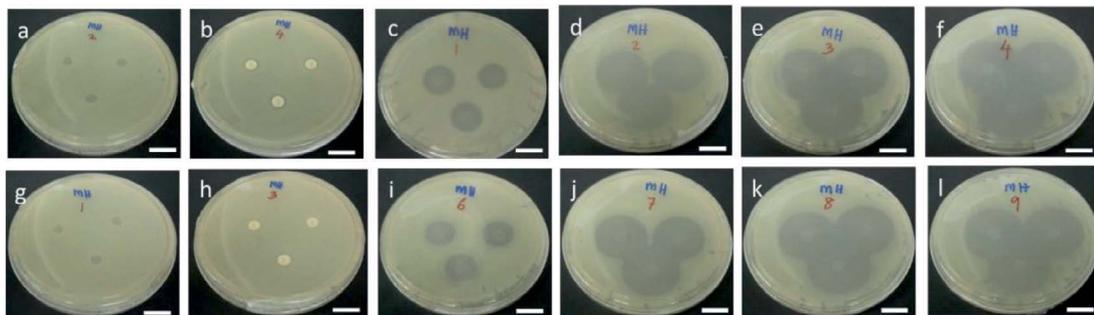


Fig. 5. Disk diffusion results of (a) GG (negative control), (b) penicillin (positive control), (c) GG-NOR001, (d) GG-NOR01, (e) GG-NOR05, and (f) GG-NOR1 films against *Escherichia coli*, respectively, and (g) GG (negative control), (h) penicillin (positive control), (i) GG-NOR001, (j) GG-NOR01, (k) GG-NOR05, and (l) GG-NOR1 films against *Staphylococcus aureus*. Scale bar represents 2 cm.

norfloxacin film at different concentrations. The inclusions of norfloxacin into the gellan gum matrix led to the increased of water absorption in pH 7 and pH 12. The inclusion of 0.01 % (w/w) norfloxacin to the gellan gum increased the swelling percentage in pH 1, pH 7, and pH 12 to 310 %, 1199 %, and 814 % compared to gellan gum films at 308 %, 987 %, and 724 %, respectively. However, increased the concentration of norfloxacin to gellan gum decreased the swelling properties except in pH 7 and pH 12. This is due to the different pH environment of buffer solution. Gellan gum absorbs lesser extent in acidic conditions because of the carboxylic group of L-ascorbic acid becomes protonated to COOH and turns the materials less soluble³⁹. In pH 12, the solubility was increased due to the residue of D-glucuronic acid of carboxylic groups is in the anionic form. The maximum of water uptake in pH 7 of the composite films are due to the highest number of anionic groups in buffer solution. The hydrophilicity properties of the norfloxacin itself also might be one another factor of the increasing of water uptake by the composite films.

For the released study of gellan gum with norfloxacin film, wavelength of 272 nm⁴⁰ was selected for estimation of norfloxacin from the overlain spectra as shown in Fig. 3. The calibration curve was constructed by recording UV-Visible absorbance difference against concentration of norfloxacin.

The drug released was studied by using 0.01, 0.1, 0.5, and 1 % w/w concentration of norfloxacin as active materials. Norfloxacin was incorporated into the gellan gum solution to examine the release properties. The release study was investigated in controlled pH and standard temperature which is at pH7 and 37°C, respectively. Since the composite film will be applied for pharmaceutical, so the neutral pH and human body temperature have been chosen to control the conditions.

Fig. 4 shows the drug release behaviours of GG-NOR composite films in phosphate buffer solution. Based on the results, it shows that the amount of drug release was run out within 5 min for both GG-NOR001 and GG-NOR01, while within 10 and 20 min for GG-NOR05 and GG-NOR1, respectively. It shows that the concentrations of drug incorporated in the film influence the release

rate of norfloxacin. The film incorporated with the lowest norfloxacin concentration shows the fastest release rate while the slowest release rate was shown by the highest incorporated drug. It is noted that the release rate of drug can be controlled with the norfloxacin content.

Apart from that, the released drug also controlled by absorbed water of gellan gum. The anionic side chains that presence in gellan gum molecules was enhanced the hydration activity causing it soluble in Milli-Q-water⁴¹. Gellan gum film will dissolve and forms pores filled with liquid and released the drug through diffusion process⁴¹.

The antibacterial activity of GG-NOR composite films was examined by qualitative (disk method) presented in Fig. 5. The inhibition zone was recorded after 24 h against *Staphylococcus aureus* and *Escherichia coli* executed by norfloxacin.

The incorporated of norfloxacin at high concentration inhibited the bacterial growth at larger diameter. For example, the GG-NOR1 composite films had inhibited 2.5 and 2.9 times larger than that of a GG-NOR001 composite film against *Staphylococcus aureus* and *Escherichia coli*, respectively. The inhibition zone of GG-NOR001 about 5.3 mm and 5.0 mm, while GG-NOR1 inhibited about 13.0 mm and 14.3 mm resisted *Staphylococcus aureus* and *Escherichia coli*, respectively (Table 2).

The GG-NOR composite films were effectively executed Gram-positive and Gram-negative bacterial due to it could inhibit deoxyribonucleic acid synthesis intracellular in bacteria⁴²⁻⁴⁸. It was modified the enzyme structures and caused an abnormal change in production of polypeptide. Since it has different molecular structure compare to the other types of quinolones which has fluorine atom and piperazine ring at six and seven position, respectively, which is obtained from converting nalidixic acid into quinolone structure, it was offered dual effectiveness which is increased potency against gram-negative organisms and also provided anti-pseudomonal activity⁴².

CONCLUSION

The chemical and physical interaction also antibacterial activity of GG and GG-NOR film were examined. Based on the morphological observation, the smooth appearance of cross-

section proved the blends was homogeneously mixed. The swelling properties were increased proportionally in pH 7 to the concentration of norfloxacin. In contrast, the water vapour transmission rates value decreased with increased of norfloxacin added. The release rate of norfloxacin was increased with increase of the concentration norfloxacin incorporated in gellan gum film. Therefore, the inhibition zone was increased at higher concentration of norfloxacin incorporated into film.

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CONFLICTS OF INTEREST

The authors declare that there is no conflict of interest.

AUTHORS' CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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DATA AVAILABILITY

All datasets generated or analyzed during this study are included in the manuscript and/or the Supplementary Files.

ETHICS STATEMENT

This article does not contain any studies with human participants or animals performed by any of the authors.

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